

# Mouse Models of Cancer-associated Thrombosis

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# Outline

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- **Background**
- **Role of tumor-derived TF+ MVs in venous thrombosis in mice with pancreatic tumors**
- **Role of neutrophils and NETs in venous thrombosis in mice with pancreatic tumors**

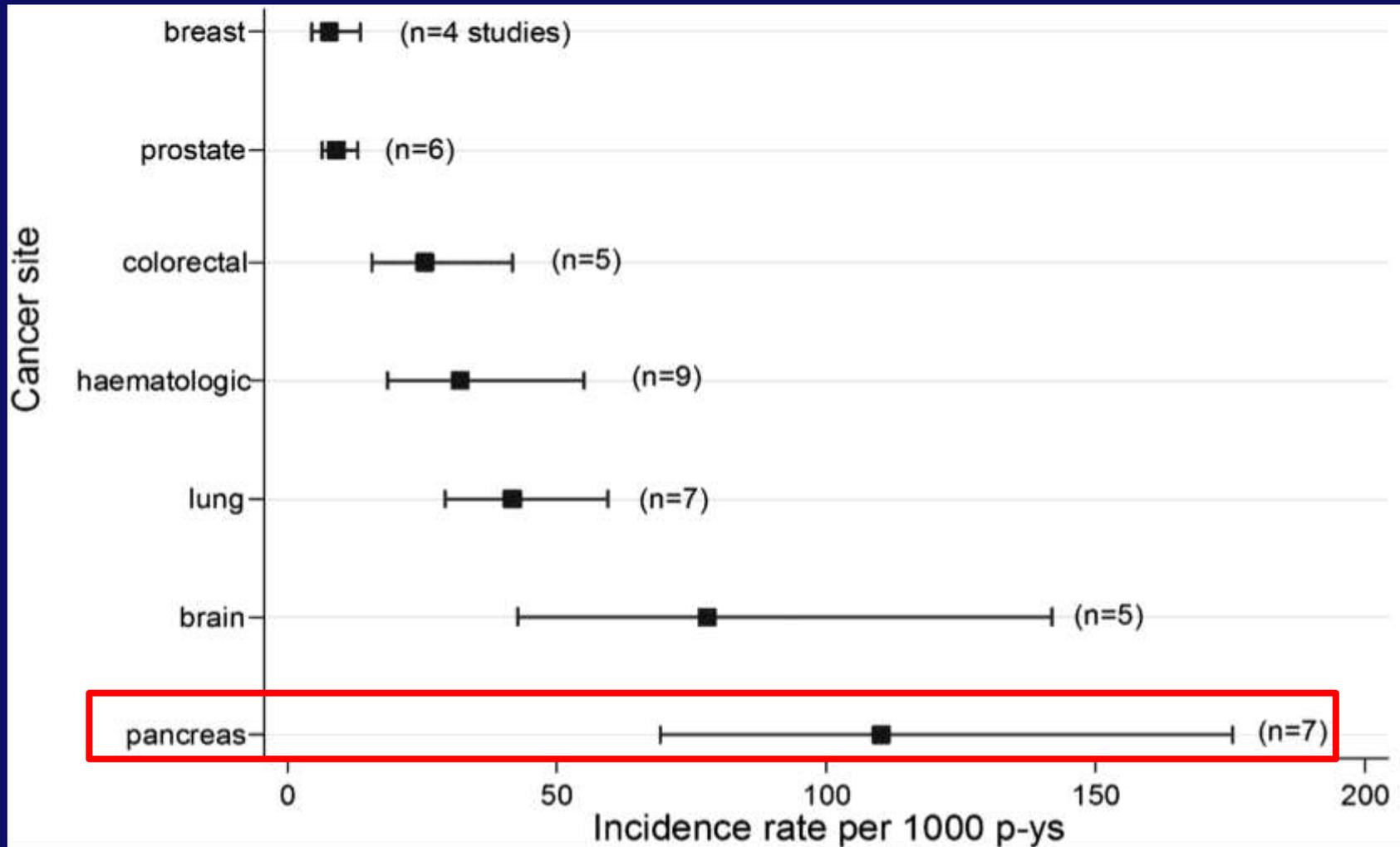
# Cancer and Venous Thrombosis

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**Cancer patients have an a 4-7 fold increased risk for venous thrombosis – cancer-associated thrombosis (CAT)**

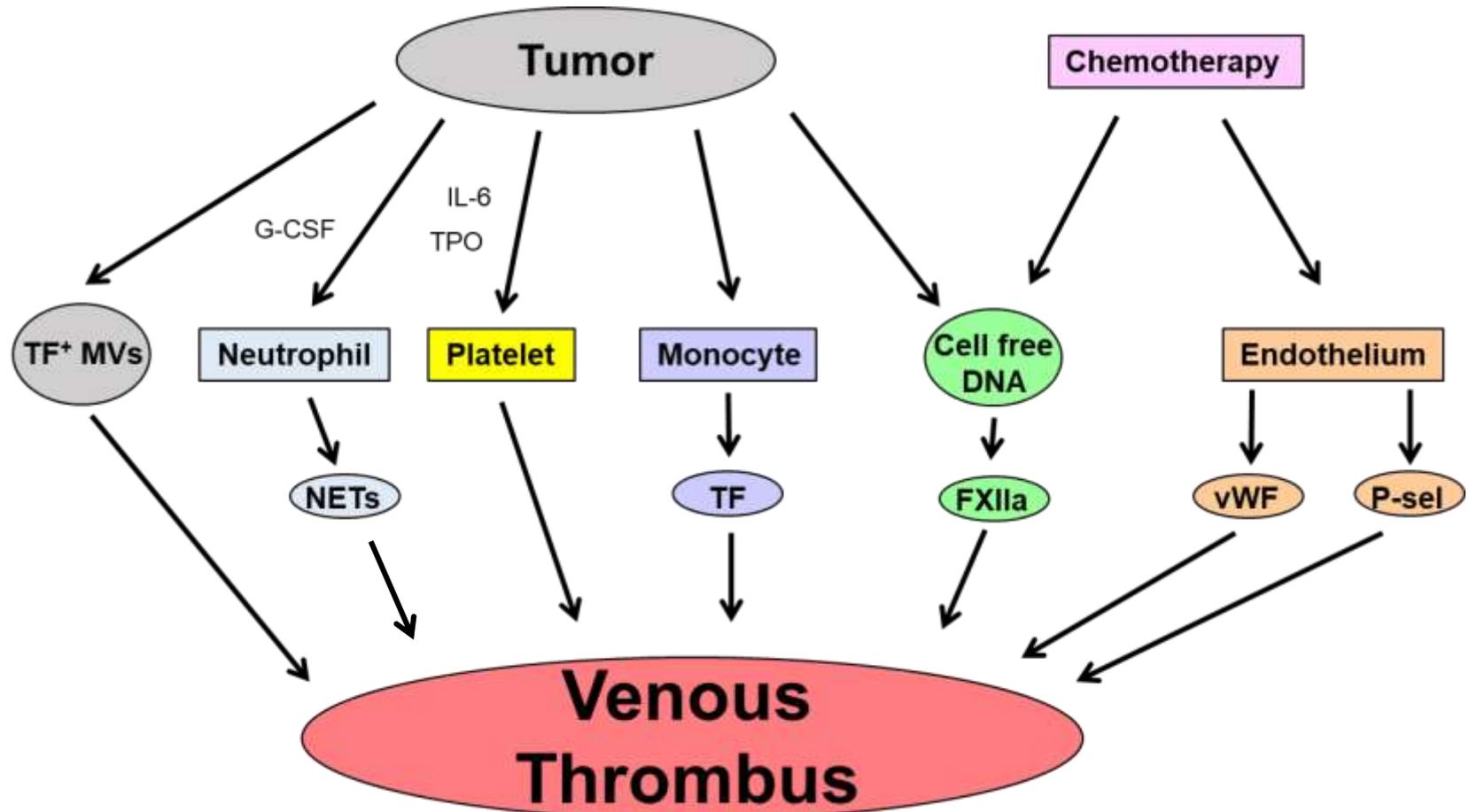
**Blom JW et al. JTH 2006; Khorana AA et al. JTH 2007;  
Timp JF et al. Blood 2013**

# Incidence of Venous Thrombosis in Patients with Different Types of Cancer



Modified from Timp JF et al. Blood 2013

# Possible Pathways of Cancer-associated Thrombosis



**Mouse models are  
important tools to study  
the mechanisms of venous  
thrombosis in cancer  
patients**

# Mouse Models of Cancer-Associated Thrombosis

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- **Mouse strain- immunocompetent vs immunodeficient**
- **Thrombosis model**
- **Site of tumor growth- S.Q. vs orthotopic**
- **Tumor-type**

# Mouse Models of Cancer-associated Thrombosis

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## Immunocompetent mice

Can investigate the contribution of immune cells.  
Can use different KO mice.  
Limited number of murine cancer lines.



## Immunodeficient mice

Can use human cancer cell lines and patient-derived xenografts.  
Easy to image orthotopic tumors expressing reporters.  
Mice lack various immune cells.

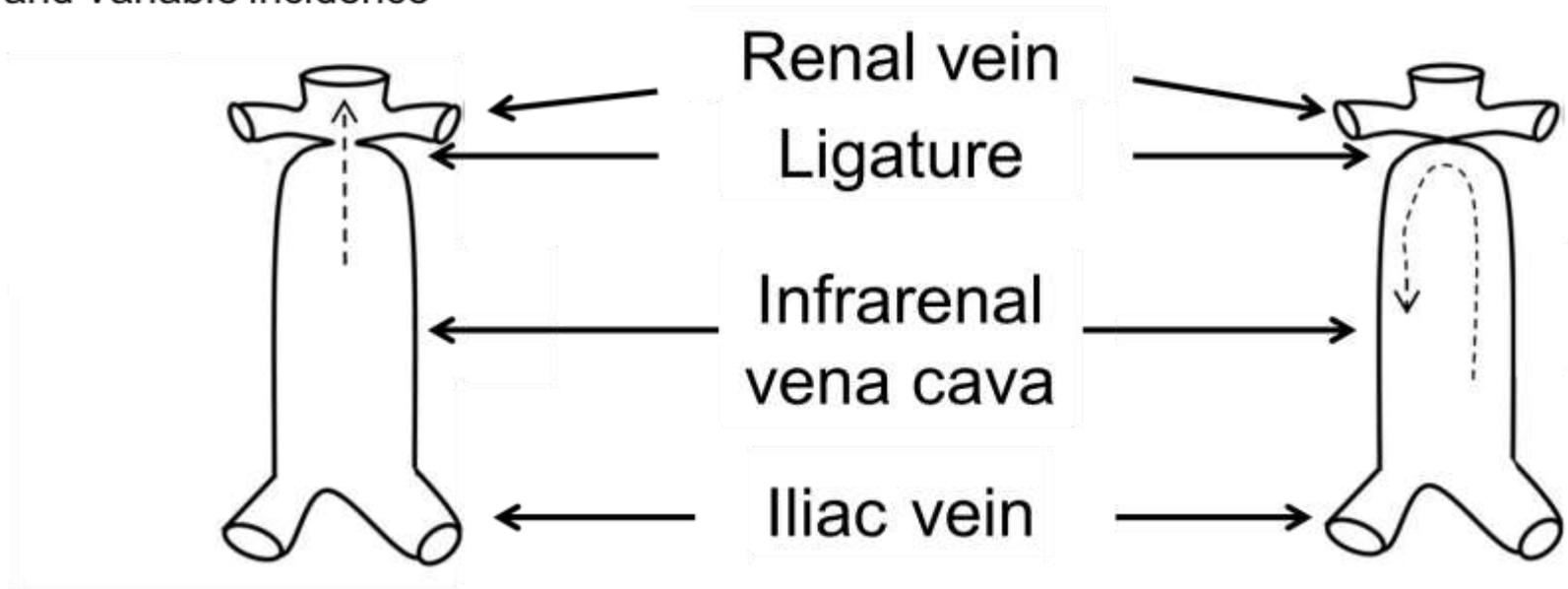
# Mouse Venous Thrombosis Models: Infrarenal Vena Cava Stenosis and Stasis

## Stenosis model:

1. Maintains blood flow
2. Small thrombus (~5mg) formation and variable incidence

## Stasis model:

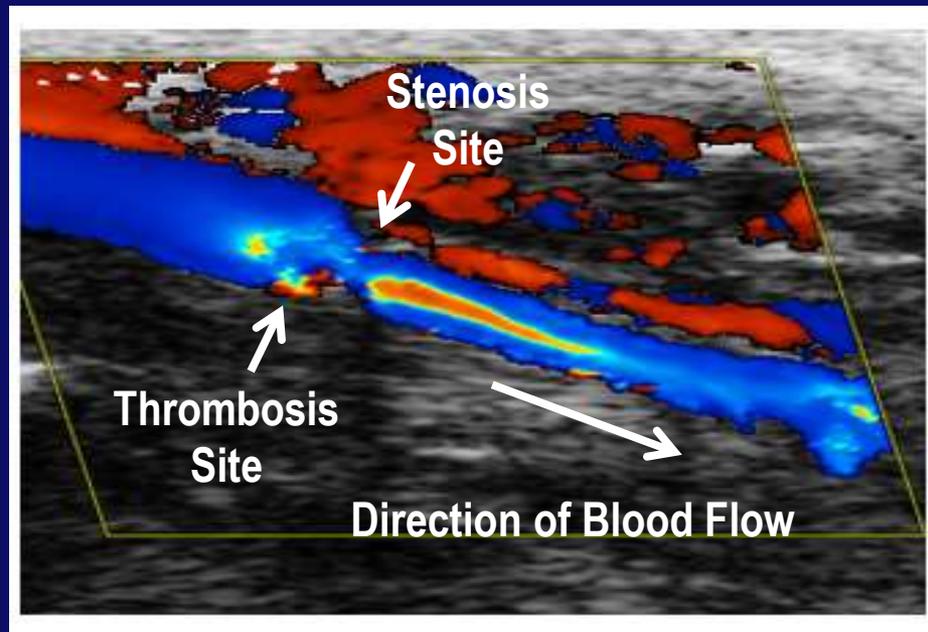
1. No blood flow
2. Large thrombus (~20mg) with ~100% incidence



# Kinetic Analysis of Thrombosis Formation

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## Color Doppler



Geddings J et al. JTH 2014

# Summary of Allograft Mouse Models of Cancer-associated Thrombosis

Mouse strain	Cell line	Tumor site	Thrombosis model	Result	Ref
C57BL/6	LLC lung	S.Q.	FeCl <sub>3</sub> /mesenteric	Occlusion time ↓	Thomas et al JEM 2009
C57BL/6	M27 lung	Orthotopic	FeCl <sub>3</sub> /IVC	Clot weight ↑	Aghourian et al Blood 2016
BALB/c	4T1 breast	Orthotopic	Rose Bengal/Jagular	Occlusion time ↓	Leal et al Sci Rep 2016
C57BL/6	Pan02 pancreatic	S.Q.	FeCl <sub>3</sub> /mesenteric	Occlusion time ↓	Thomas et al JEM 2009
C57BL/6	Pan02 pancreatic	S.Q.	Laser/cremaster	Clot ↑	Mesouar et al Int J Cancer 2015
C57BL/6	Pan02 pancreatic	S.Q.	IVC/stenosis	Clot incidence & weight ↑	Thomas et al JTH 2015

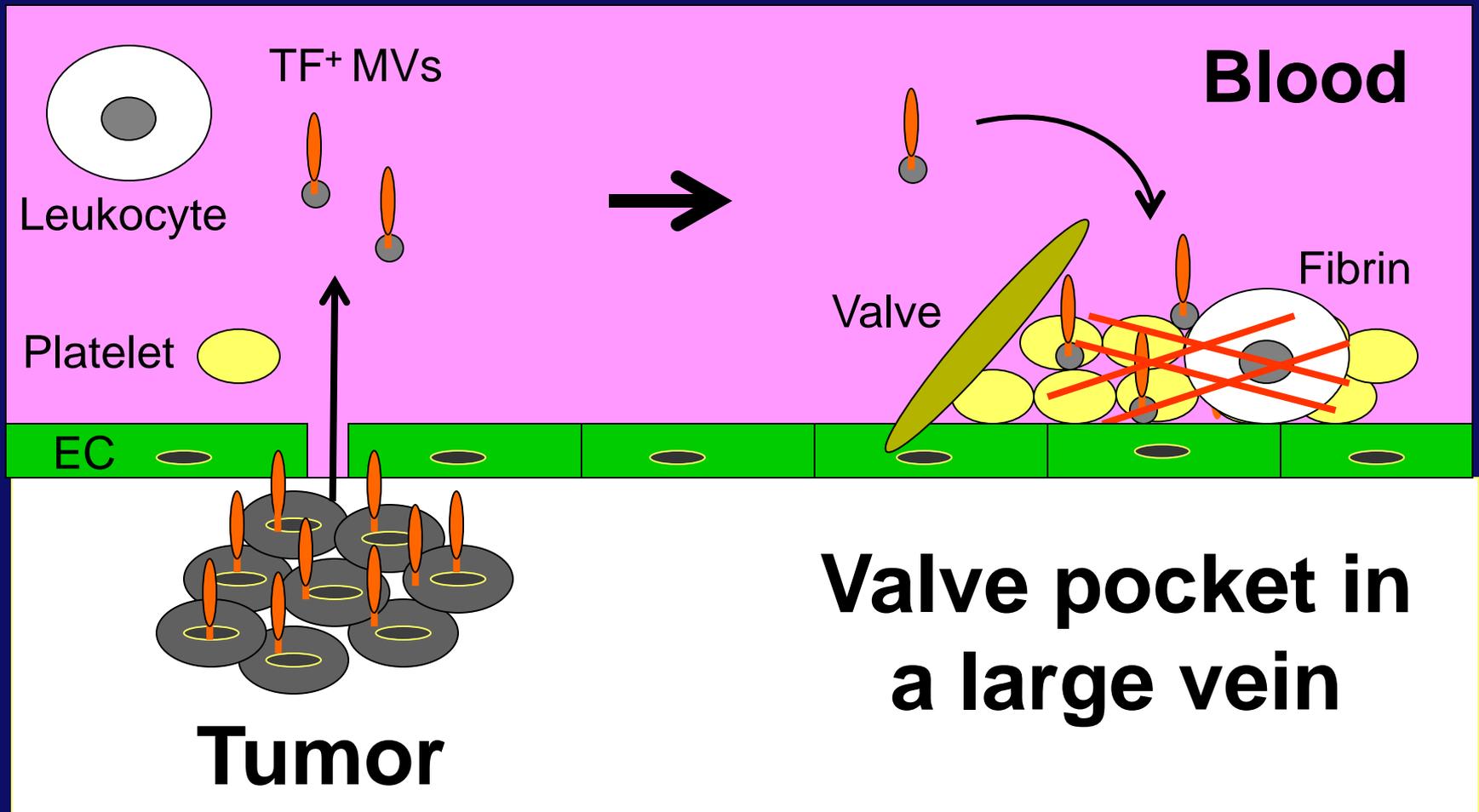
Hisada Y and Mackman Thromb Res 2018

# Summary of Xenograft Mouse Models of Cancer-associated Thrombosis

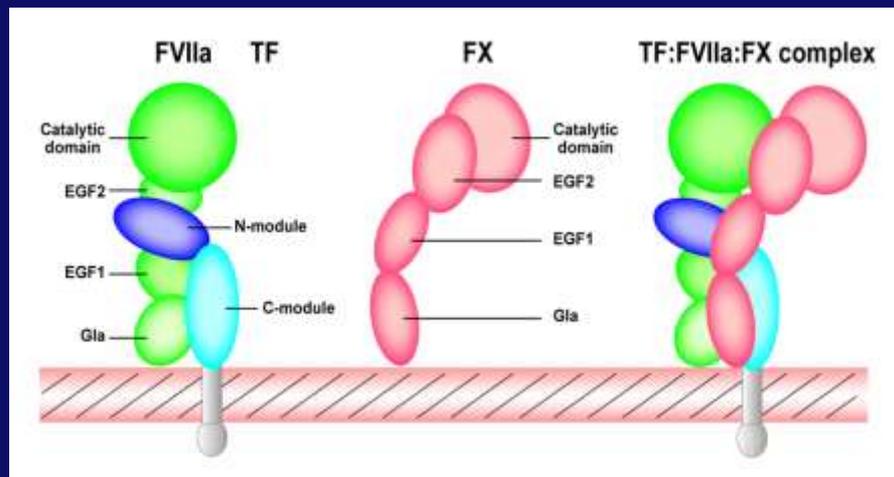
Cell line	Thrombosis model	Result	Ref
A549 lung	IVC stenosis (3h)	Clot weight ↑	Chen et al Mol Cancer 2015
HPAF-II pancreatic	IVC stenosis (3h)	No change	Wang et al Blood 2009
BxPc-3 pancreatic	IVC stenosis (0-24h)	Clot incidence ↑	Geddings et al JTH 2015
BxPc-3 pancreatic	IVC stasis (0-48h)	Clot weight ↑	Hisada et al JTH 2017

Hisada Y and Mackman Thromb Res 2018

# Hypothesis: Tumor-derived TF<sup>+</sup> MVs Trigger Venous Thrombosis in Cancer Patients



# Tissue Factor (TF): The Primary Activator of Blood Coagulation

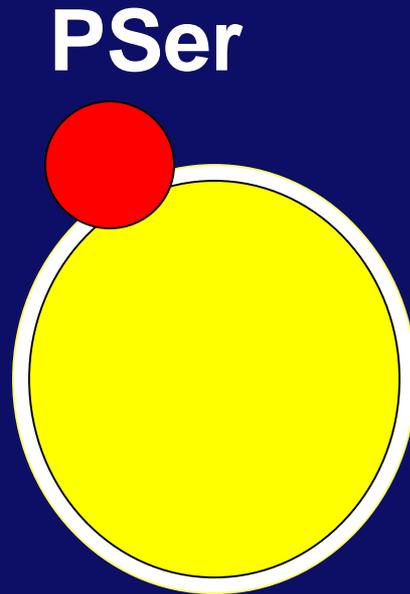


- 47 kDa transmembrane glycoprotein
- Receptor for FVII/FVIIa
- The TF:FVIIa complex activates both FIX and FX

# Types of Procoagulant MVs

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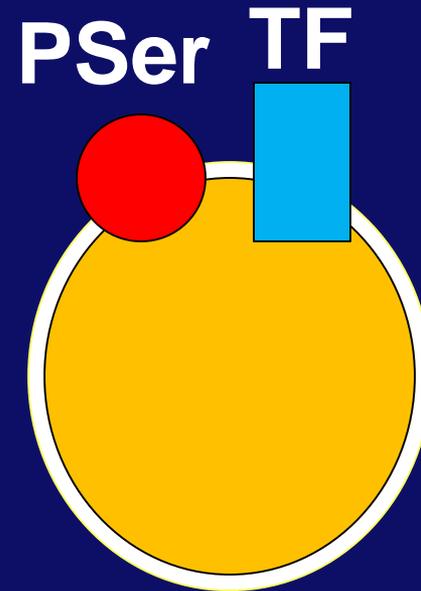
Microvesicles (MVs) are defined as small (0.1-1  $\mu\text{m}$ ) membrane vesicles that are released from activated or apoptotic cells



Procoagulant  
activity  
Origin

**++**

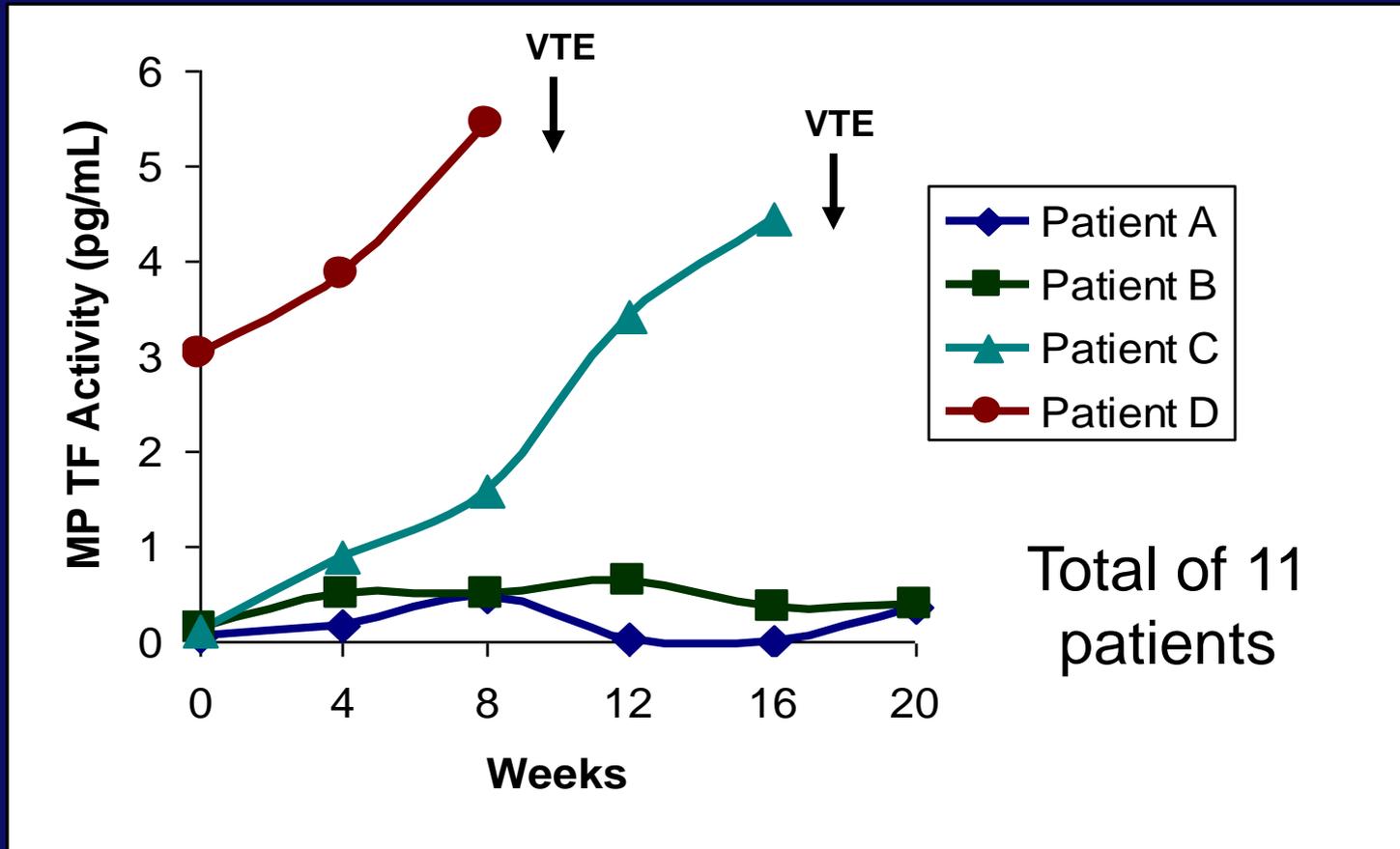
**Platelet/  
RBC**



**+++**

**Monocyte/  
tumor cell**

# Levels of MV TF Activity Increase in Pancreatic Cancer Patients before VTE



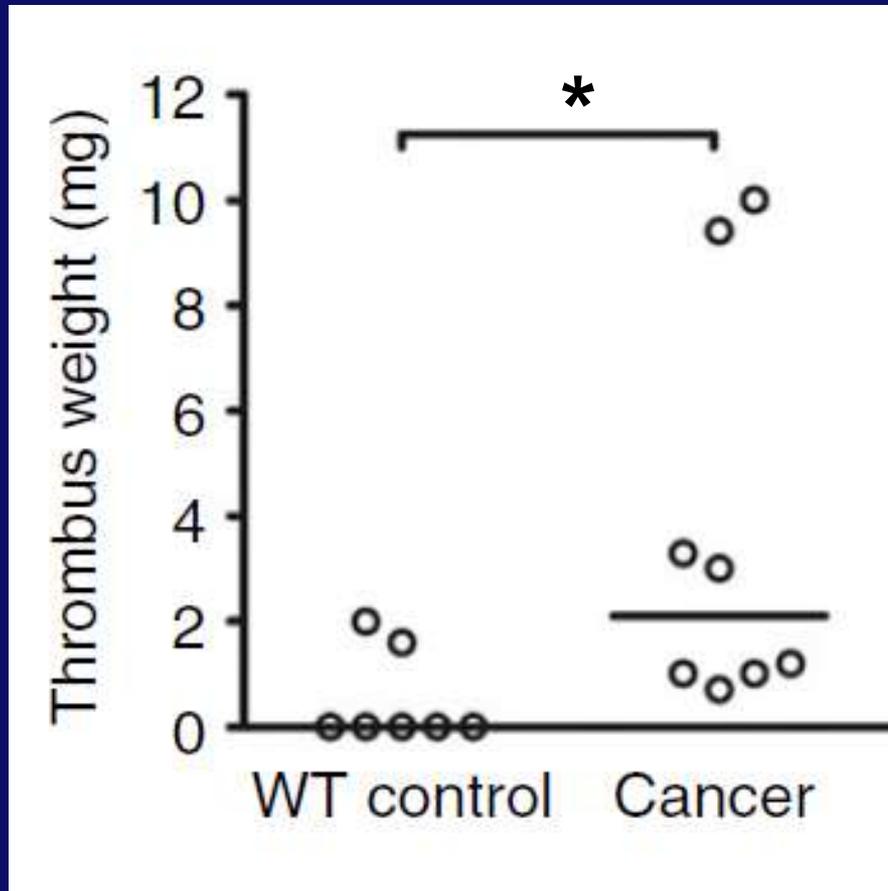
Khorana A et al JTH 2008

# Association of MV TF Activity and VTE in Pancreatic Cancer Patients

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Study	Patients	Time of Follow-up	Association between MV TF and VTE?
Khorana, 2008	2/9 patients developed VTE	5 months	Yes
Van Doormaal, 2012	3/13 patients developed VTE	6 months	Yes
Thaler, 2012	12/60 patients developed VTE	2 years	Weak
Bharthuar, 2013	*52/117 patients developed VTE	6 months	Yes
Van Es, Resubmitted	9/89 patients developed VTE	6 months	Weak
Khorana, In revision	7/39 patients developed VTE	3 months	Yes
Ilich, In preparation	4/22 patients developed VTE	6 months	Yes

# Increased Thrombosis in C57BL/6 Mice bearing Murine Pancreatic Tumors

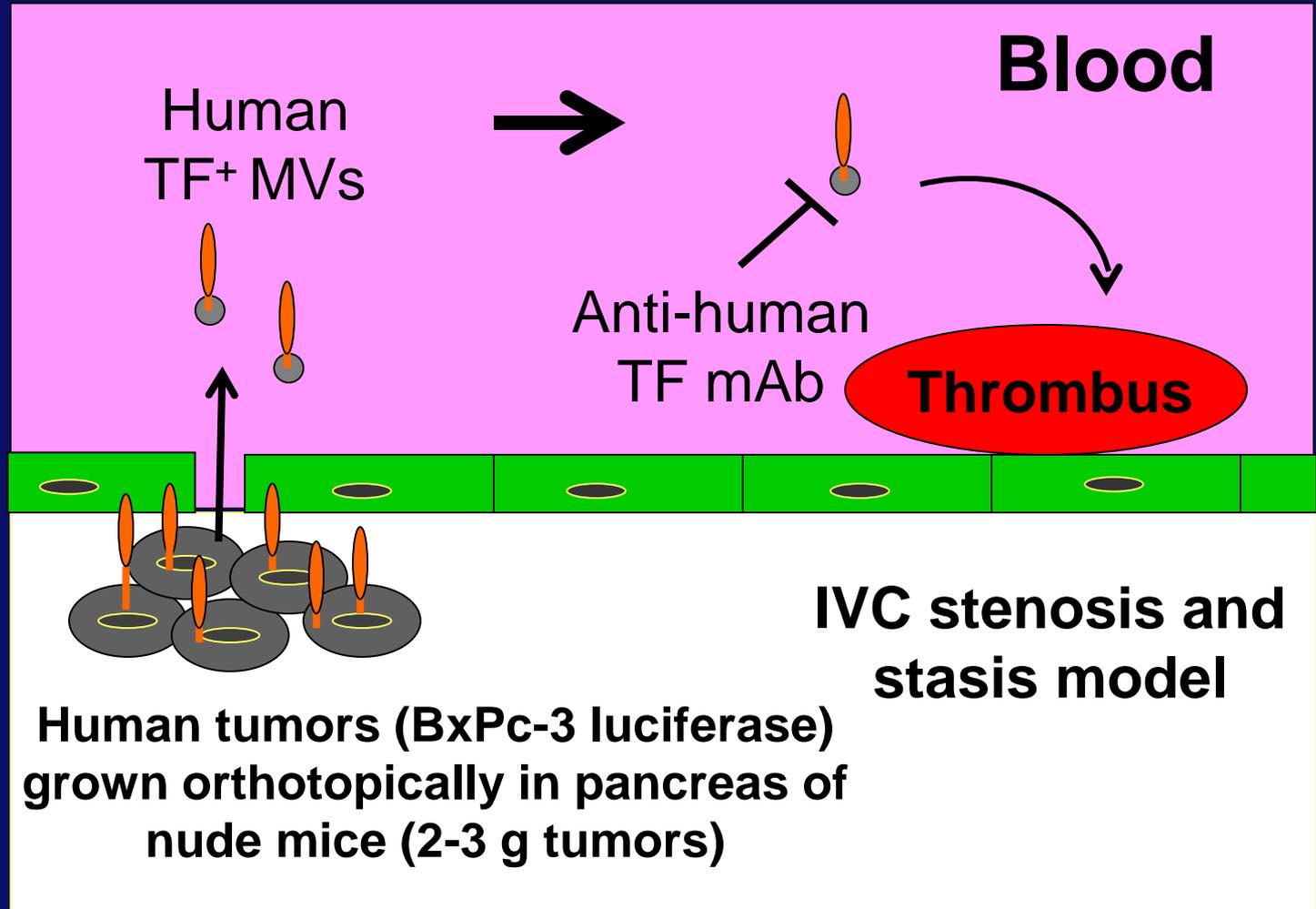


Clot measured at  
3 h in the IVC  
stenosis model in  
mice bearing  
Panc02 tumors

# Model to Study the Role of Tumor-derived, TF<sup>+</sup> MVs in Venous Thrombosis in Mice

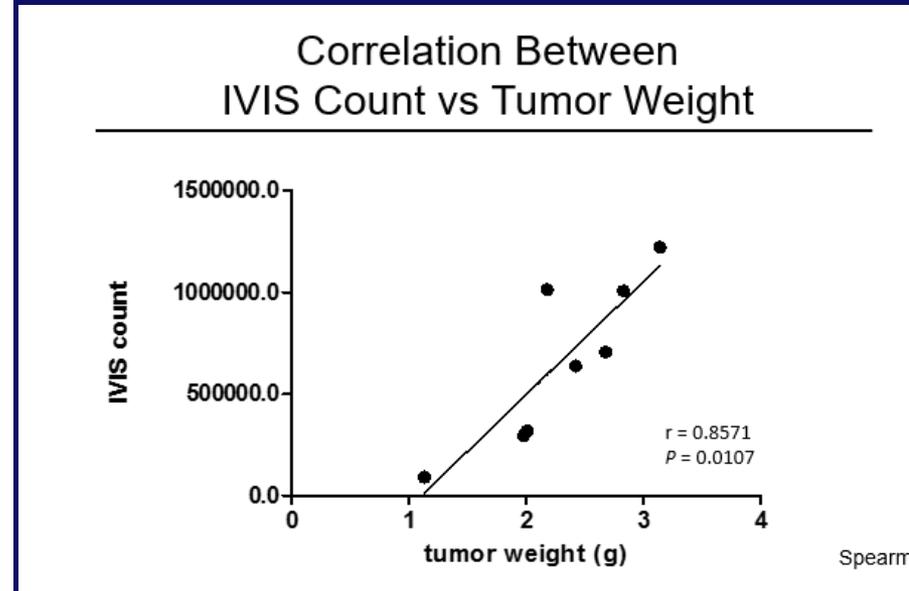
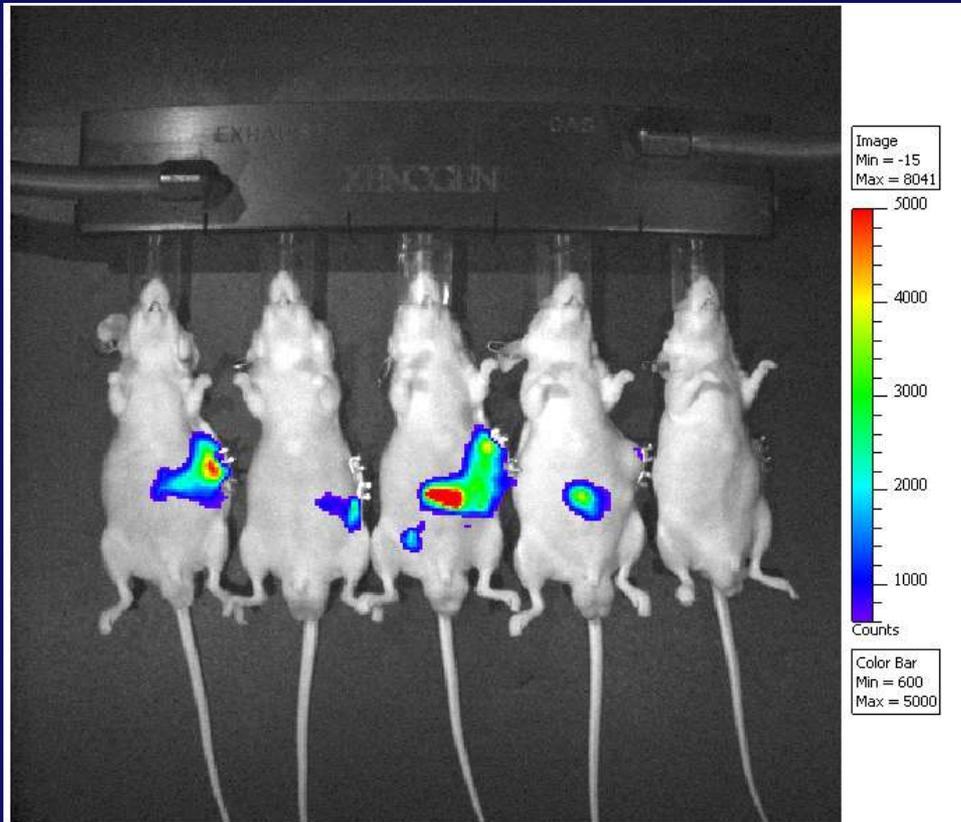


Julia Geddings

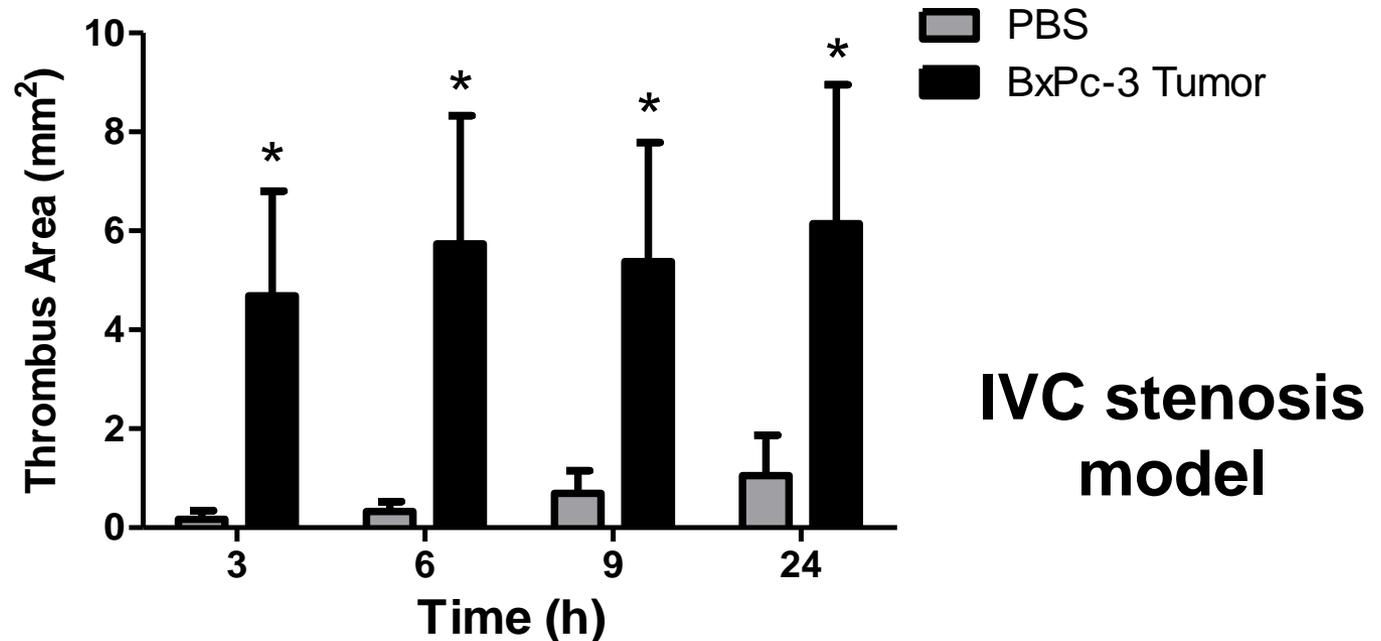


Yohei Hisada

# Imaging of BxPc-3-luciferase Tumors in the Pancreas of Mice



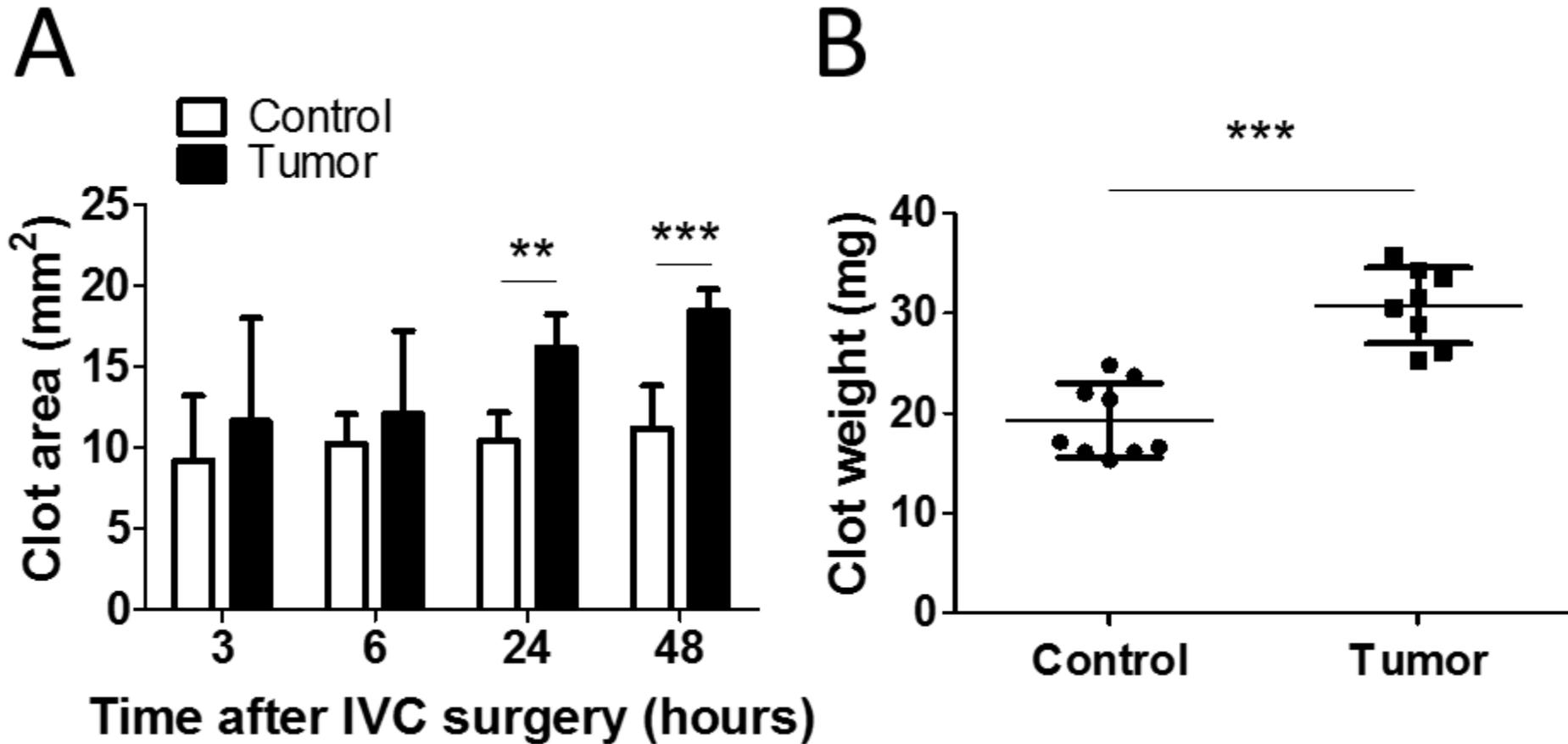
# Increased Thrombosis in Nude Mice bearing Human Pancreatic Tumors



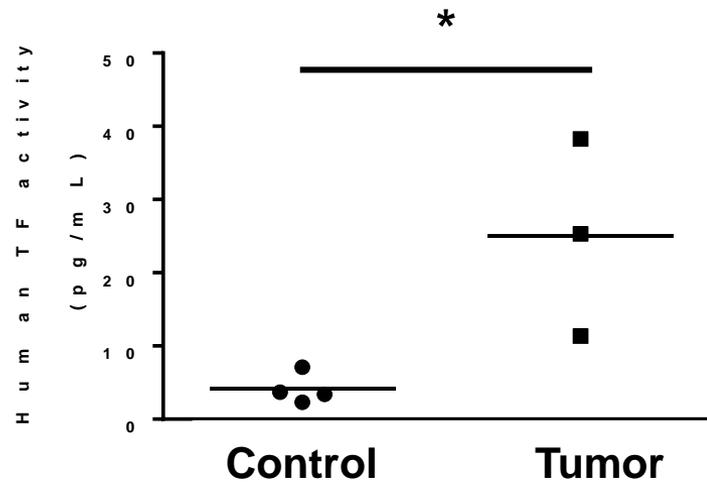
Thrombus Incidence

Time (h)	3	6	9	24
PBS	17%	33%	33%	33%
BxPc-3 Tumor	50%	50%	50%	50%

# Human Pancreatic Tumors increase Clot Size in Nude Mice

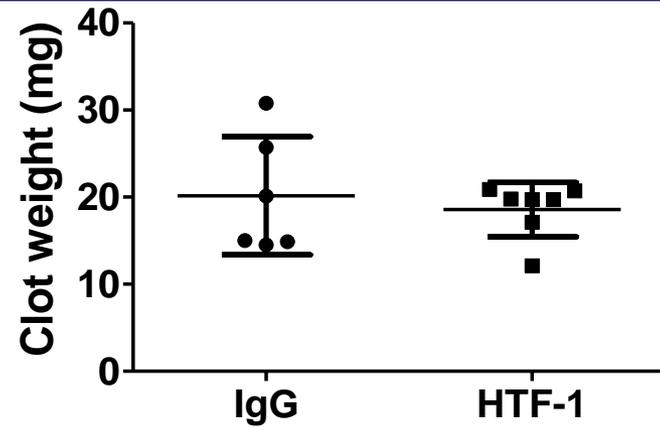
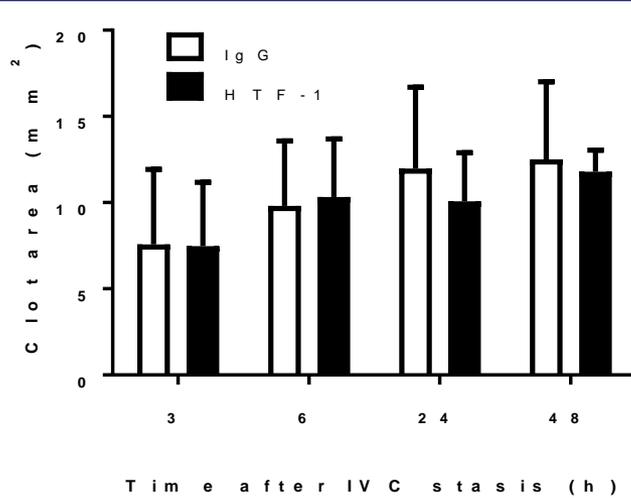


# Clots from Tumor-bearing Mice Contain Human TF



# Inhibition of Human TF Reduces Clot Size in Tumor Bearing Mice

Control



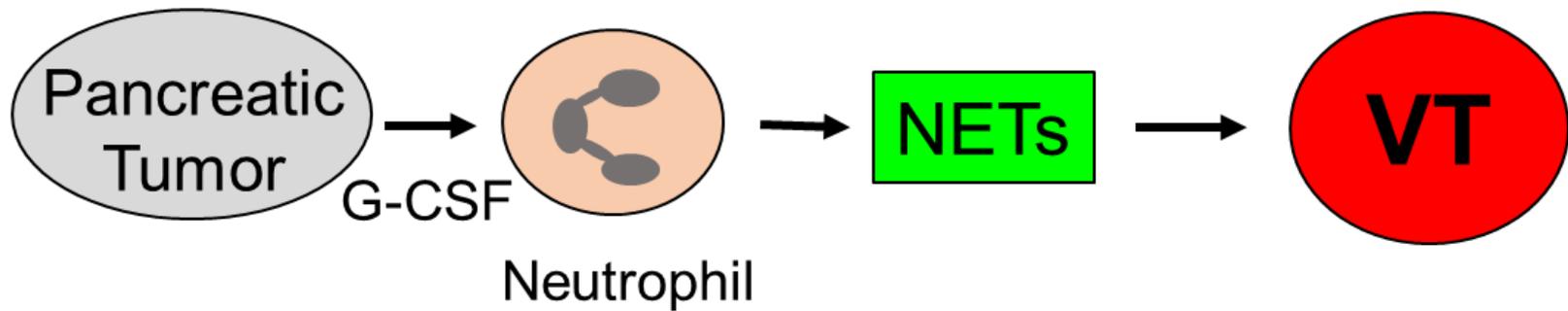
# Conclusion

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**Tumor-derived TF<sup>+</sup> MVs  
enhance venous  
thrombosis in mice with  
pancreatic tumors**

# Hypothesis: Neutrophils and NETs Contribute Venous Thrombosis in Cancer Patients

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# Neutrophil Extracellular Traps- NETs

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**NETs are extracellular DNA fibers comprising of histones and neutrophil antimicrobial proteins- formed by a cell-death program called NETosis**

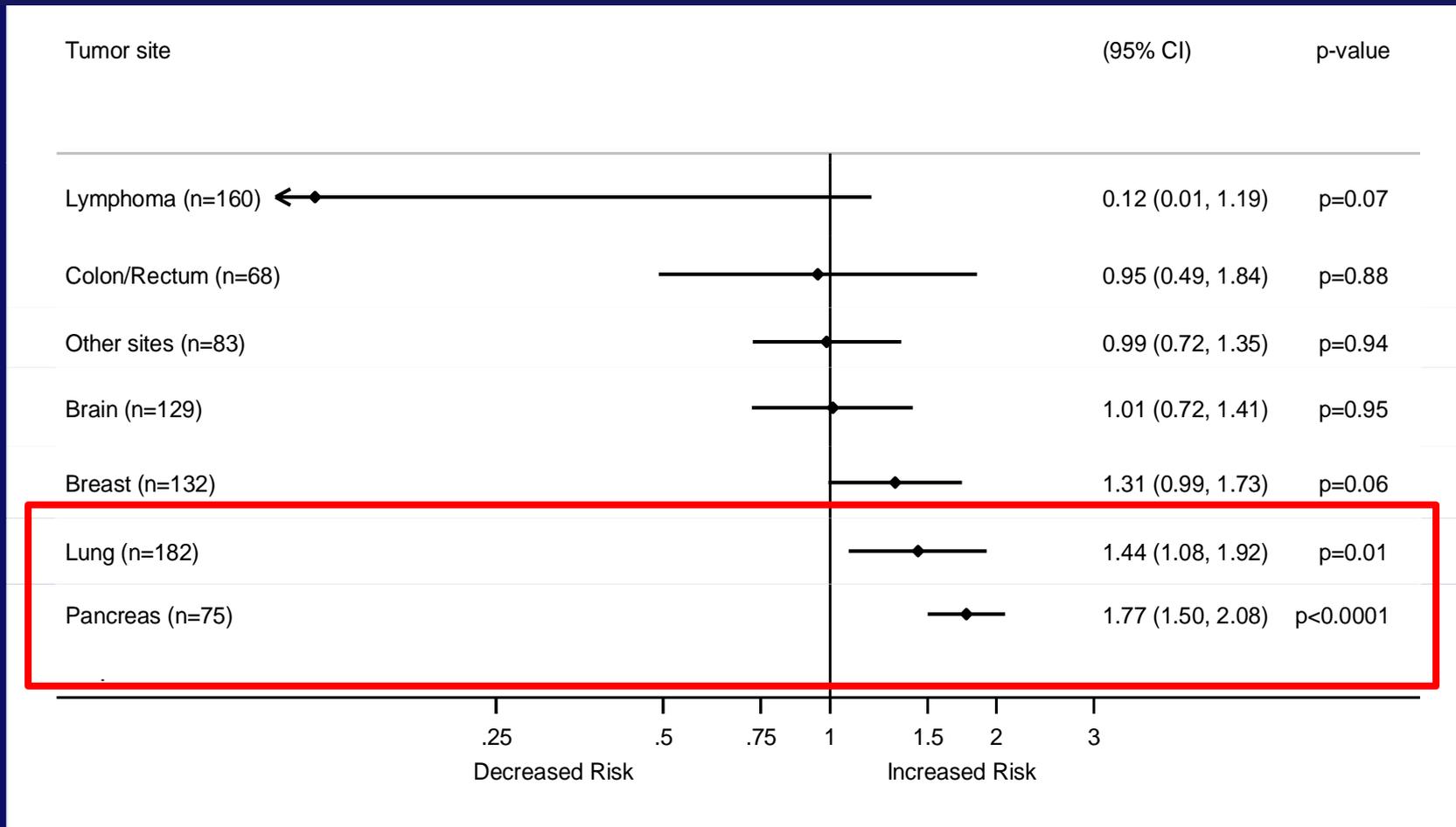
Brinkmann et al Science 2004 Neutrophils  
extracellular traps kill bacteria

# NETs and Thrombosis

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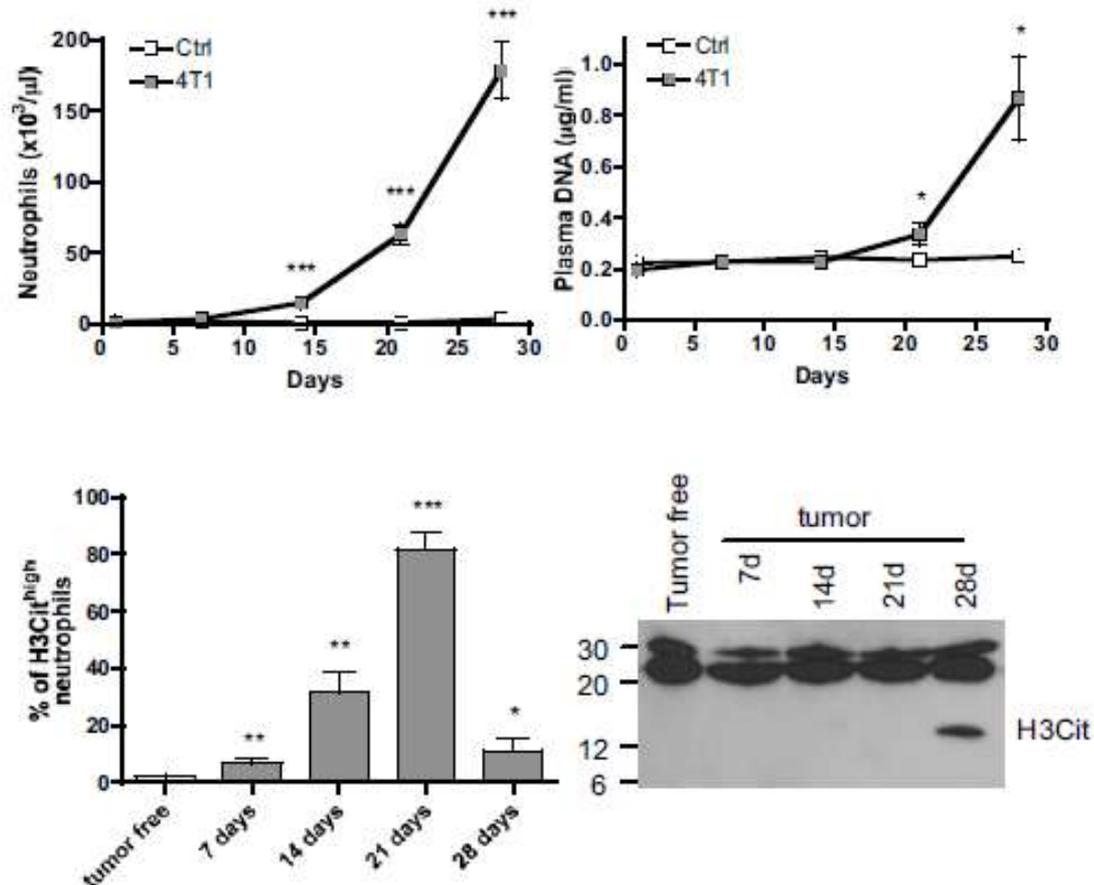
- **NETs are found in venous and arterial clots.** Fuchs et al PNAS 2010; Longstaff et al JBC 2013; Mangold et al Circ Res 2015.
- **NETs contribute to venous thrombosis in mouse models.** Brill et al JTH 2012; Martinod et al PNAS 2013.

# Levels of H3Cit are Associated with VTE Lung and Pancreatic Cancer

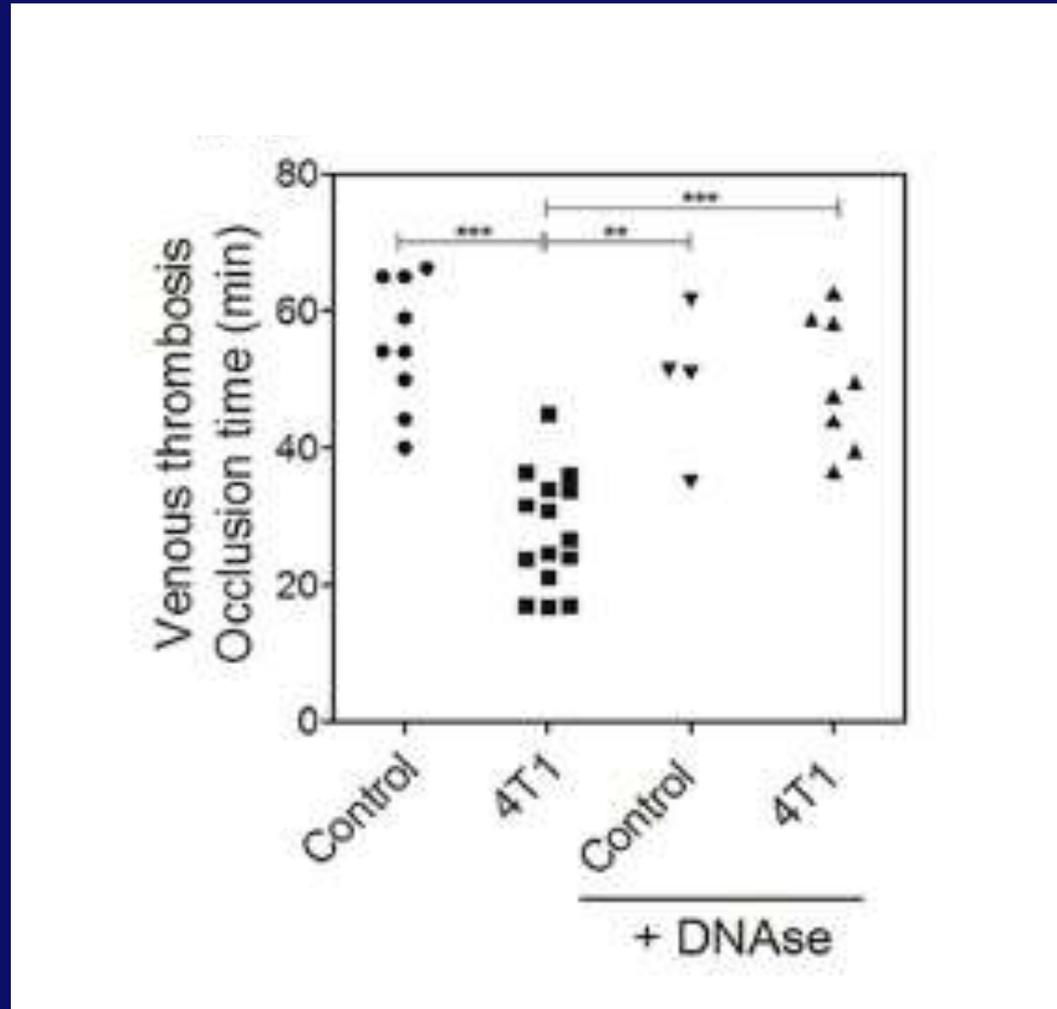


Modified from Mauracher LM et al. JTH 2018

# Mice bearing 4T1 Breast Tumors have increased Neutrophils and NETs

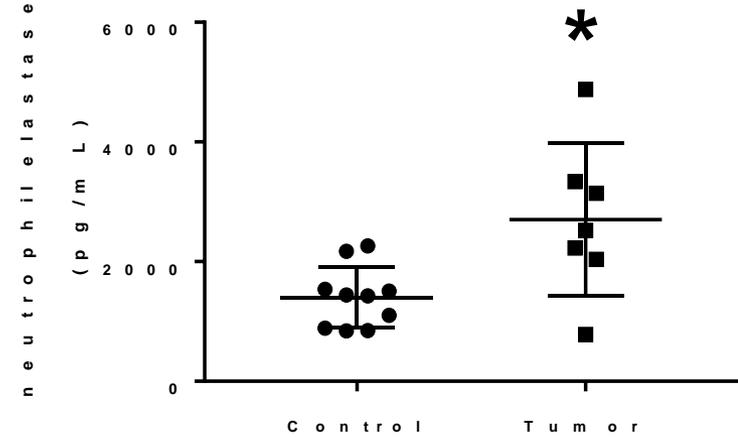
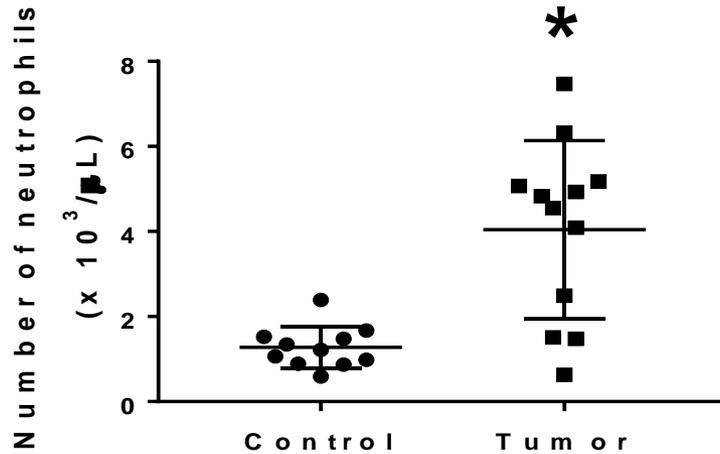


# Mice bearing 4T1 Breast Tumors have increased Venous Thrombosis

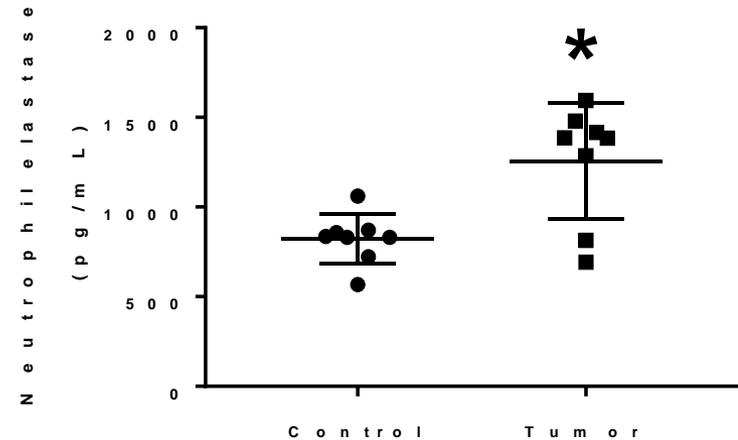
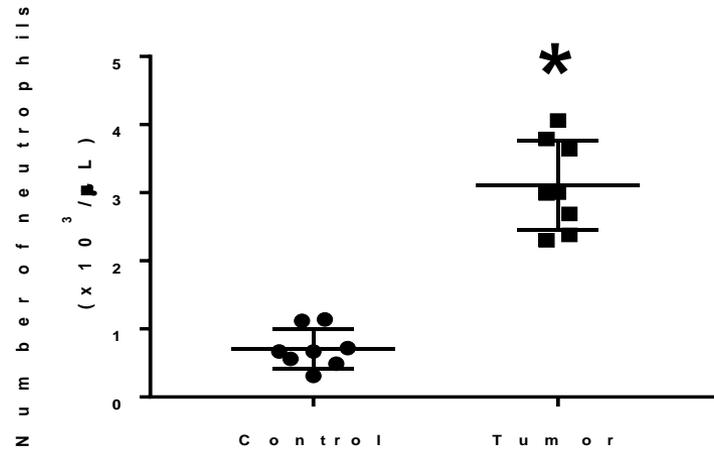


# Mice bearing Human and Murine Pancreatic Tumors have increased Neutrophils and Neutrophil Elastase

**BxPc-3  
(Human)**

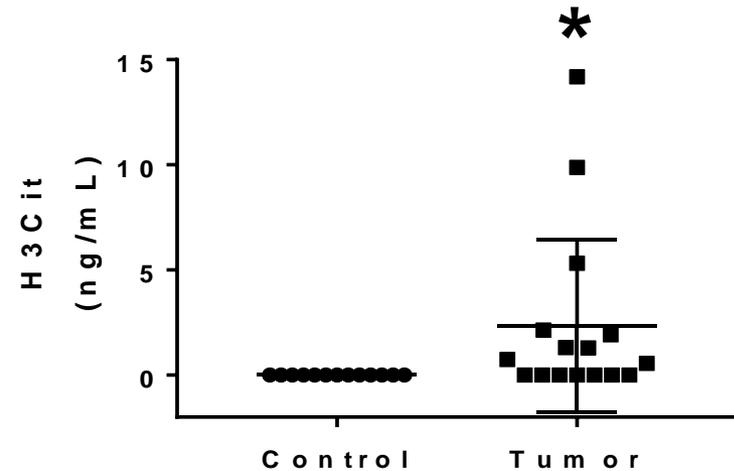
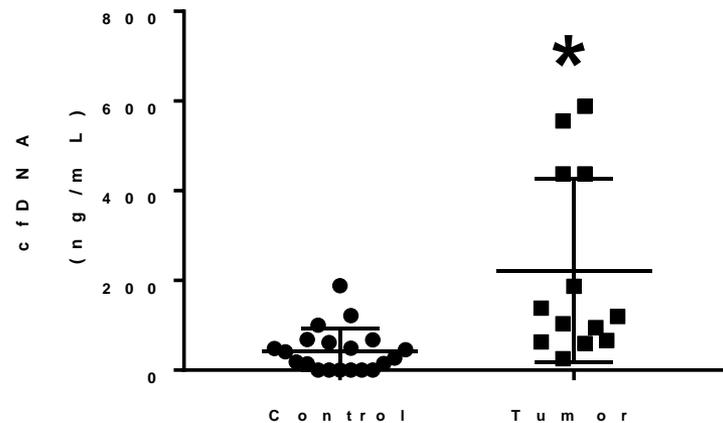


**Pan02  
(Mouse)**



Hisada unpublished data

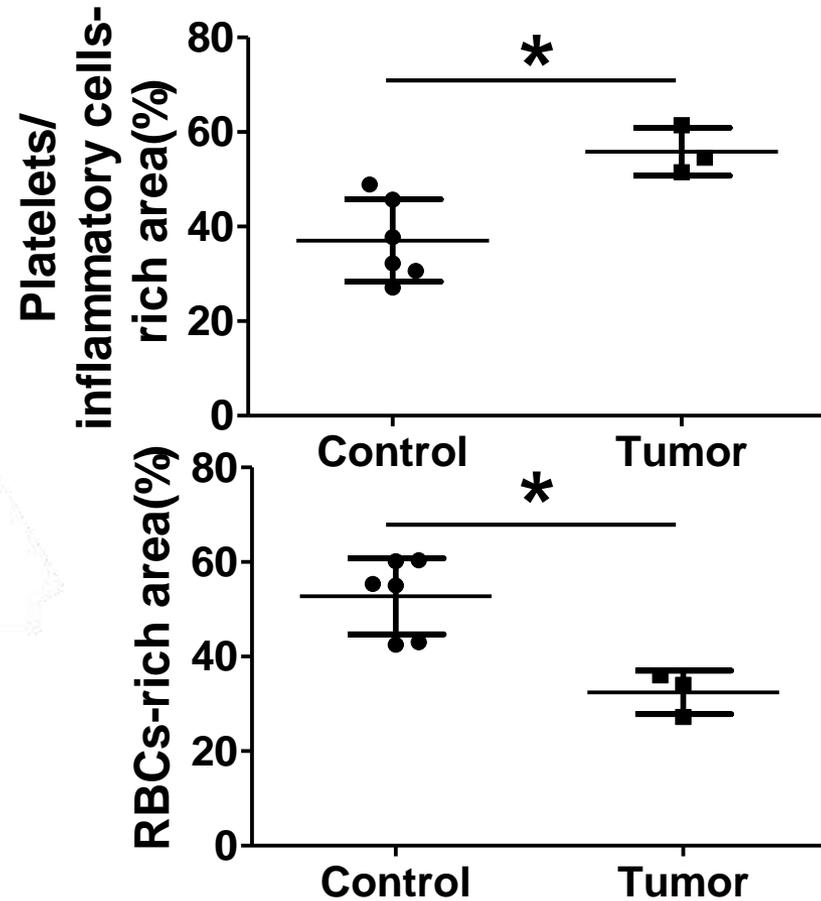
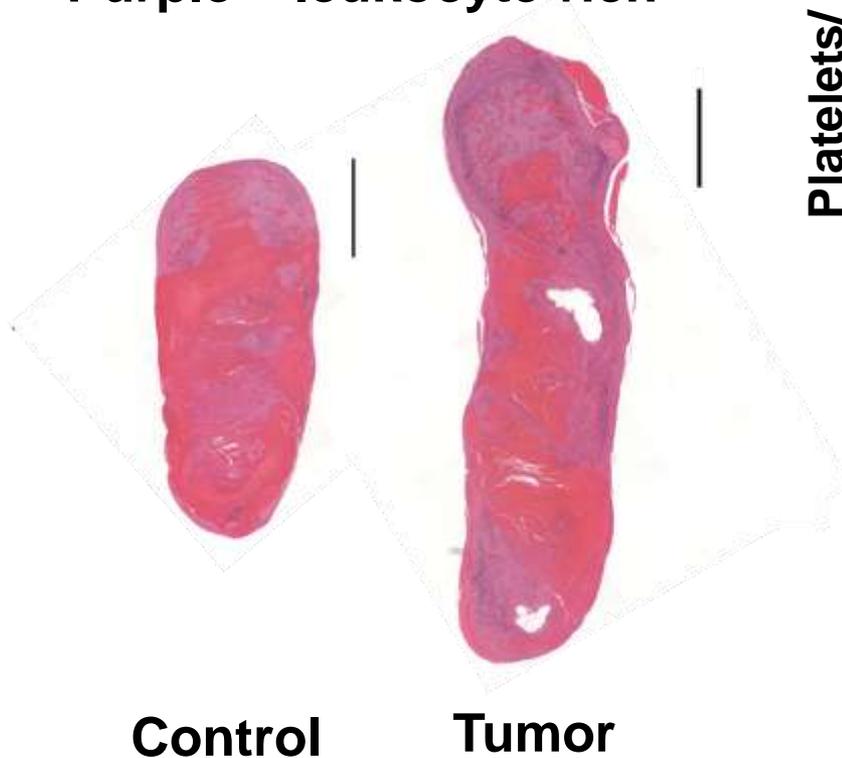
# Mice bearing Human Pancreatic Tumors have increased Plasma cfDNA and NETs



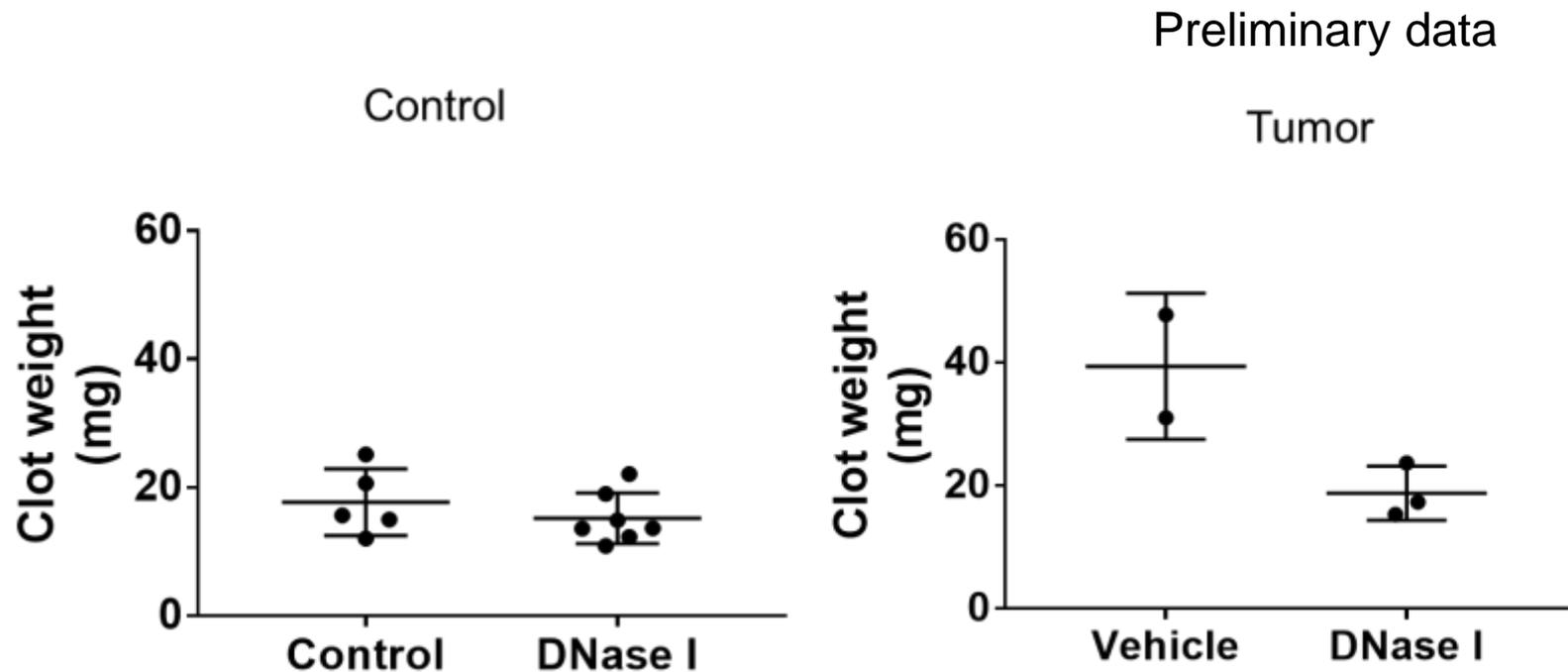
Hisada unpublished data

# Clots from Mice bearing BxPc-3 Tumors have increased Leukocytes

Red = RBC-rich  
Purple = leukocyte-rich



# Effect of DNase I on Thrombus Size in an IVC Stasis Model in Control and Tumor bearing Mice



Hisada unpublished data

# Conclusion

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**Neutrophils and NETs  
appear to contribute to  
venous thrombosis in mice  
with pancreatic tumors**

# Acknowledgements

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## **Mackman Lab**

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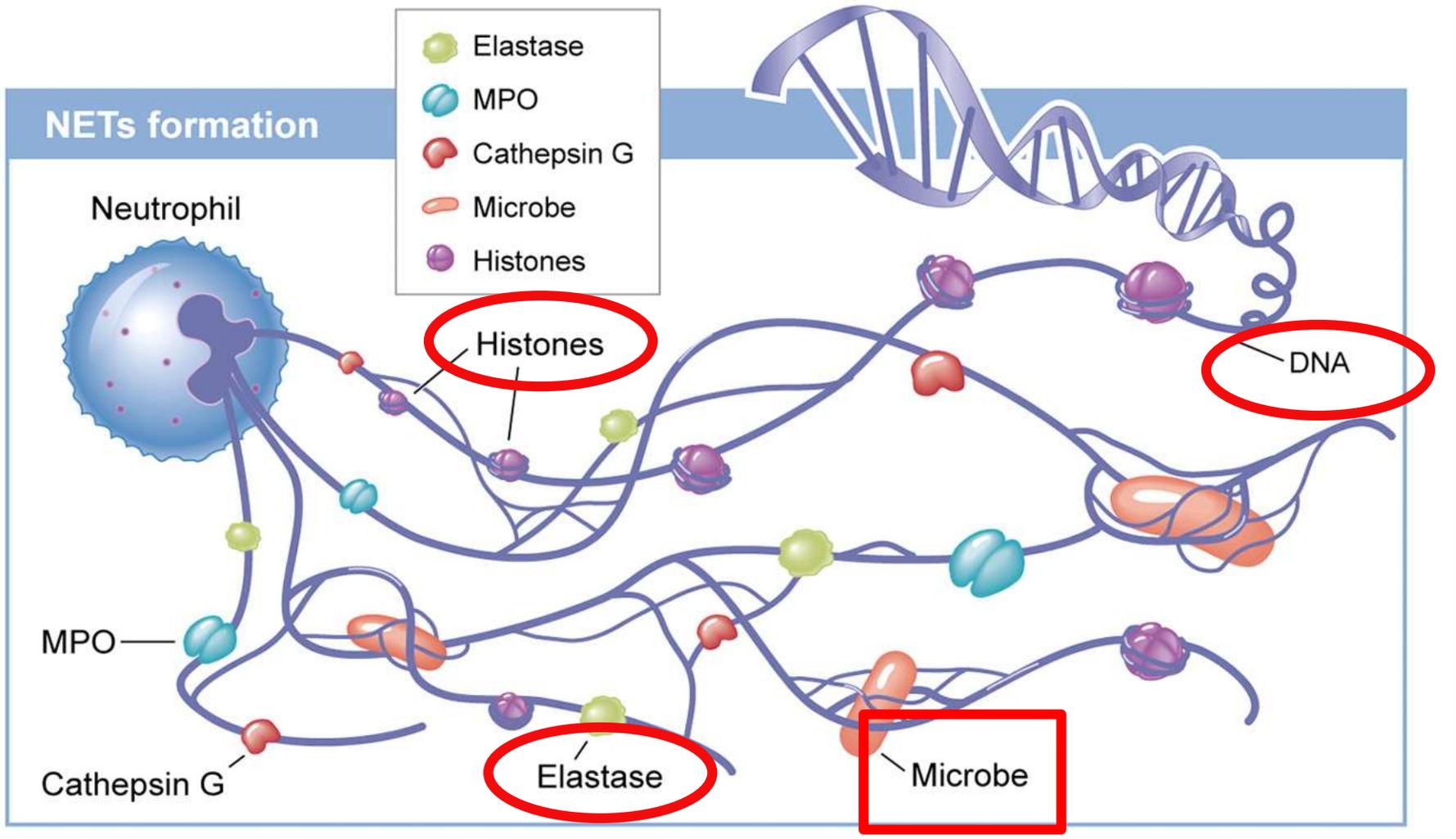


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# Neutrophil Extracellular Traps (NETs)

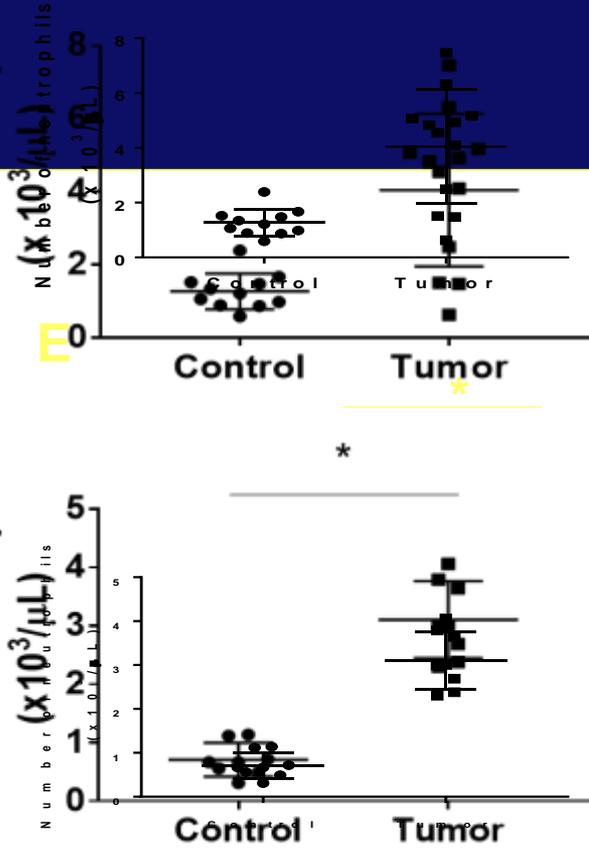


# Conclusion

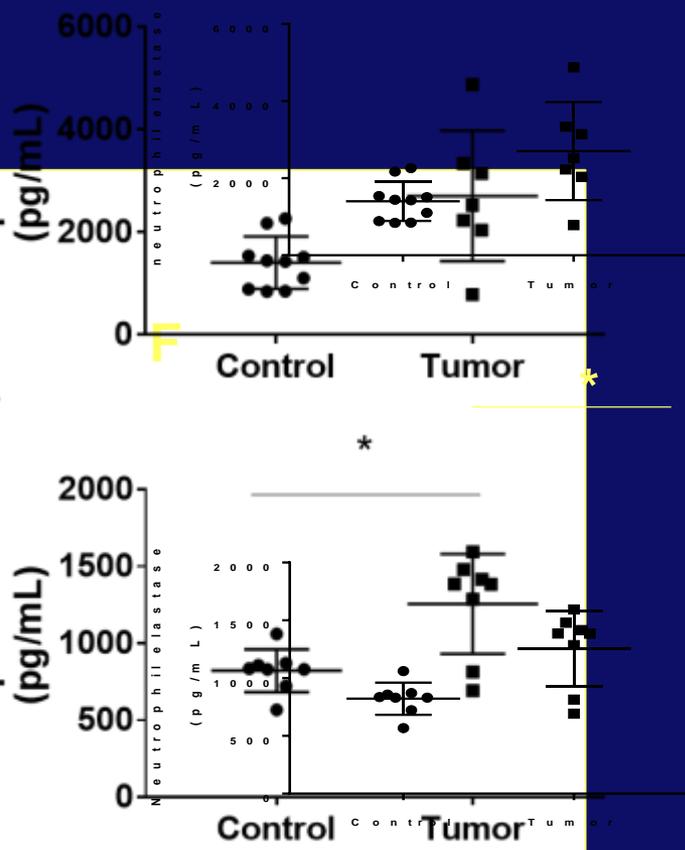
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**Tumor-derived TF<sup>+</sup> MVs  
enhance venous  
thrombosis in mice with  
pancreatic tumors**

Number of neutrophils  $\square$  Number of neutrophils  $\square$



Neutrophil elastase  $\square$  Neutrophil elastase  $\square$



**B**

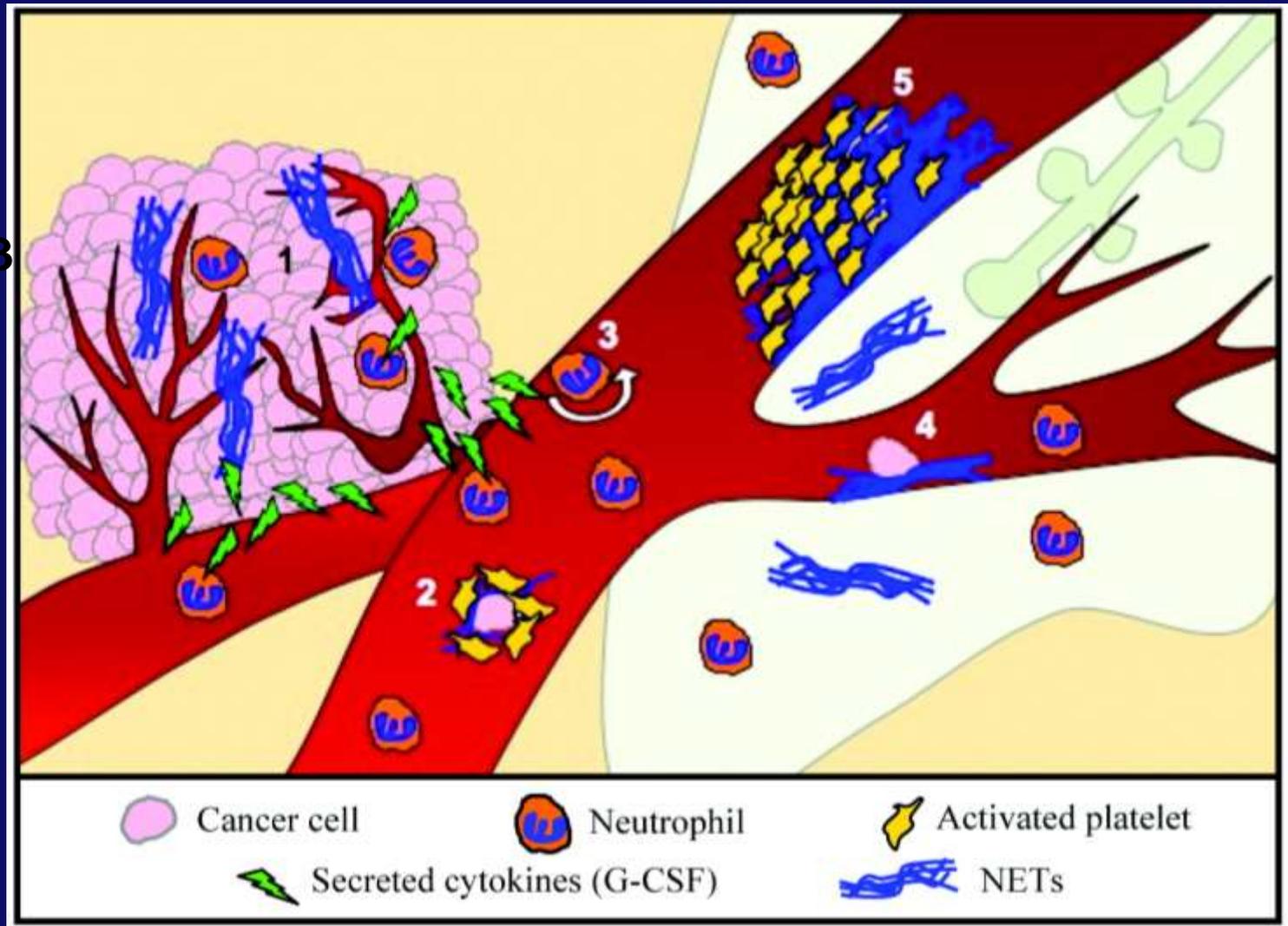
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**C**

\*

# NETs and Cancer-associated Thrombosis

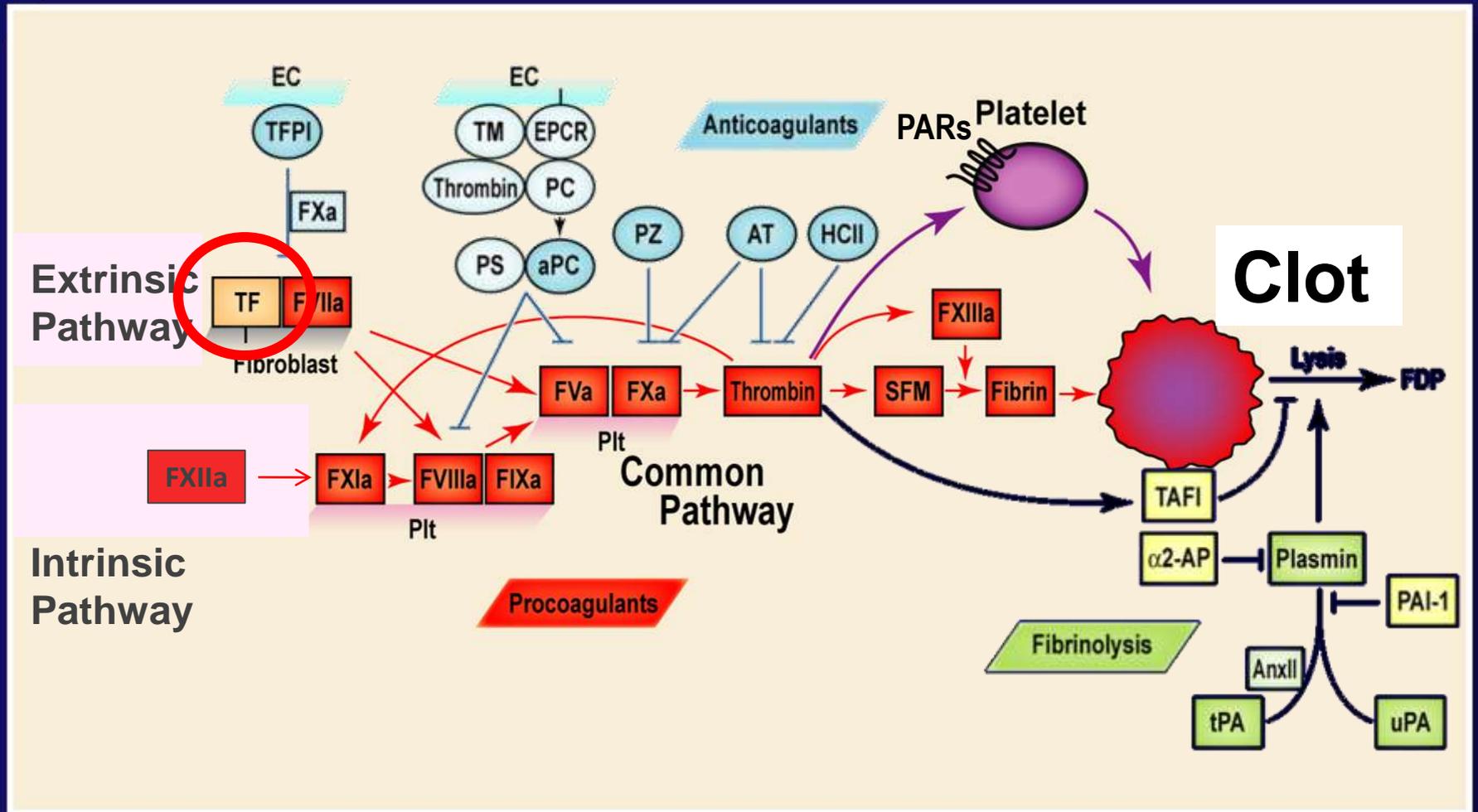
BxPc-3



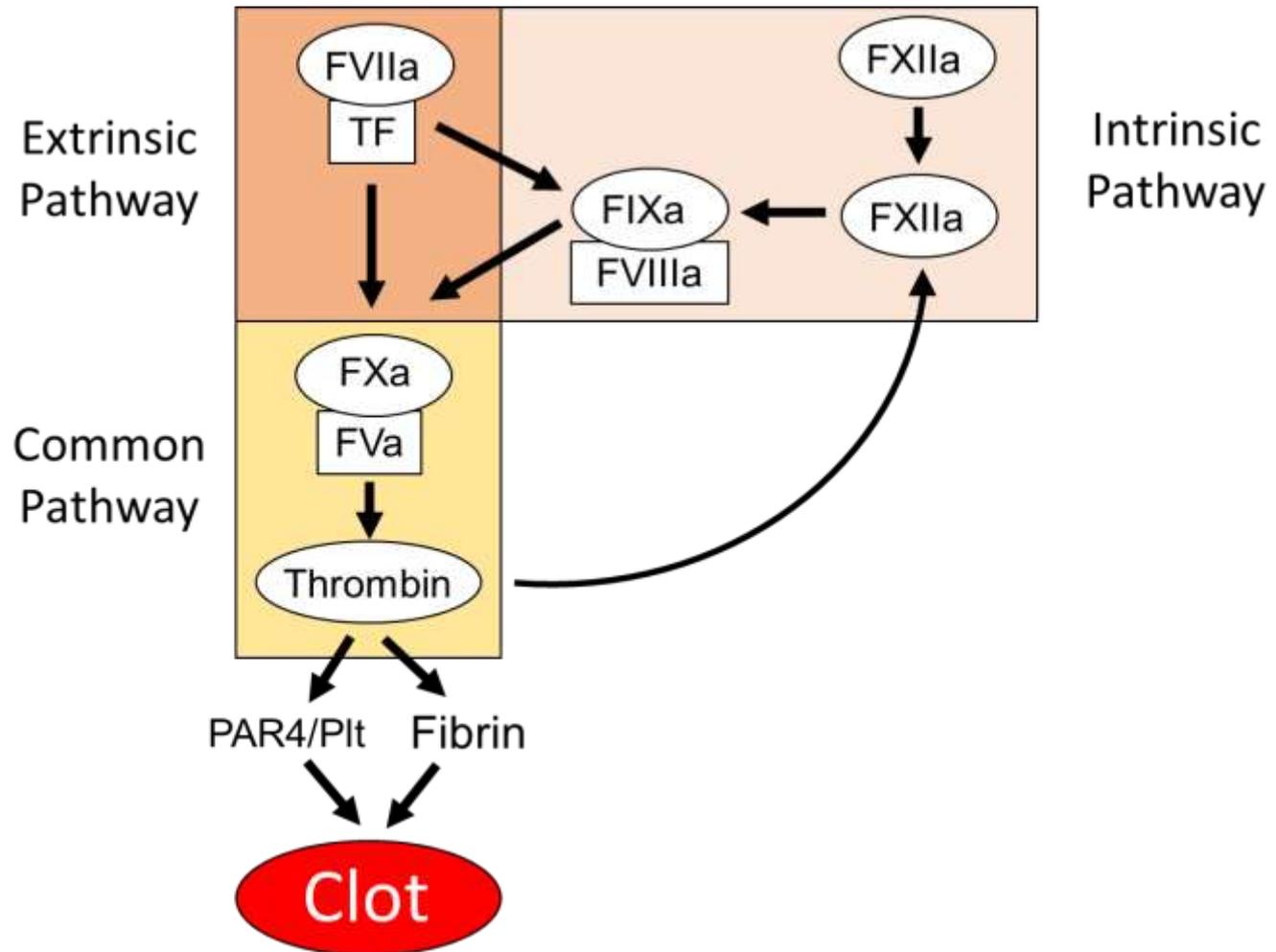
# Terminology

- **Hemostasis** - the arrest of bleeding
- **Thrombosis** - clotting within a blood vessel that may cause reduce blood flow and tissue oxygenation
- **Hemophilia** - a disorder of the blood marked by a permanent tendency to hemorrhages
- **Thrombophilia** - a disorder of the blood in which there is a tendency to the occurrence of thrombosis

# Formation and Lysis of a Clot



# Simplified Blood Coagulation Cascade

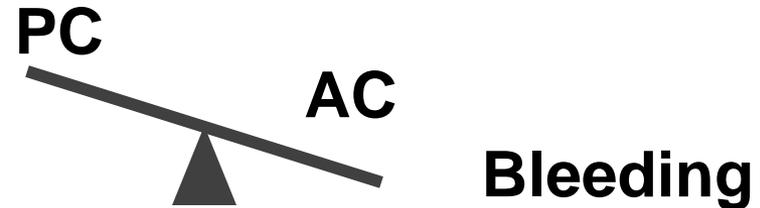


# Balancing Clotting

Normal Hemostasis



Hemophilia  
A (FVIII), B (IX) and C (XI)



Thrombophilia

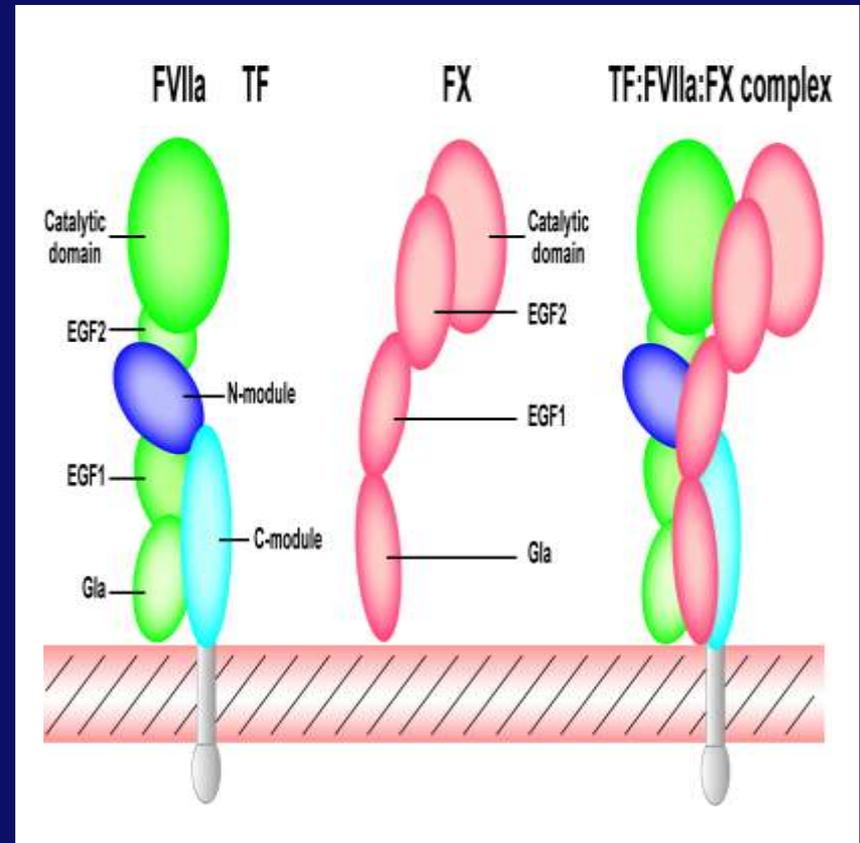


Restore hemostasis in hemophilia patients by decreasing levels of ACs

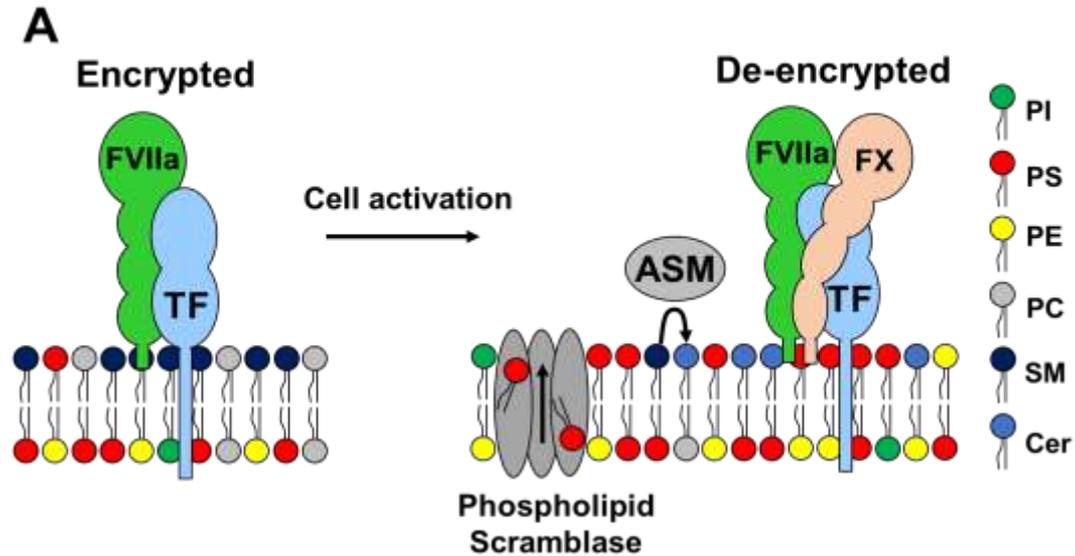
PC, procoagulant; AC, anticoagulant

# Tissue Factor (TF): The Primary Activator of Blood Coagulation

- 47 kDa transmembrane glycoprotein
- Receptor for FVII/FVIIa
- The TF:FVIIa complex activates both FIX and FX

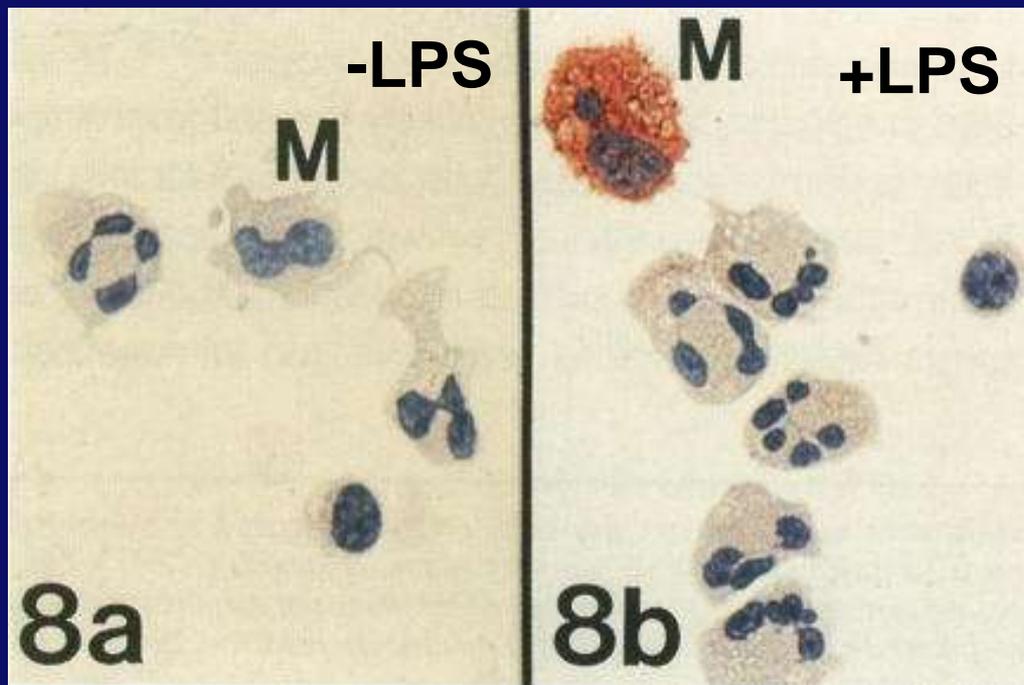


# Mechanisms of TF De-encryption



# LPS Induction of TF Expression in Monocytes

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Host Defense

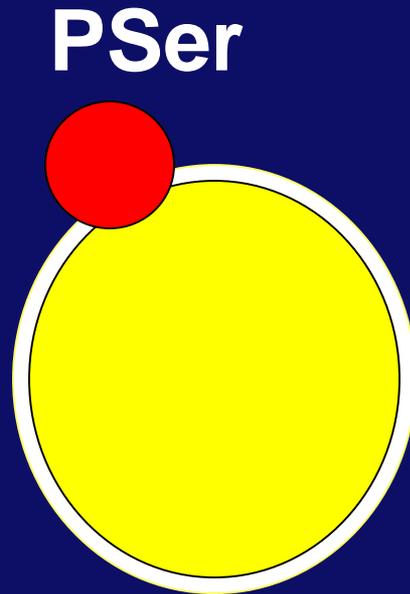
6 hrs

Drake et al AJP 1989

# Types of Procoagulant MVs

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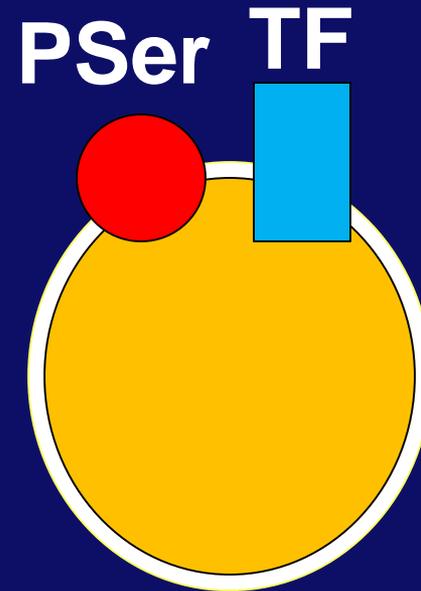
Microvesicles (MV) were originally defined as small (0.1-1  $\mu\text{m}$ ) membrane vesicles that are released from activated or apoptotic cells



Procoagulant  
activity  
Origin

++

Platelet/  
RBC

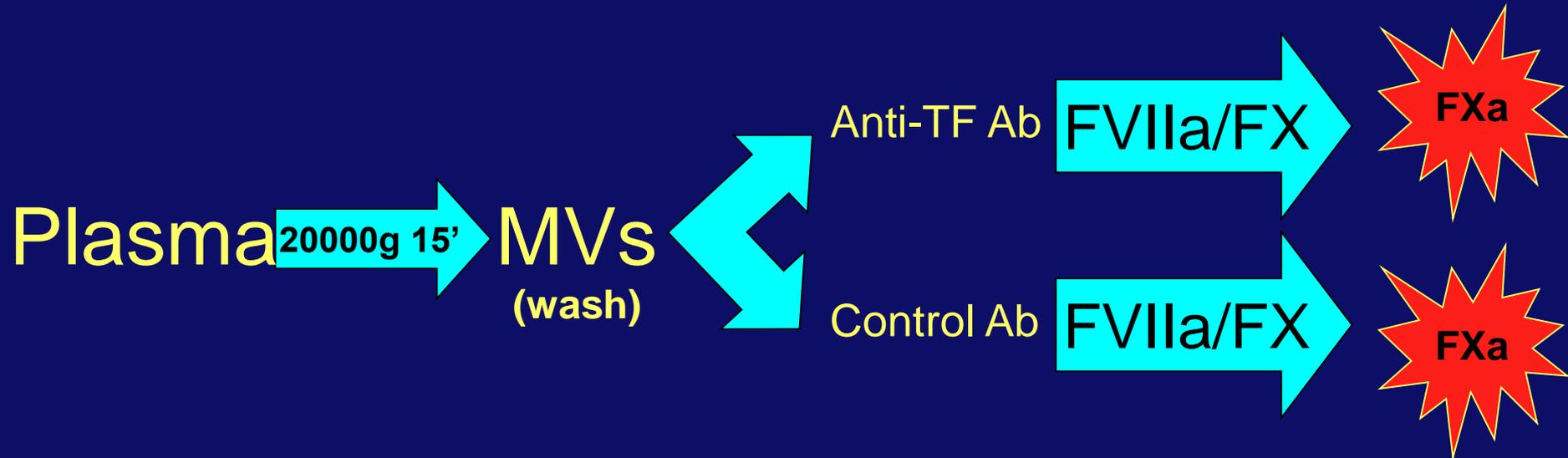


+++

Monocyte/  
tumor cell

# Development of a New Assay to Measure Levels of MV TF Activity in Plasma

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**TF-dependent factor Xa activity**

**Healthy individuals do not have detectable levels of TF<sup>+</sup> MVs in the circulation**

# Diseases with Elevated Levels of Circulating TF+ MVs

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- Cancer
- Endotoxemia
- Sickle cell disease
- Liver injury

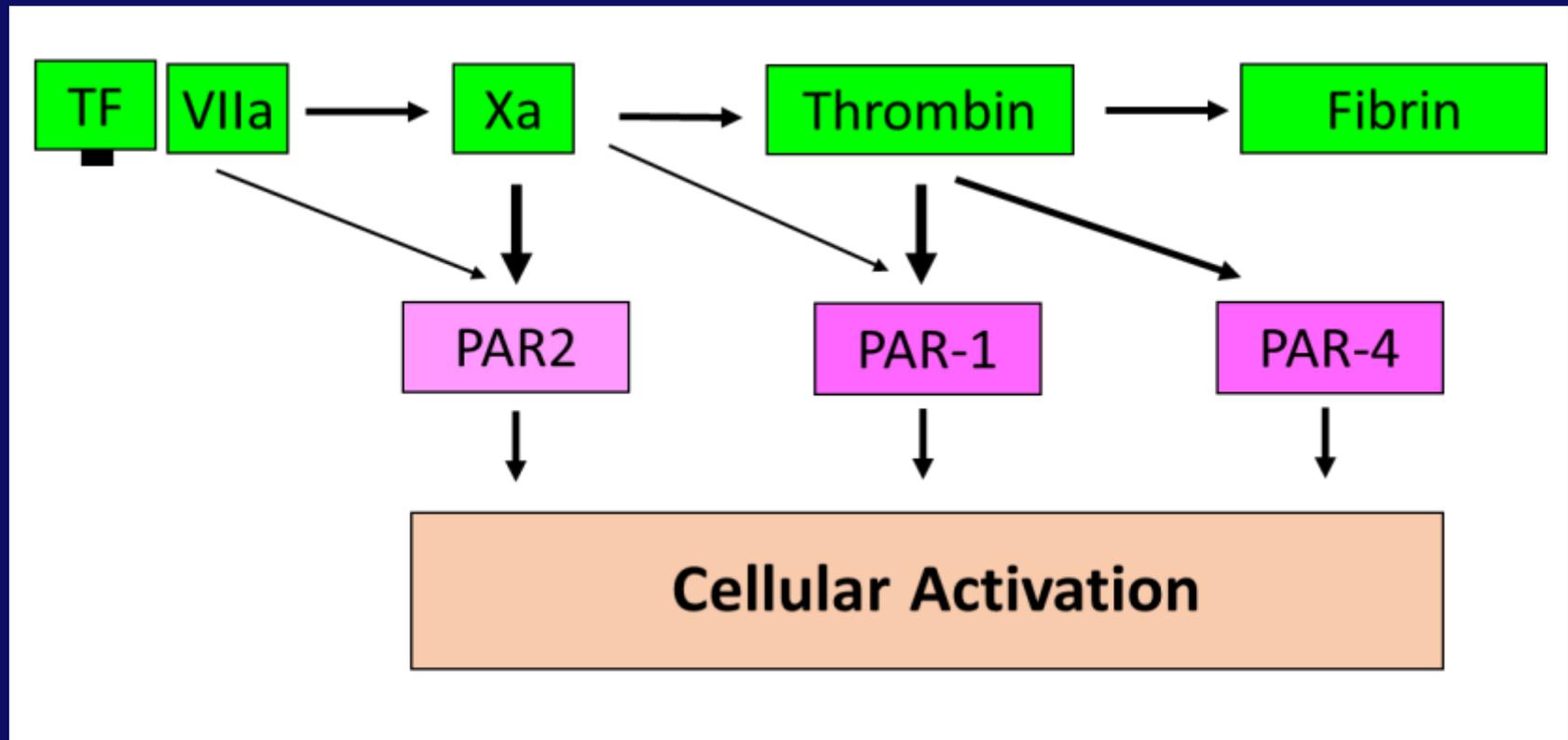
**MV TF = biomarker of thrombotic risk**

- Severe influenza A/H1N1 infection

Owens A and Mackman N Circ Res 2011;  
Hisada Y et al Throm Res 2016

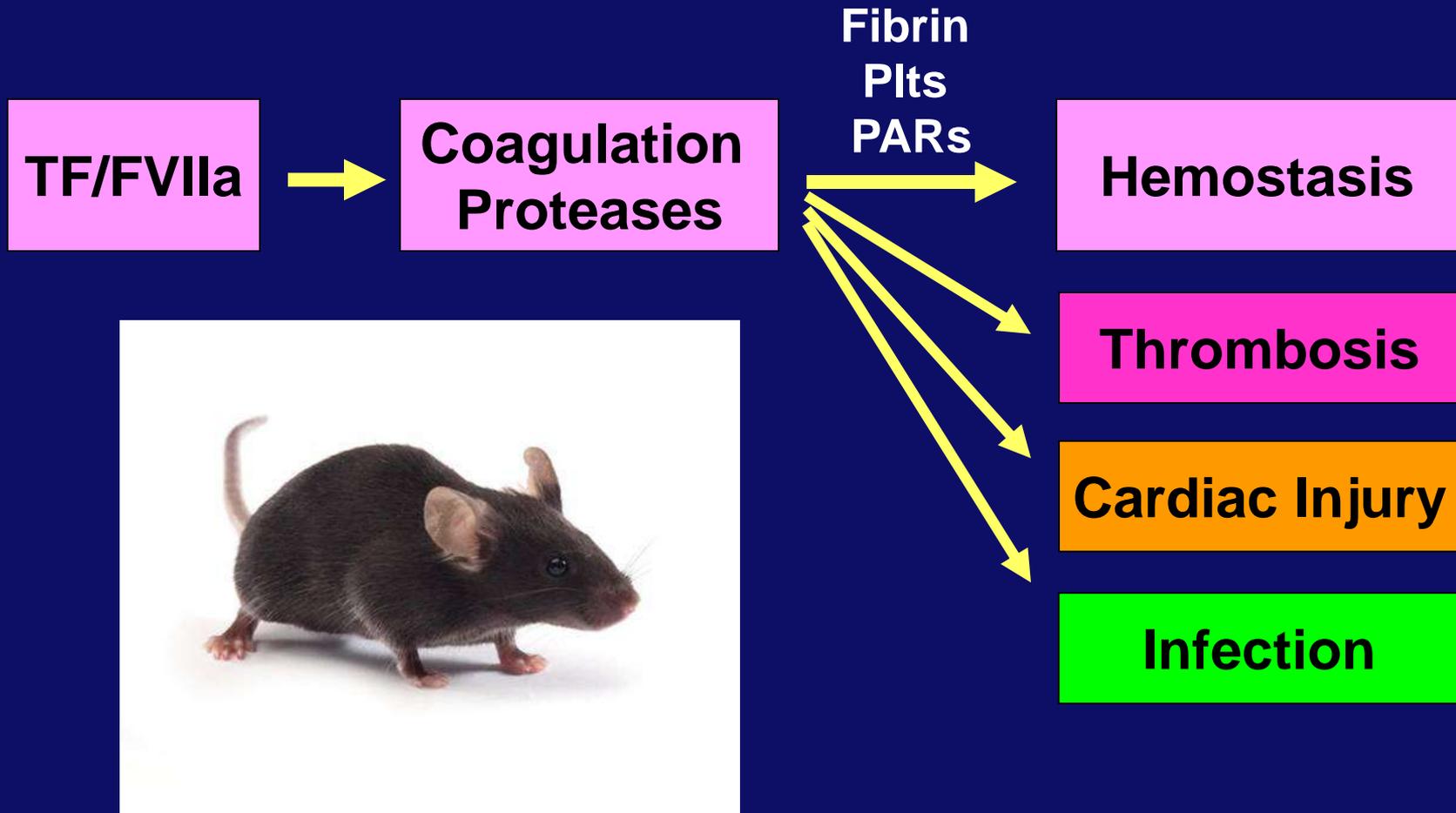
# Coagulation Proteases Activate PARs

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# Mouse Models to Study Disease

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# Hemostasis

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- **Hemostasis** is a process which causes bleeding to stop. It is the first stage of wound healing. This involves coagulation, blood changing from a liquid to a gel (Wikipedia). Hemostasis also reduces the risk of infection.

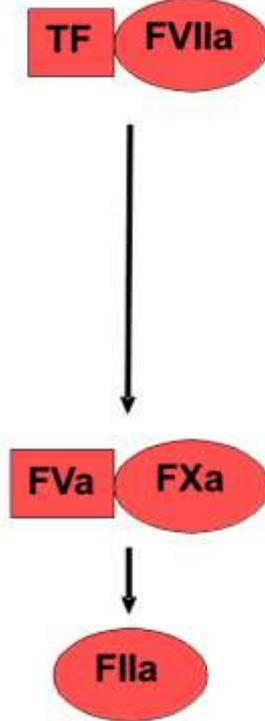


# Effect of Deletion of Different Procoagulant and Anticoagulant Proteins in Mice

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# Effect of a Deficient of Extrinsic Pathway Factors on Survival in Mice

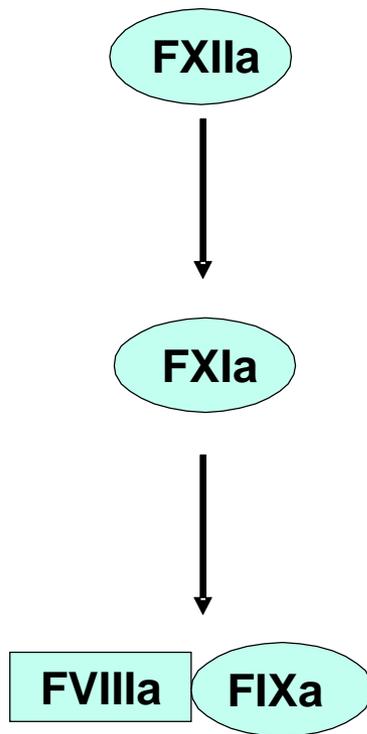


All die before wean

- TF** Bugge PNAS 1996  
Carmeliet Nature 1996  
Toomey Blood 1996
- FVII** Rosen Nature 1997
- FX** Dewerchin T&H 2000
- FV** Cui Nature 1996
- FII** Sun PNAS 1998

# Effect of a Deficiency of Contact and Intrinsic Pathway Factors on Survival in Mice

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Mice are Viable

**FXII** Pauer T&H 2004

**FXI** Gailani B C & F 1997

**FIX** Lin Blood 1997

**FVIII** Bi Nat Genet 1995

# Hemostatic Defects in Mice Lacking FVIII, FIX or FXI

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Mouse	Increase in tail bleeding
FXII <sup>-/-</sup>	No
FXI <sup>-/-</sup>	No
FIX <sup>-/-</sup>	Yes
FVIII <sup>-/-</sup>	Yes

# Effect of a Deficiency of Anticoagulant Proteins on Survival in Mice

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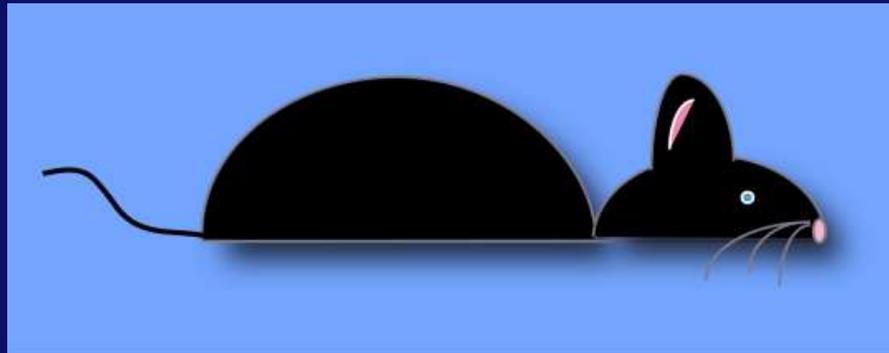
Gene	Survival at wean (%)	
TFPI	0	Huang Z et al Blood 1997
AT	0	Ishiguro K et al JCI 2000
TM	0	Healy A et al PNAS 1995
EPCR	0	Gu J et al JBC 2002
PC	0	Jalbert L et al JCI 1998
PS	0	Burstyn-Cohen et al JCI 2009

Each of the three anticoagulant pathways is required for regulation of the coagulation protease cascade

# Rescue of mTF<sup>-/-</sup> Embryos with a Human TF (hTF) Transgene

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Cross = mTF<sup>+/-</sup>,hTF<sup>+</sup> x mTF<sup>+/-</sup>,hTF<sup>+</sup>

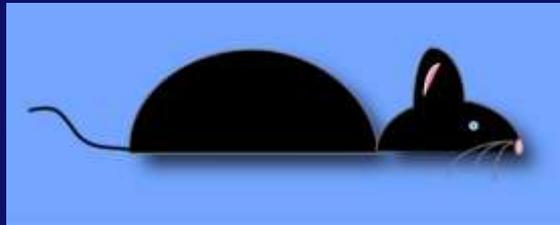


Low TF mice mTF<sup>-/-</sup>,hTF<sup>+</sup>

**Low FVII mice** Rosen E et al T&H 2005

# Generation of Mice with Cell Type-specific Deletion of the TF Gene

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**TF<sup>flox/flox</sup> mice**

Cell type-specific deletion of the TF gene when  
crossed with different Cre<sup>+</sup> lines

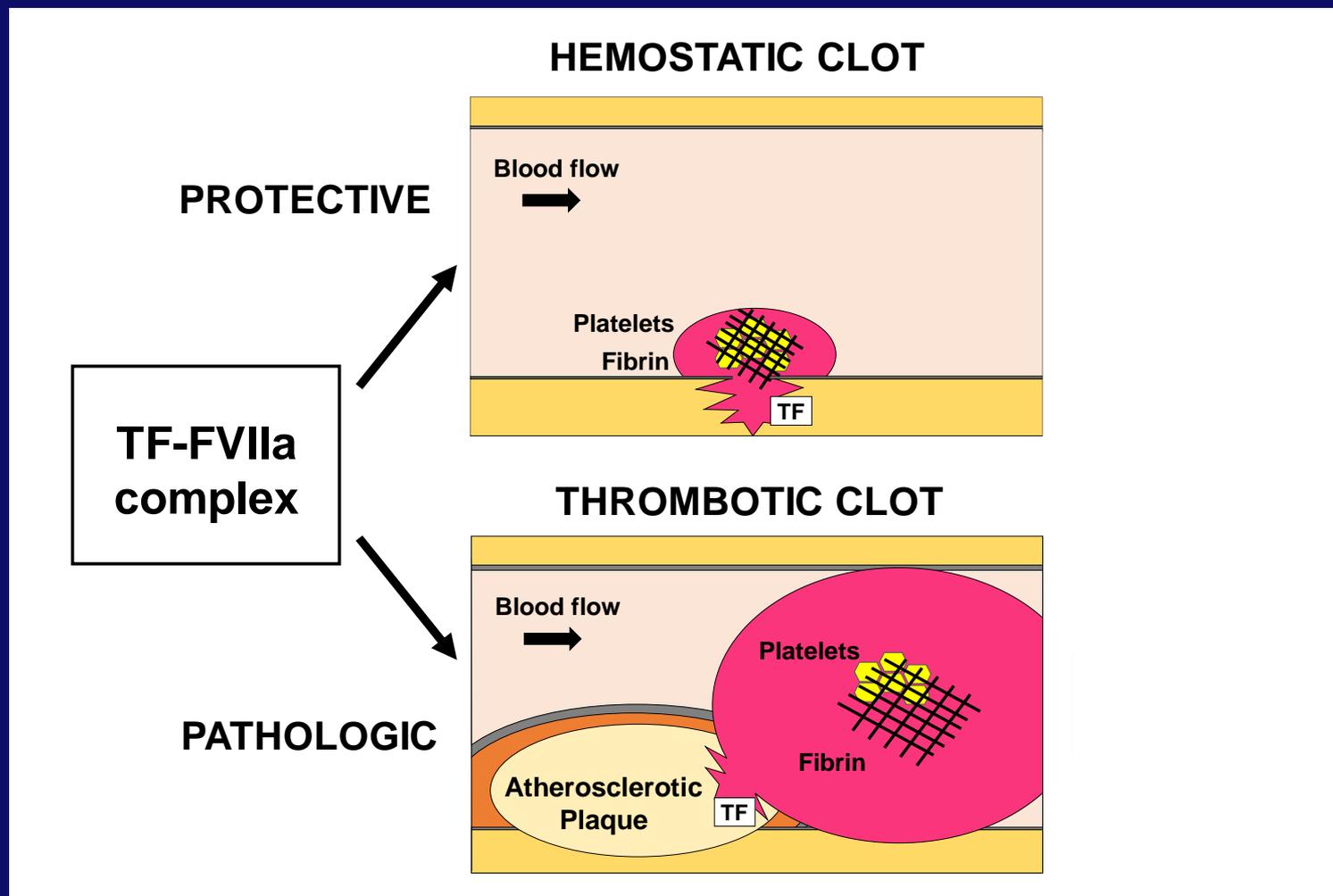
**Pawlinski ..Mackman JTH 2007**

# Generation of Mice with Cell Type-Specific Deletion of the TF Gene

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- **Mlc2v-Cre (cardiomyocytes).** Pawlinski et al, JTH 2007.
- **SM22-Cre (vascular smooth muscle cells).** Wang et al, Blood 2009.
- **LysM-Cre (myeloid cells).** Pawlinski et al, Blood 2010.
- **Tie2-Cre (EC and hemat cells).** Pawlinski et al, Blood 2010.
- **Alb-Cre (hepatocytes).** Sullivan et al, Blood 2013.
- **SPC-Cre (lung epithelial cells).** Shaver et al, AJRCMB 2015.
- **Nestin-Cre (CNS).** Wang et al JCI Insight 2016.

# Roles of TF in Hemostasis and Thrombosis

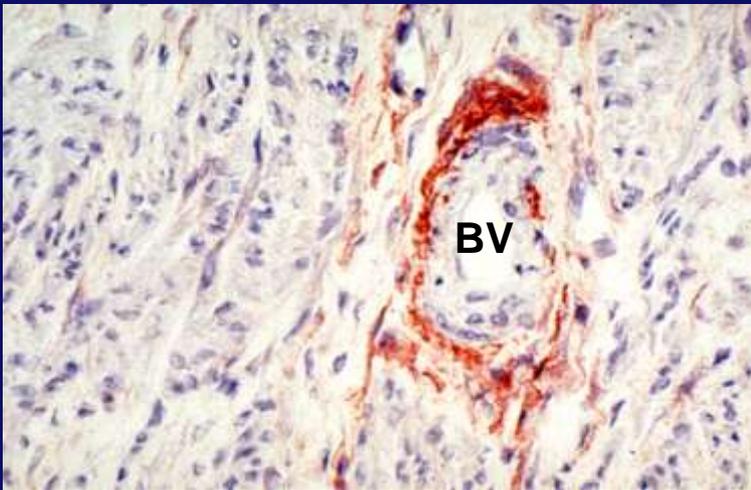


Grover and Mackman ATVB in press

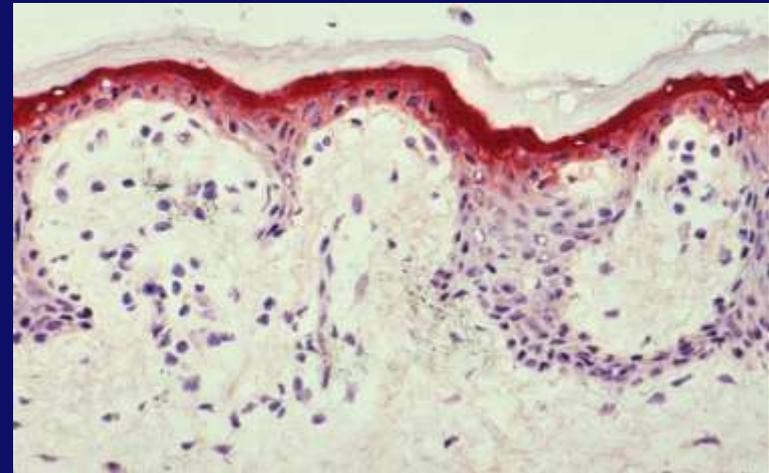
# Role of TF in Hemostasis

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**Blood Vessel**



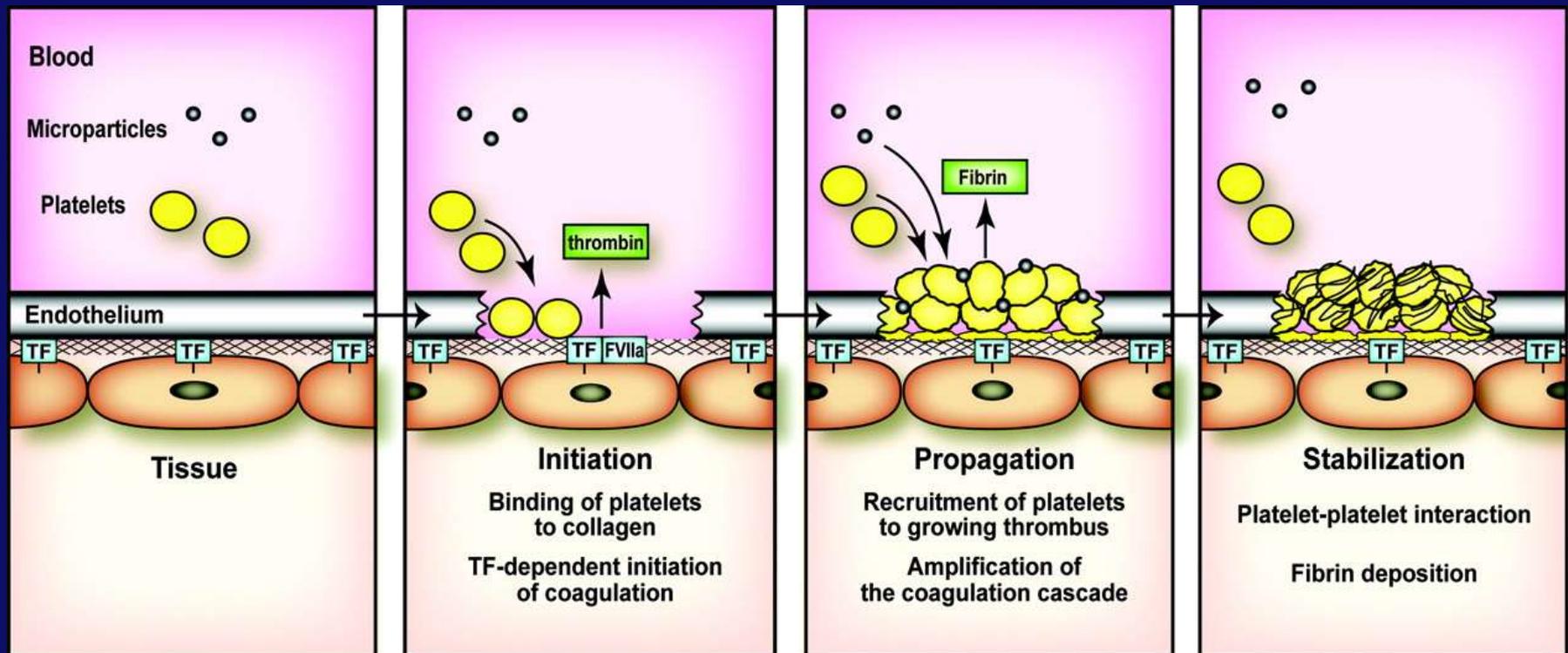
**Skin**



**TF forms a “hemostatic envelope”  
around blood vessels and body surfaces**

**Drake et al AJP 1989**

# Perivascular TF Plays a Critical Role in Hemostasis After Vessel Injury



TF is the only clotting factor not present in blood

# Expression of TF in Different Organs

---

**High**

**uterus**

**skin**

**brain**

**lung**

**placenta**

**Medium**

**heart**

**kidney**

**testis**

**Low**

**skeletal muscle**

**liver**

**spleen**

**thymus**

**Hypothesis:**

**TF provides additional hemostatic protection to select tissues**

**Analyze the hemostatic defects in Low TF mice**

# Spontaneous Hemostatic Defects in Low TF Mice

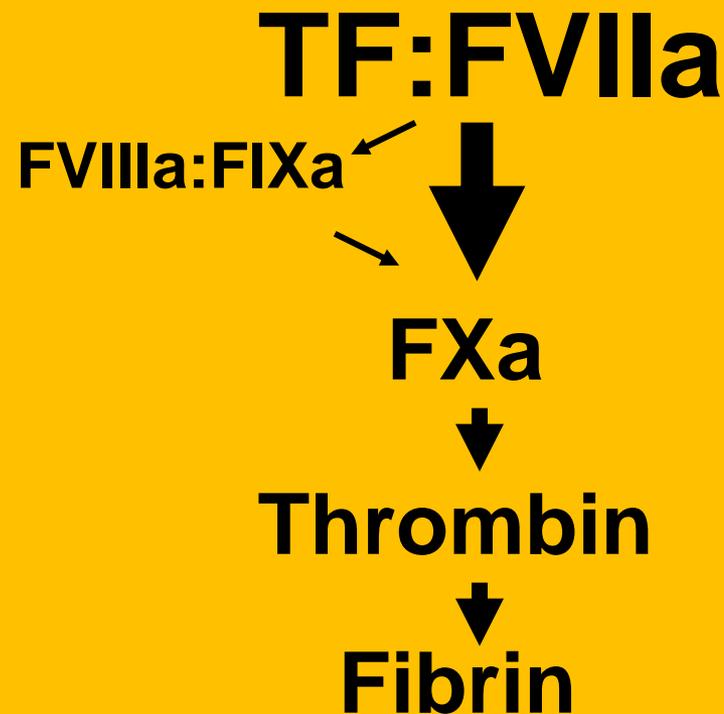
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<b>Uterus</b>	Fatal hemorrhage post-partum	Erlich PNAS 1999
<b>Placenta</b>	Blood pools	Erlich PNAS 1999
<b>Heart</b>	Hemosiderin, fibrosis	Pawlinski PNAS 2002
<b>Lung</b>	Hemosiderin, fatal hemorrhages	Pedersen Blood 2005
<b>Testis</b>	Hemorrhage, calcification	Mackman JTH 2007
<b>Brain</b>	Occasional brain hemorrhage	Pawlinski T&H 2004

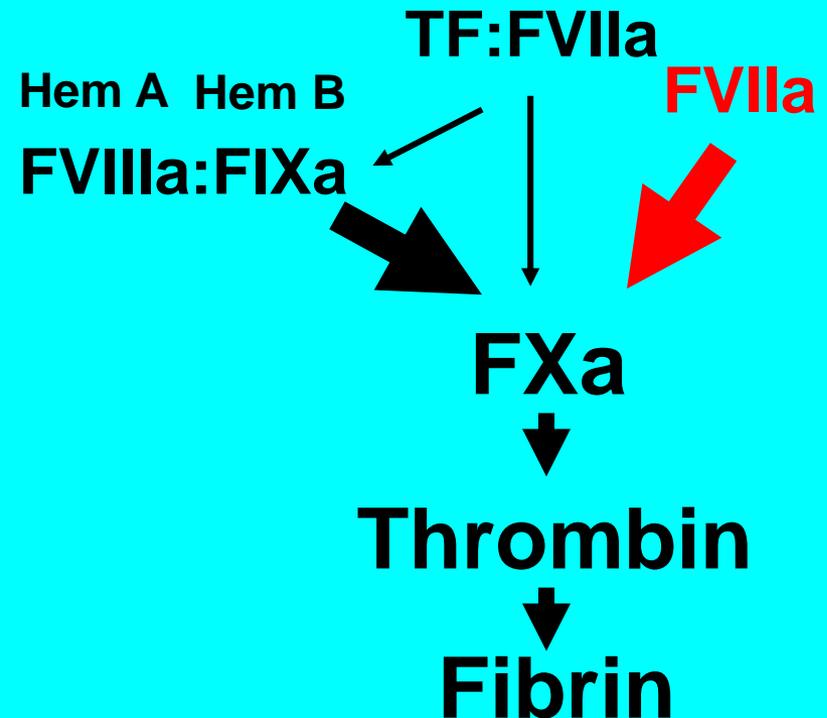
**Blood vessels in the heart and lung may be more prone to mechanical damage**

# Tissue Specific Hemostasis

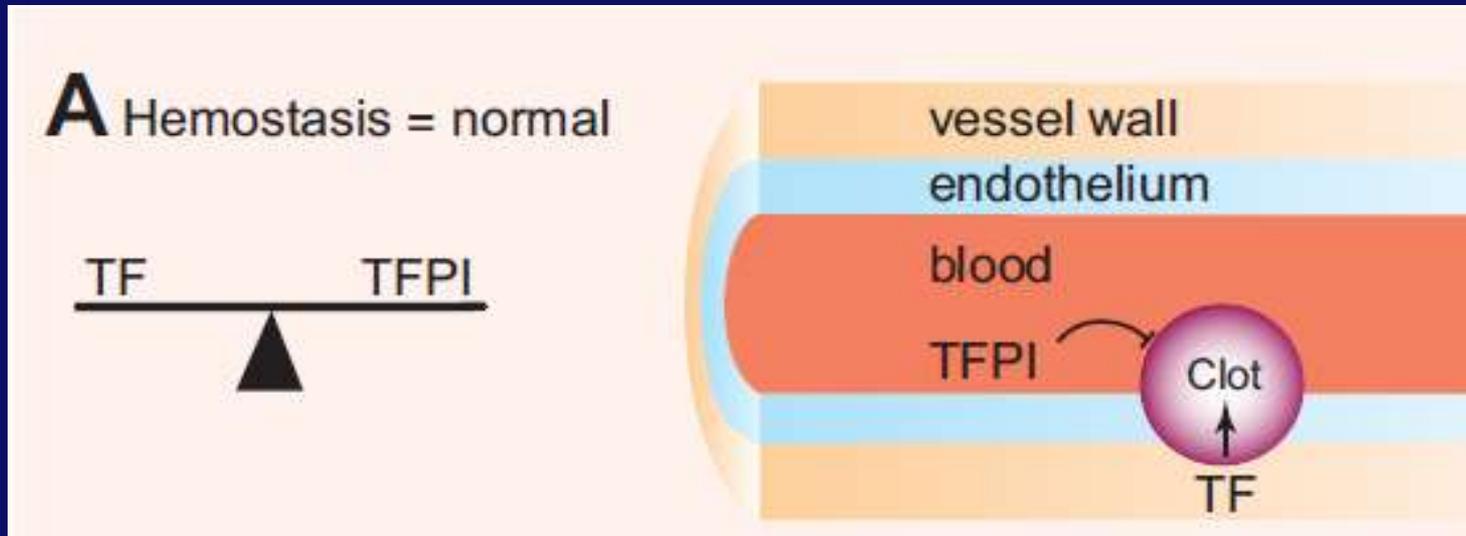
High TF: uterus, brain, heart, lung, placenta, testis



Low TF: skeletal muscle and joints



# Balancing the TF Pathway



# Re-balancing the Clotting Cascade in Mice

---

Can we rescue the lethality of embryos lacking different anticoagulants with low levels of TF?

1/ TFPI<sup>-/-</sup> embryos are rescued by reducing the level of TF.

2/ Fatal bleeding in low TF mice is rescued by reducing the level of TFPI.

# Re-balancing the Clotting Cascade in Mice

---

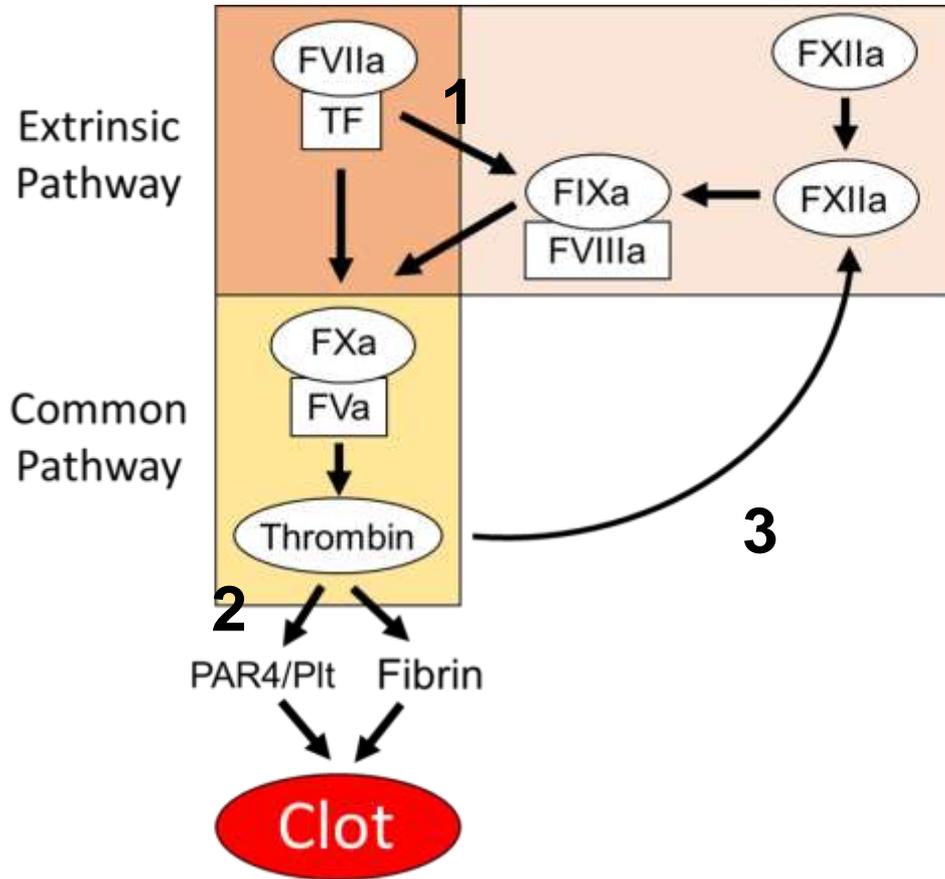
**EPCR<sup>-/-</sup> x Low TF- rescue**

Li W et al Blood 2005

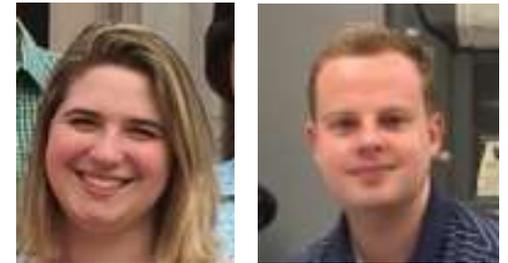
**AT<sup>-/-</sup> x Low TF- rescue**

Hayashi M et al Blood 2005

# Analysis of Different Pathways using a Genetic Approach



Intrinsic Pathway



- 1/ TF:FVIIa – FIX
- 2/ Thrombin – PAR
- 3/ Thrombin – FXI

# Effect of Combining Low Levels of TF with FIX Deficiency

Breeding Male  $mTF^{-/-}$ ,  $hTF^{+/+}$ ,  $FIX^{+/y}$  x female  $mTF^{+/-}$ ,  $hTF^{-/-}$ ,  $FIX^{+/-}$

Sex	<u>mTF</u>	<u>hTF</u>	F9	Expected (%)	E14.5 (n)	E14.5 Observed (%)
F	+/-	+	+/+	12.5	7	18.4
F	+/-	+	+/-	12.5	7	18.4
M	+/-	+	+/y	12.5	1	2.6
M	+/-	+	-/y	12.5	4	10.5
F	-/-	+	+/+	12.5	5	13.2
F	-/-	+	+/-	12.5	5	13.2
M	-/-	+	+/y	12.5	5	13.2
M	-/-	+	-/y	12.5	4	10.5
Total				100	38	100

\*2 dead

**wean**

Grover unpublished

# Low TF, FIX<sup>-/-</sup> P1 Pups Die after Birth

mTF+/-, hTF+, FIX+/-

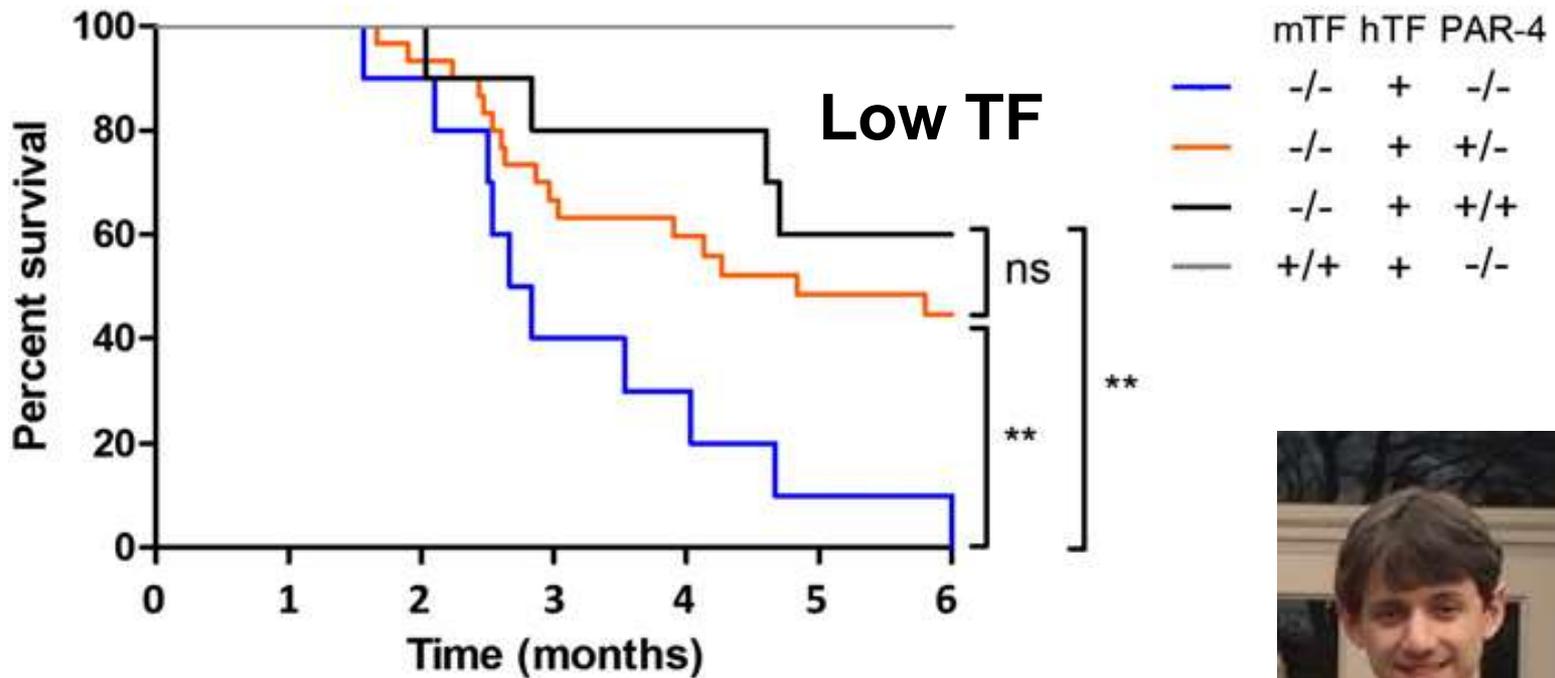


mTF-/-, hTF+, FIX-/-



# Effect of Combining Low Levels of TF with PAR4 Deficiency

Expected numbers of low TF, PAR4 deficient mice at wean



# Effect of Combining Low Levels of TF with FXI Deficiency

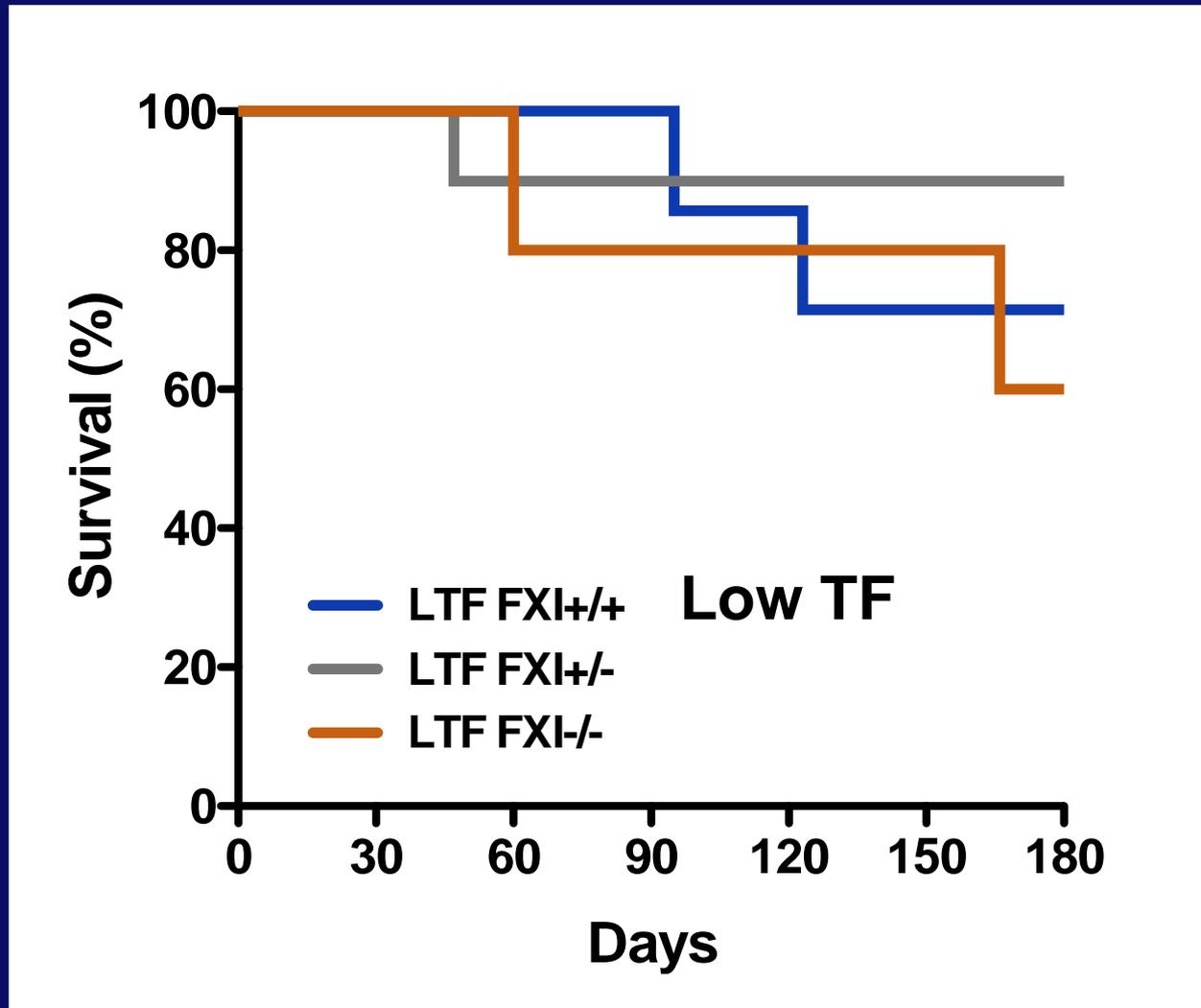
Breeding- male mTF<sup>+/-</sup>, hTF<sup>+/-</sup>, FXI<sup>+/-</sup> x female mTF<sup>+/-</sup>, hTF<sup>+/-</sup>, FXI<sup>+/-</sup>

mTF	hTF+	FXI	Observed (N)	Observed (%)	Expected (%)
+/+	-	+/+	4	4.55	1.56
+/+	-	+/-	3	3.41	3.13
+/+	-	-/-	3	3.41	1.56
+/+	+	+/+	5	5.68	4.69
+/+	+	+/-	4	4.55	9.38
+/+	+	-/-	2	2.27	4.69
+/-	-	+/+	2	2.27	3.13
+/-	-	+/-	7	7.95	6.25
+/-	-	-/-	1	1.14	3.13
+/-	+	+/+	11	12.50	9.38
+/-	+	+/-	23	26.14	18.75
+/-	+	-/-	9	10.23	9.38
-/-	-	+/+	0	0.00	0
-/-	-	+/-	0	0.00	0
-/-	-	-/-	0	0.00	0

**Low TF, FXI<sup>-/-</sup> mice are generated at the expected frequency**

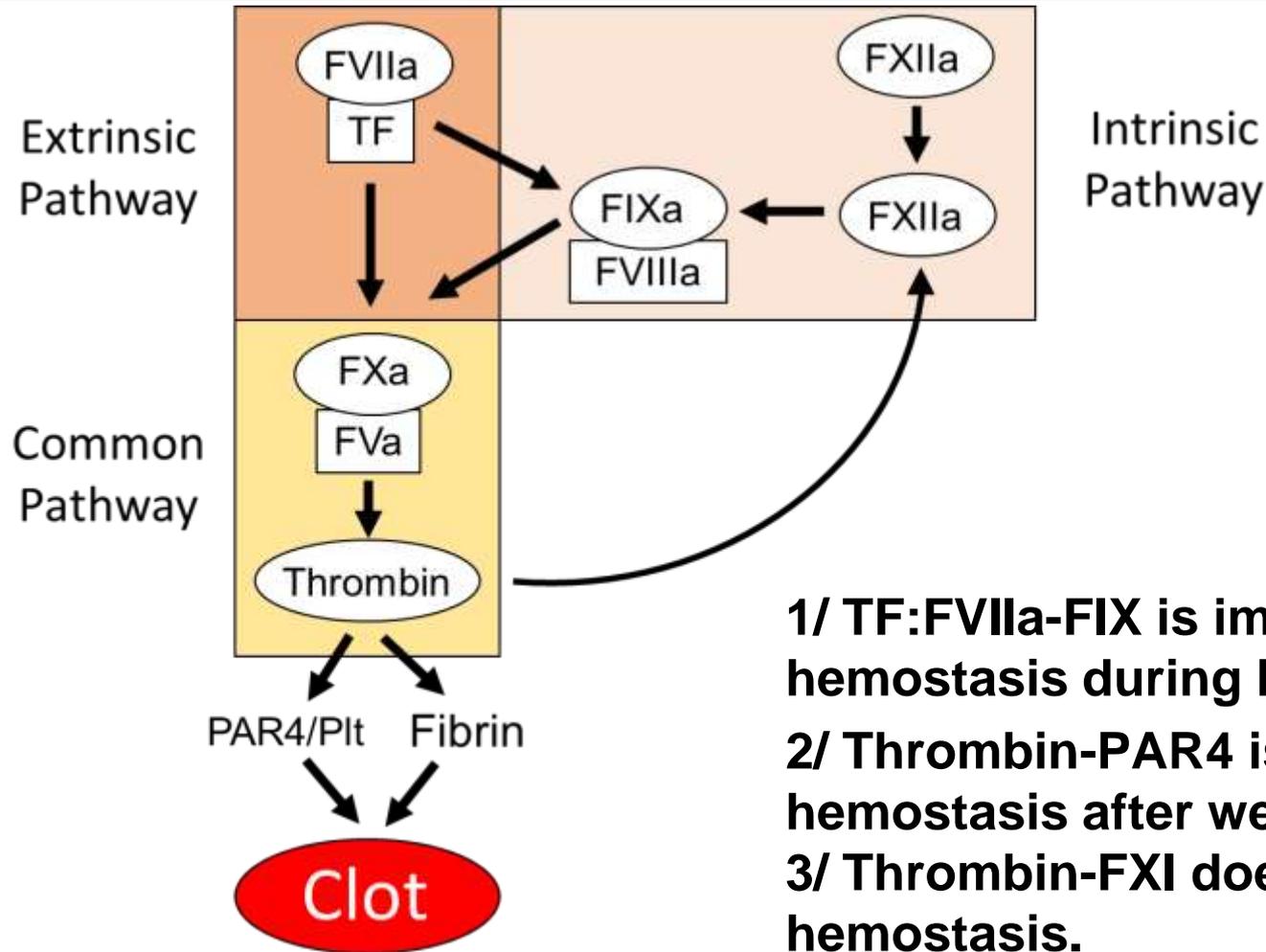
hTF+ = hTF<sup>+/-</sup> and hTF<sup>+/+</sup>

# Survival of Low TF, FXI<sup>-/-</sup> Mice



Grover unpublished

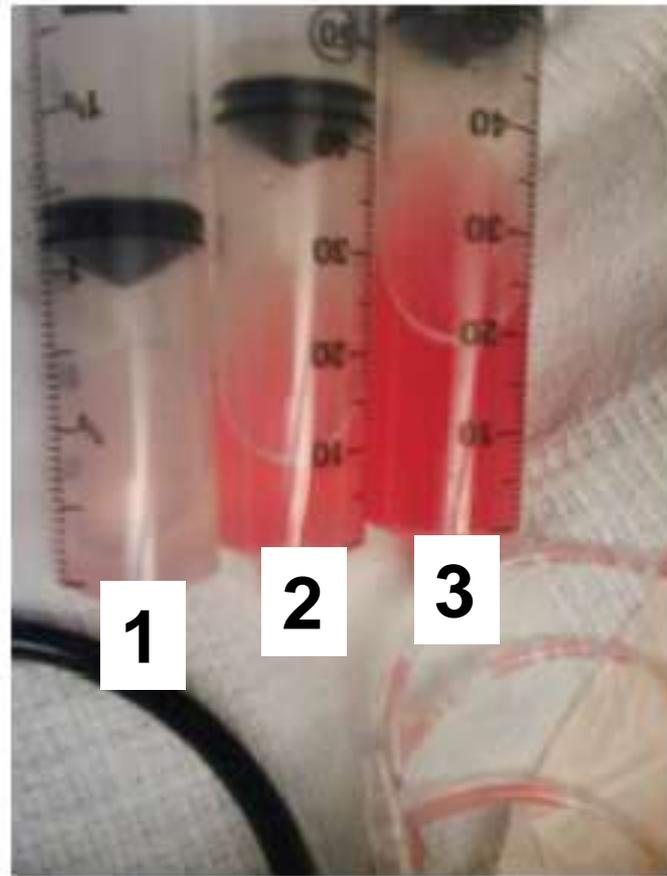
# Conclusions



- 1/ TF:FVIIa-FIX is important for hemostasis during birth.
- 2/ Thrombin-PAR4 is important for hemostasis after wean.
- 3/ Thrombin-FXI does not affect hemostasis.

# Alveolar Hemorrhage in Patients with Severe Influenza A Infection

Bronchiolar lavage fluid from a patient infected with influenza A



# Role of Lung Epithelial Cell TF in Influenza A Infection

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**Hypothesis:**

**A deficiency of TF in the lung will  
increased alveolar hemorrhage  
after influenza A infection**

**We used a genetic approach:  
(TF<sup>fl/fl</sup>, SPC-Cre mice)**

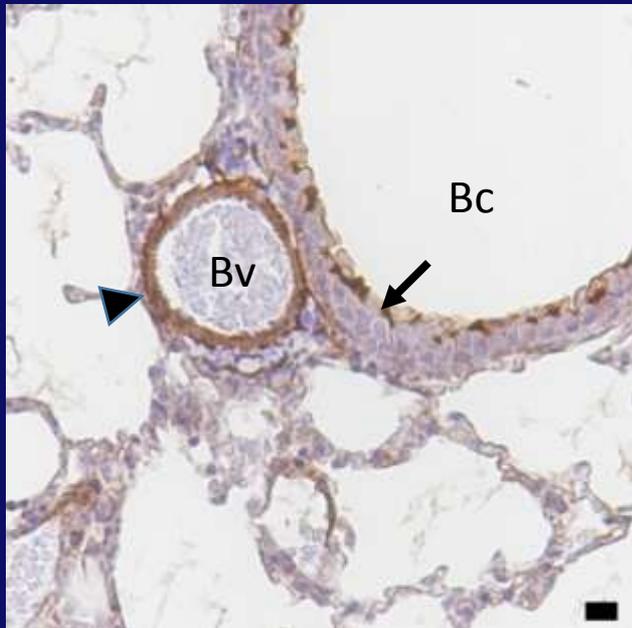


**Silvio Antoniak**

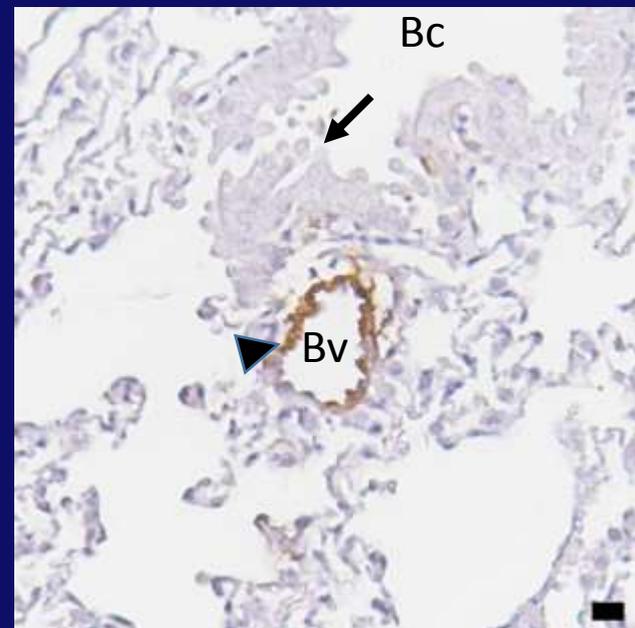
**SPC = surfactant protein C**

# Characterization of $TF^{fl/fl}$ , SPC-Cre Mice

$TF^{fl/fl}$



$TF^{fl/fl}$ , SPC-Cre<sup>+</sup>



$TF^{fl/fl}$ , SPC-Cre<sup>+</sup> have the TF gene deleted in all epithelial cells- bronchiolar and alveolar

# Mouse Model of Influenza A Infection (IAV)

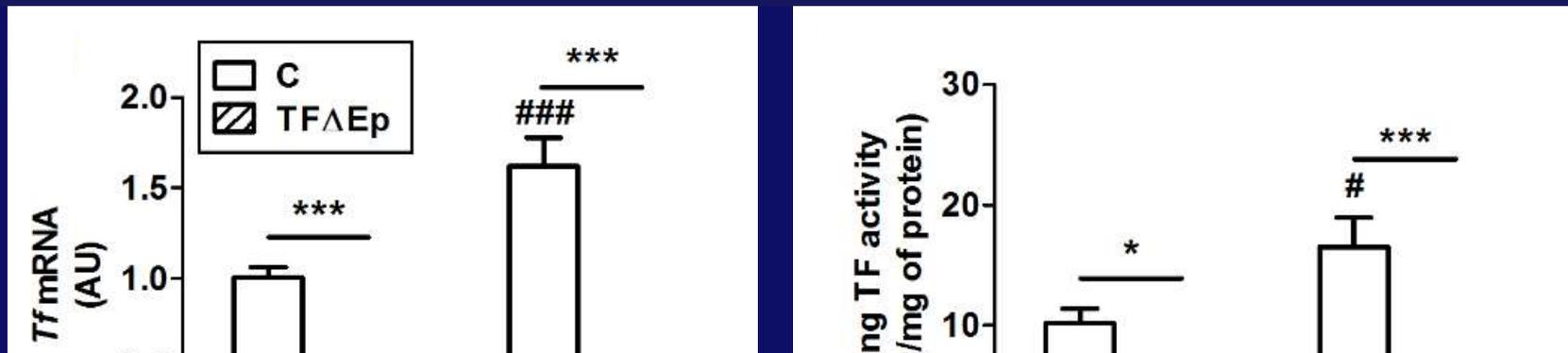
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- Influenza A is a ssRNA virus.
- Influenza A/Puerto Rico/8/1934 H1N1.
- Mouse adapted and highly pathogenic.
- Infected intranasally.

# Influenza A Infection Induces TF Expression in TF<sup>fl/fl</sup> Mice but not in TF<sup>fl/fl</sup>,SPC-Cre Mice

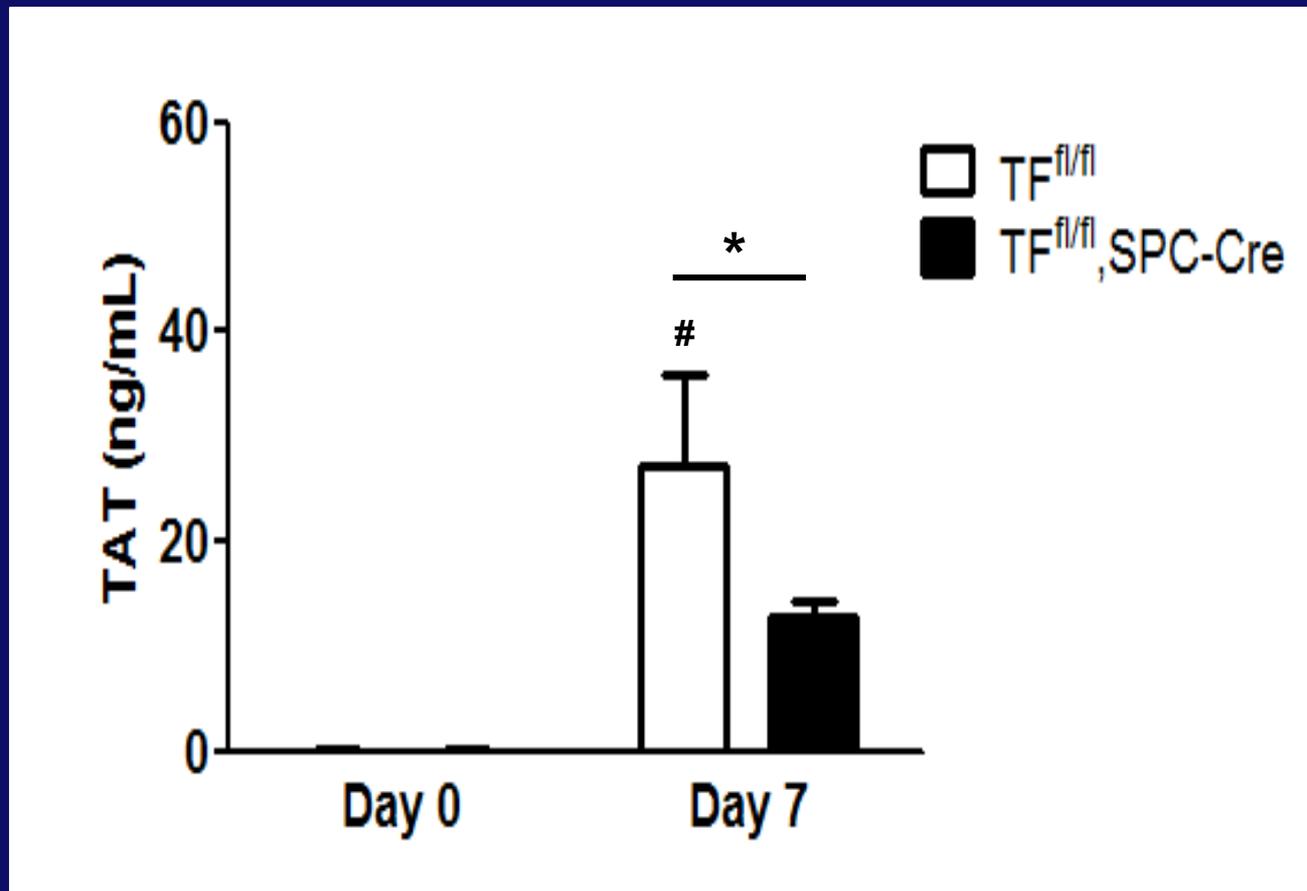
TF mRNA

TF activity



**Influenza A infection induces TF expression in lung epithelial cells-protective response**

# Reduced Activation of Coagulation in the Lungs of $TF^{fl/fl}$ ,SPC-Cre Mice after Influenza A Infection

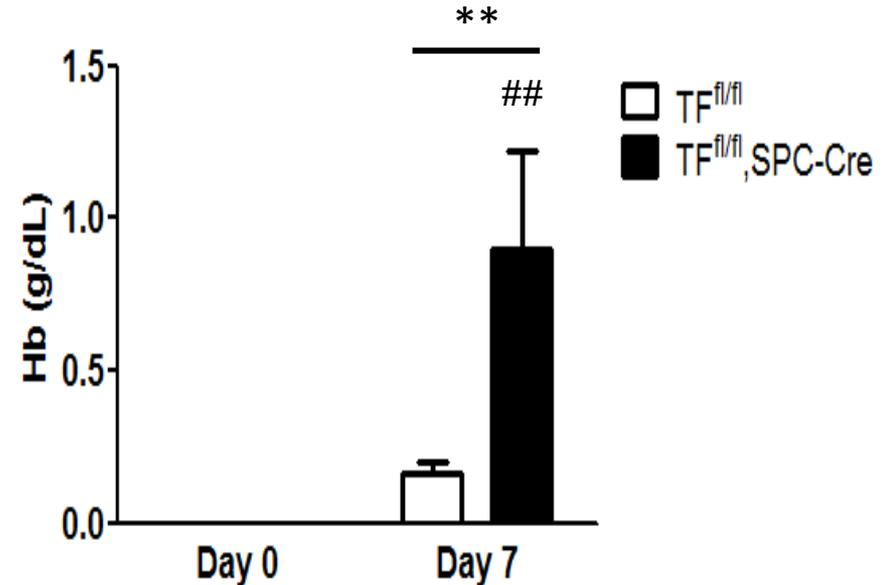
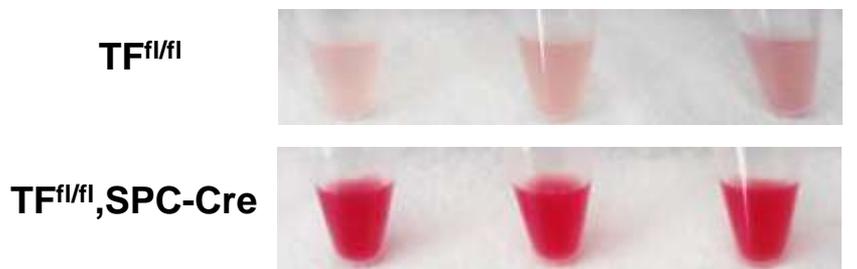


# TF<sup>fl/fl</sup>,SPC-Cre Mice have Increased Lung Hemorrhage after Influenza A Infection

Day 0



Day 7



# Conclusion

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**TF expression by lung epithelial cells is required to maintain hemostasis during influenza A infection**

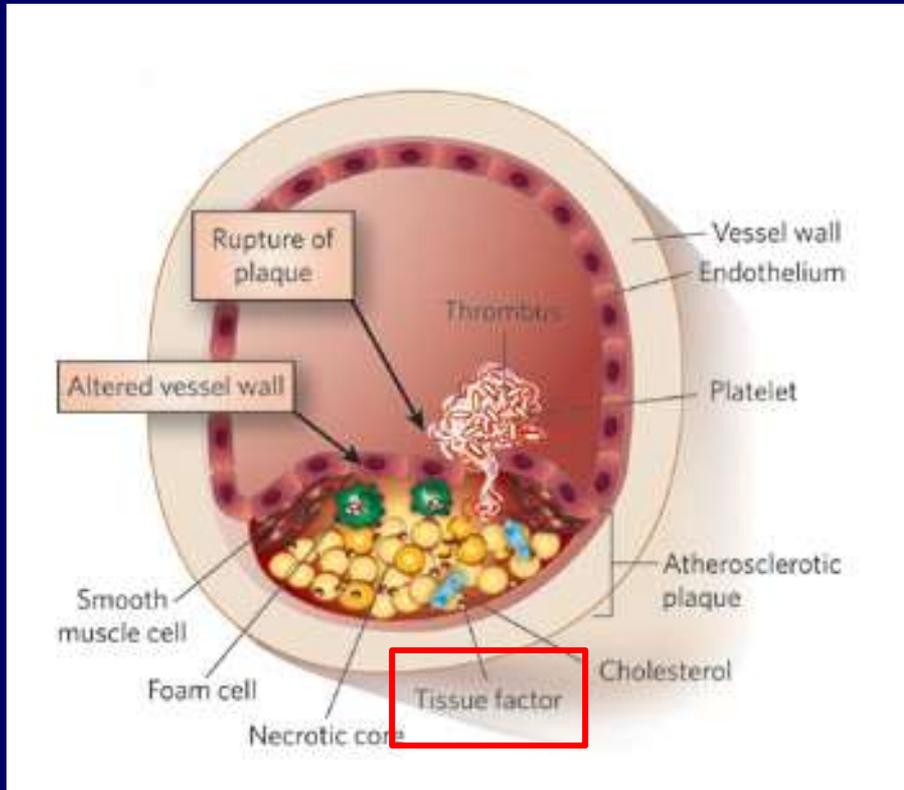
# Thrombosis

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- **Thrombosis** is the formation of a blood clot inside a blood vessel, obstructing the flow of blood through the circulatory system. Wikipedia Arterial thrombois (MI and stroke) and venous thrombosis.



# Arterial Thrombosis



- **Plaque rupture**
- **Platelet-rich thrombus**
- **TF activates clotting**
- **Clot forms rapidly**

# Mouse Model of Atherothrombosis

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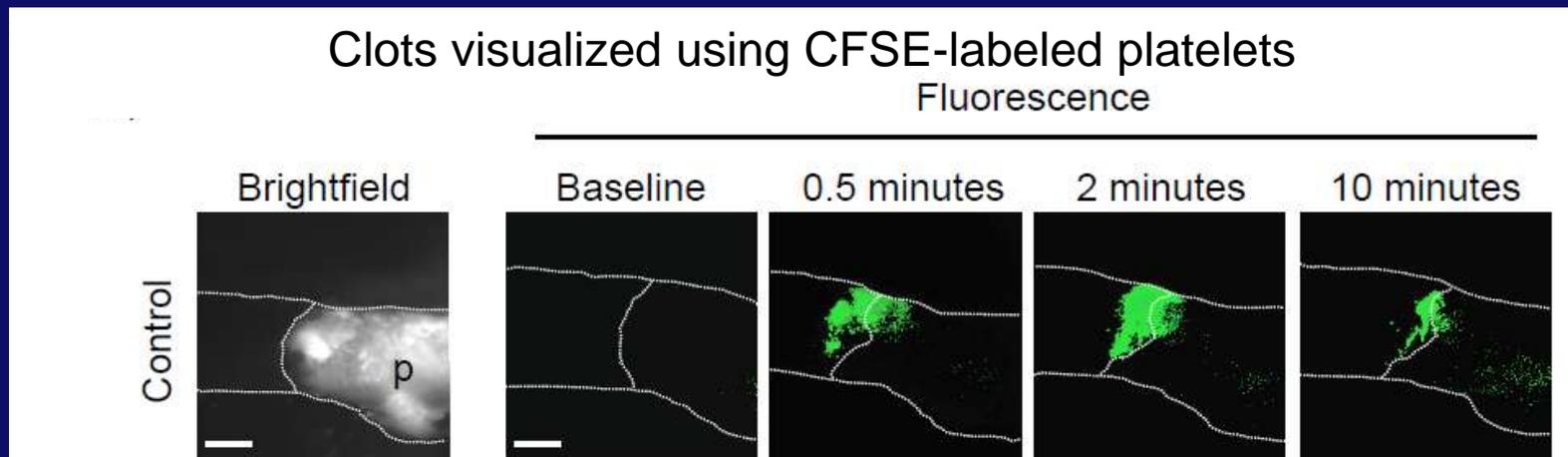
- ApoE<sup>-/-</sup> mice fed a western diet to induce atherosclerosis
- Plaques in the carotid arteries ruptured using ultrasound

Kuijper M et al ATVB 2014

Van Montfoort M et al ATVB 2014

Editorial Mackman N ATVB 2014

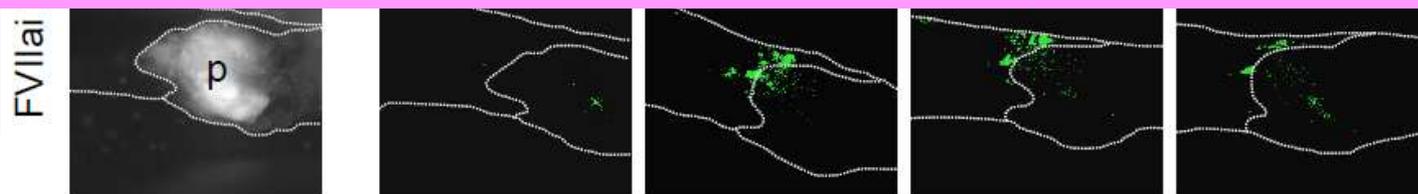
# Role of the Extrinsic (TF:FVIIa) and Intrinsic (FXIIa) Coagulation Pathways in Atherothrombosis



**The TF:FVIIa complex initiates clotting whereas FXII amplifies the thrombus**

Inh of  
of

Inhibition  
of TF:FVIIa



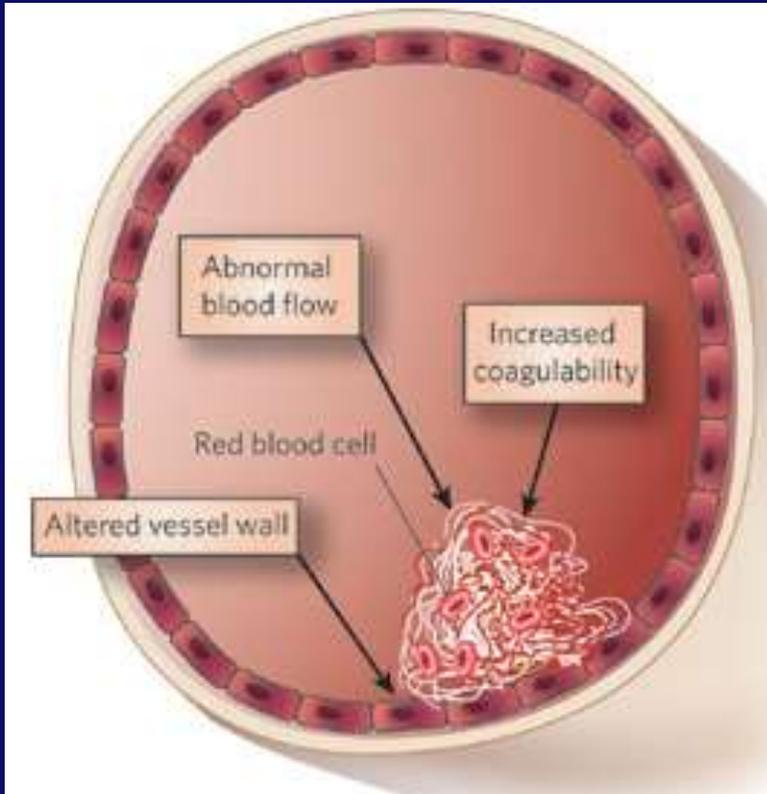
# Conclusion

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**TF in atherosclerotic plaques  
activates coagulation after plaque  
rupture**

# Venous Thrombosis

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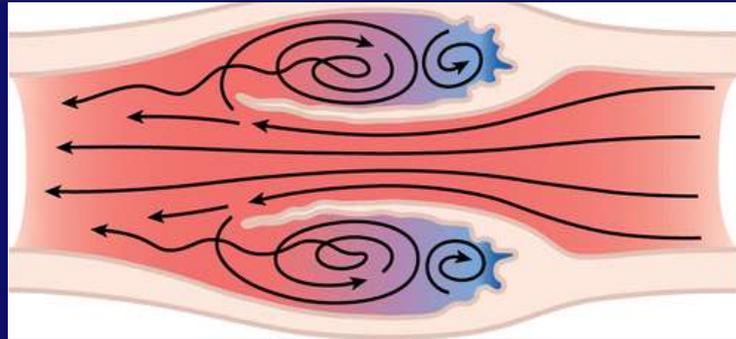
- Fibrin-rich
- Intact endothelium
- Form in valve pockets
- Form slowly (hours to days)

Venous thromboembolism (VTE) - DVT and PE

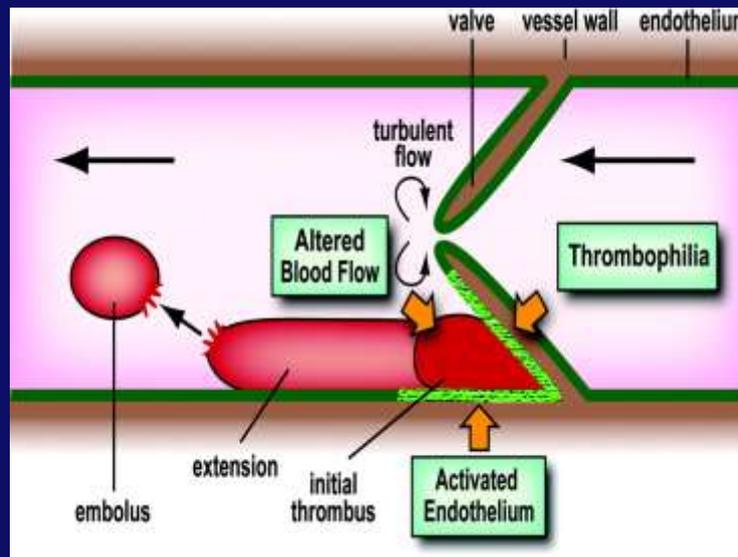
Mackman Nature 2008

# Formation of a Deep Vein Thrombus

Hypoxia in valve pockets

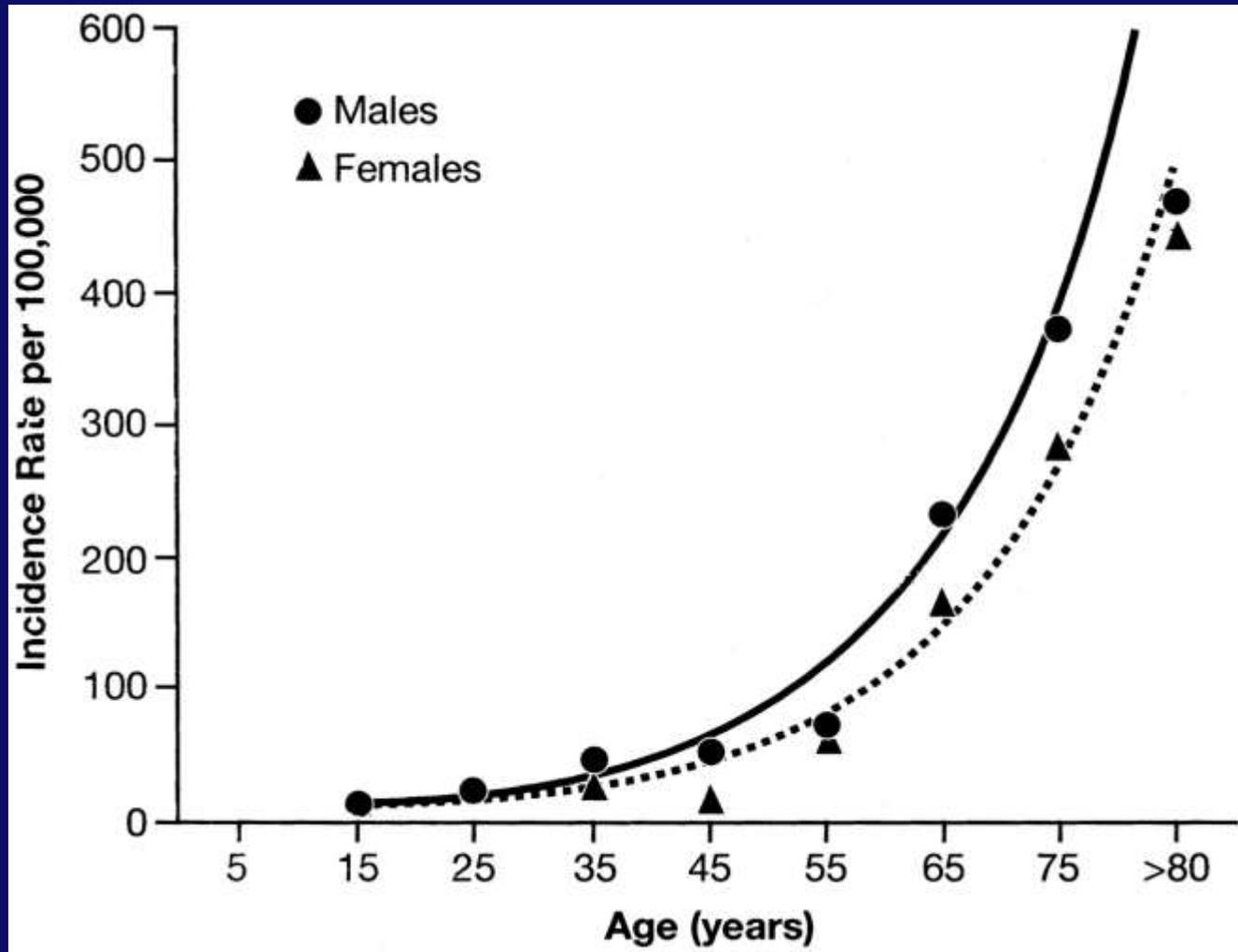


Bovill Ann Rev Physiol 2011



Moll and Mackman ATVB 2008

# Annual Incidence of VTE by Age and Sex



Richard H. White *Circulation*. 2003;107:1-4-1-8

# Cancer and Venous Thrombosis

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**Cancer patients have an a 4-7 fold increased risk for venous thrombosis – cancer-associated thrombosis (CAT)**

**~20% of patients with idiopathic VTE have cancer**

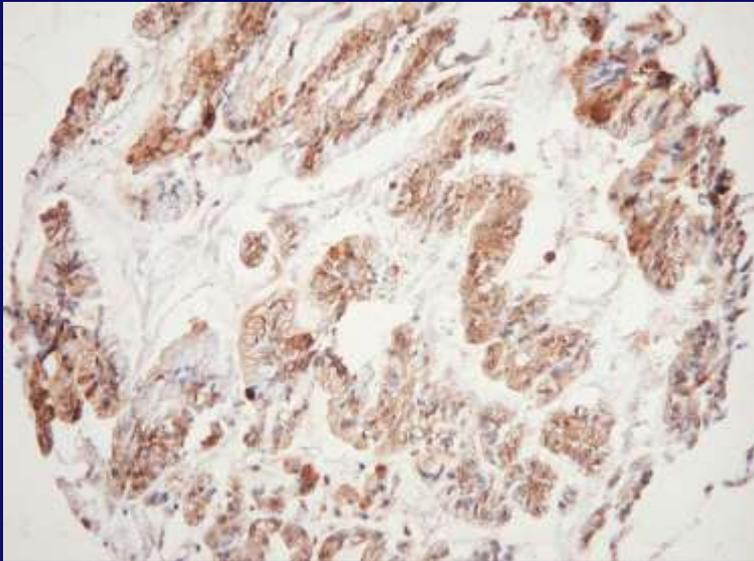
# Incidence of Symptomatic VTE within One Year of Cancer Diagnosis

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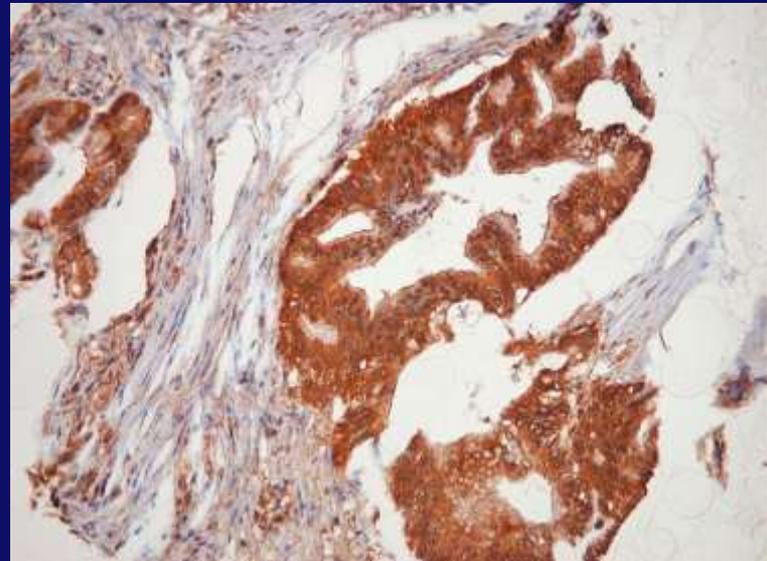
■ Pancreas	5.3- 26%	High
■ Brain	1.6- 26%	
■ Lung	1.6- 13.6%	Medium
■ Colon	3.1- 10.3%	
■ Breast	0.4- 8.1%	Low
■ Prostate	0.5- 1.4%	

# TF Expression in Pancreatic Cancer

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**Grade 1**



**Grade 3**

Khorana et al Clin Cancer Res 2007

# **Cancer Cells and Procoagulant Microvesicles**

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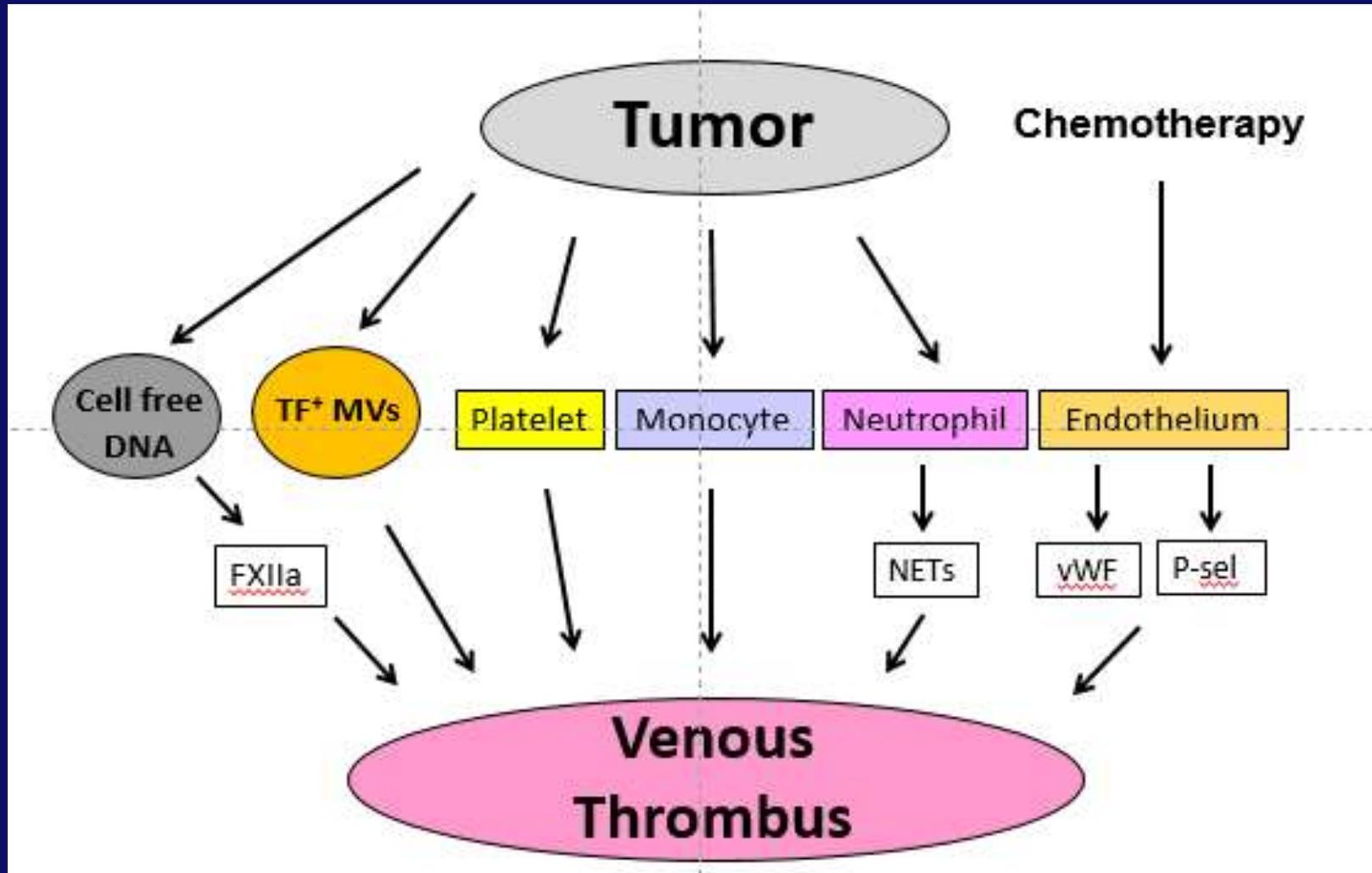
**Tumor cells shed plasma membrane vesicles when cultured in vitro. Shed vesicles carrying procoagulant activity may account for activation of the clotting system associated with malignancy**

**Dvorak HF et al Science 1981**

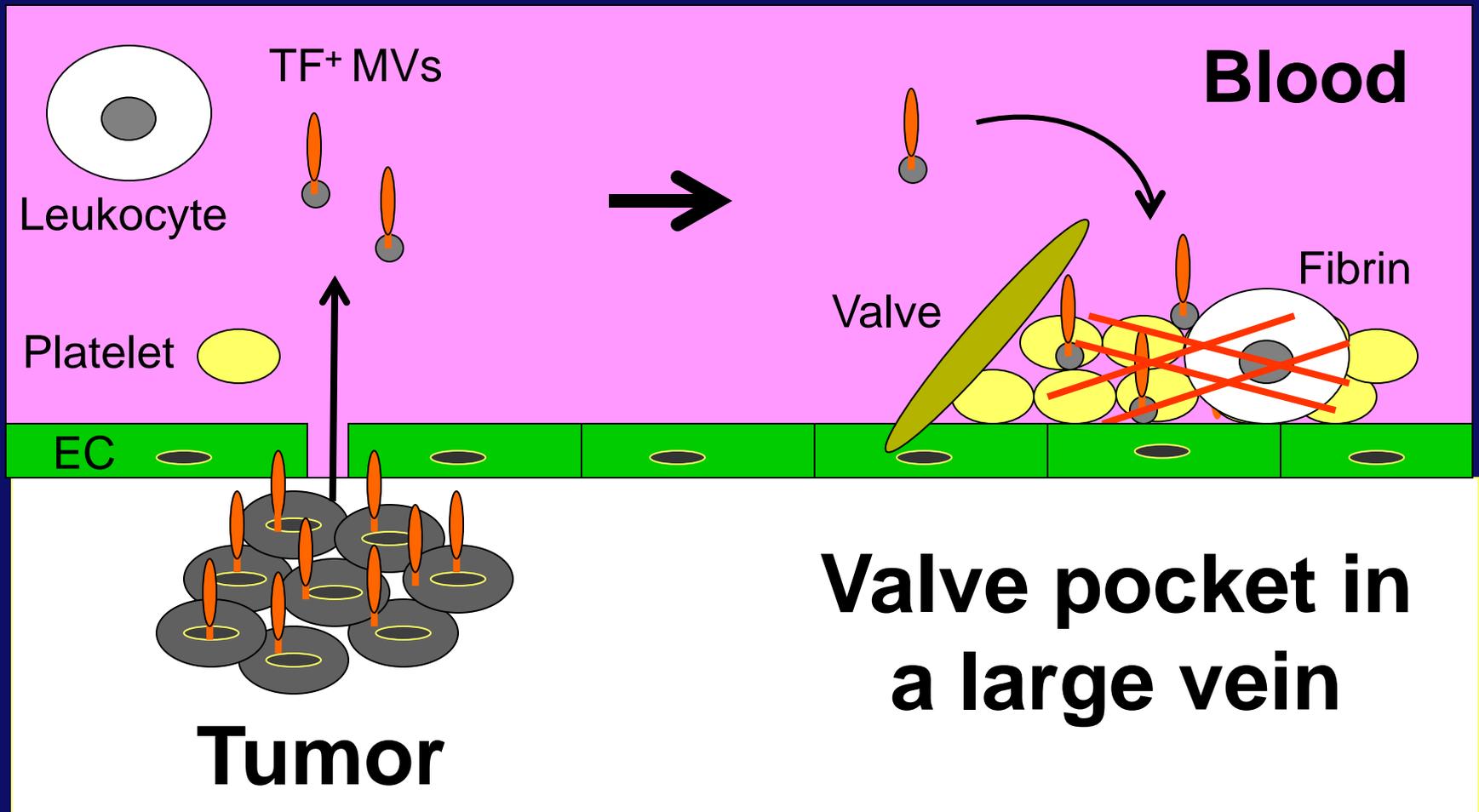
**The procoagulant activity of tumor-derived MVs is due to TF**

**Dvorak et al Cancer Res 1983; Bastida et al Blood 1984; Yu and Rak JTH 2004**

# Mechanisms of Cancer-associated Thrombosis



# Hypothesis: Tumor-derived TF<sup>+</sup> MVs Trigger Venous Thrombosis in Cancer Patients



# Mouse Studies

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## Hypothesis

**Tumor-derived, TF<sup>+</sup> MVs activate coagulation and increase clot size in a venous thrombosis model**

# Summary of Results with TF and Mouse Tumor Models

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Human CRC and pancreatic tumors in mice release TF+ MVs into the blood. Yu et al Blood 2005; Davila et al JTH 2008; Wang J-G et al Blood 2012.

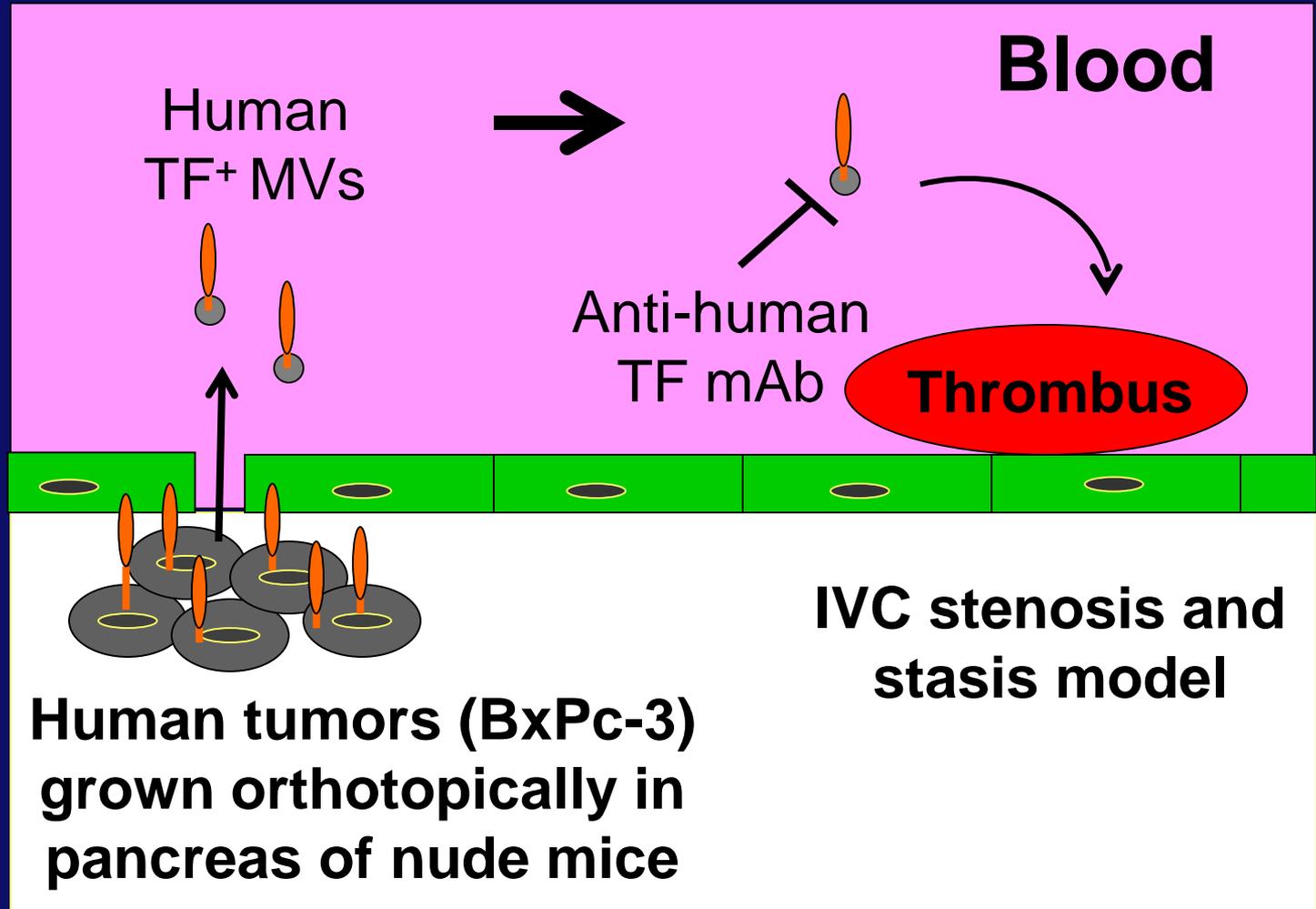
Mice with SQ murine pancreatic tumors have increased thrombosis. Human tumors in mice release MVs containing TF into the blood. Thomas C et al JEM 2009, Thomas C et al JTH 2015.

**Do tumors enhance  
clot size in mice?**

# Model to Study the Role of Tumor-derived, TF+ MVs in Venous Thrombosis in Mice



Julia Geddings

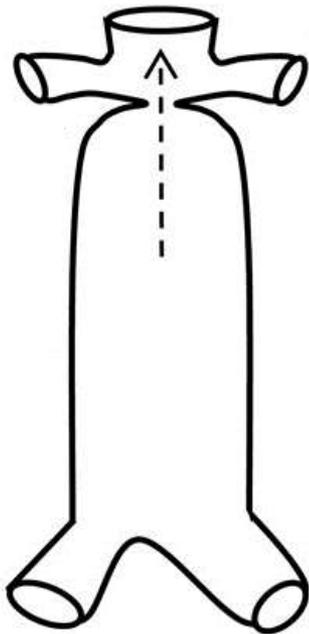


Yohei Hisada

# Mouse Venous Thrombosis Models: Infrarenal Vena Cava Stenosis and Stasis

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## Stenosis

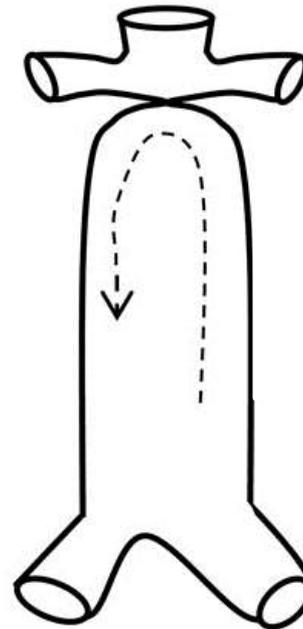


Renal  
Veins

Infrarenal  
IVC

Iliac  
Veins

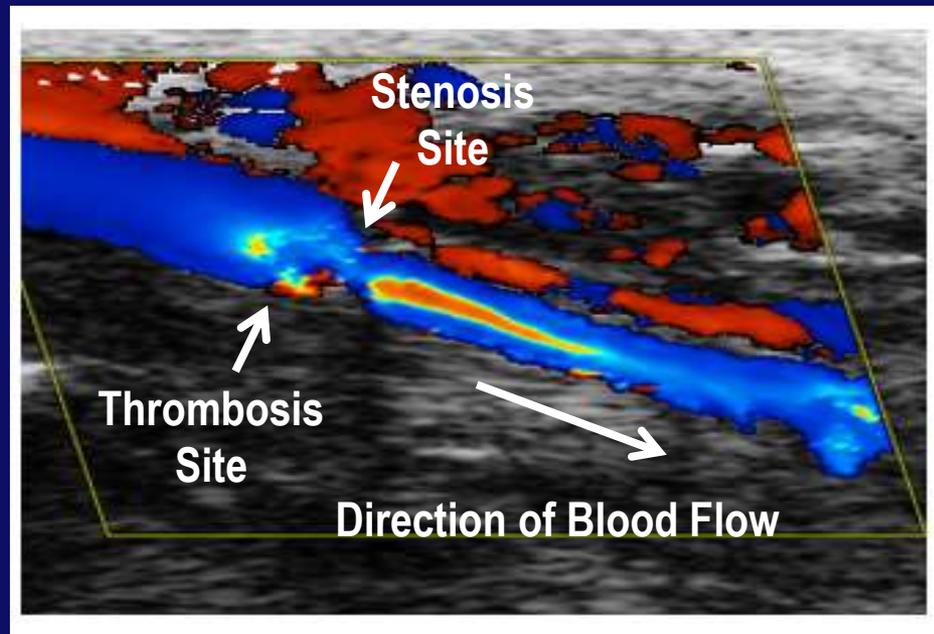
## Stasis



# Kinetic Analysis of Thrombosis Formation

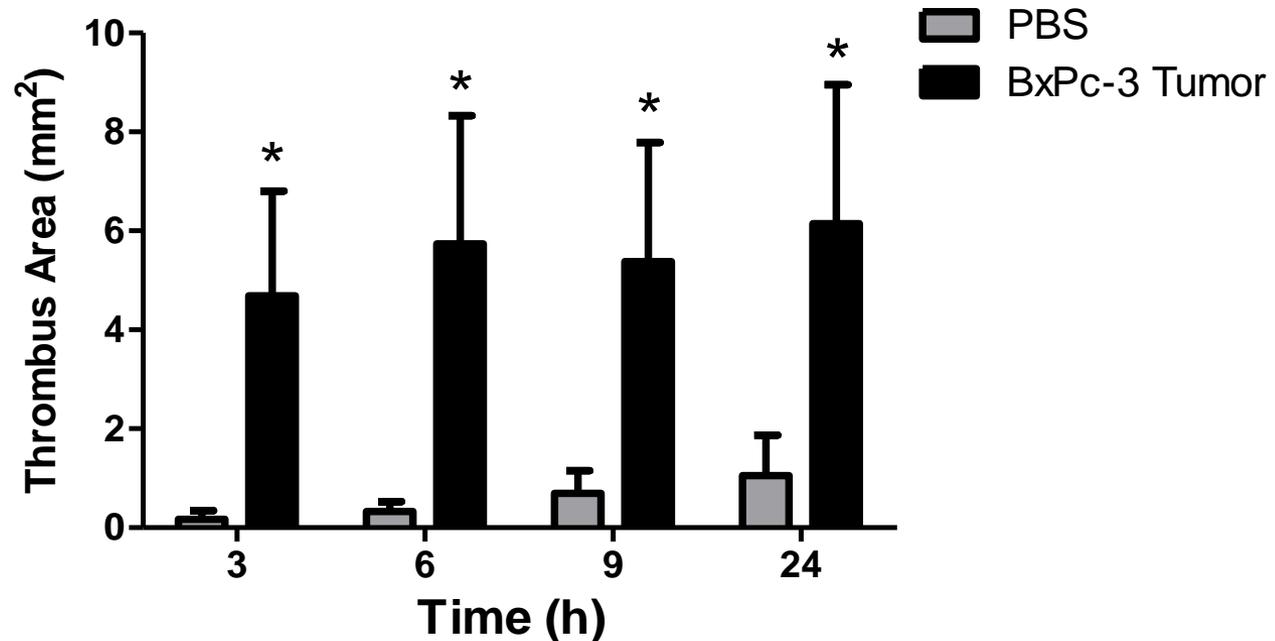
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## Color Doppler



Geddings J et al. JTH 2014

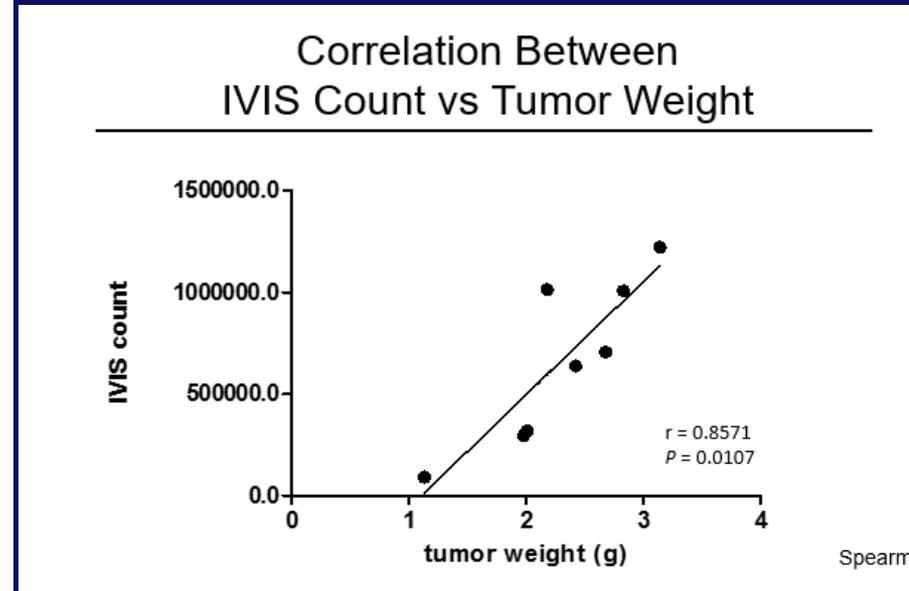
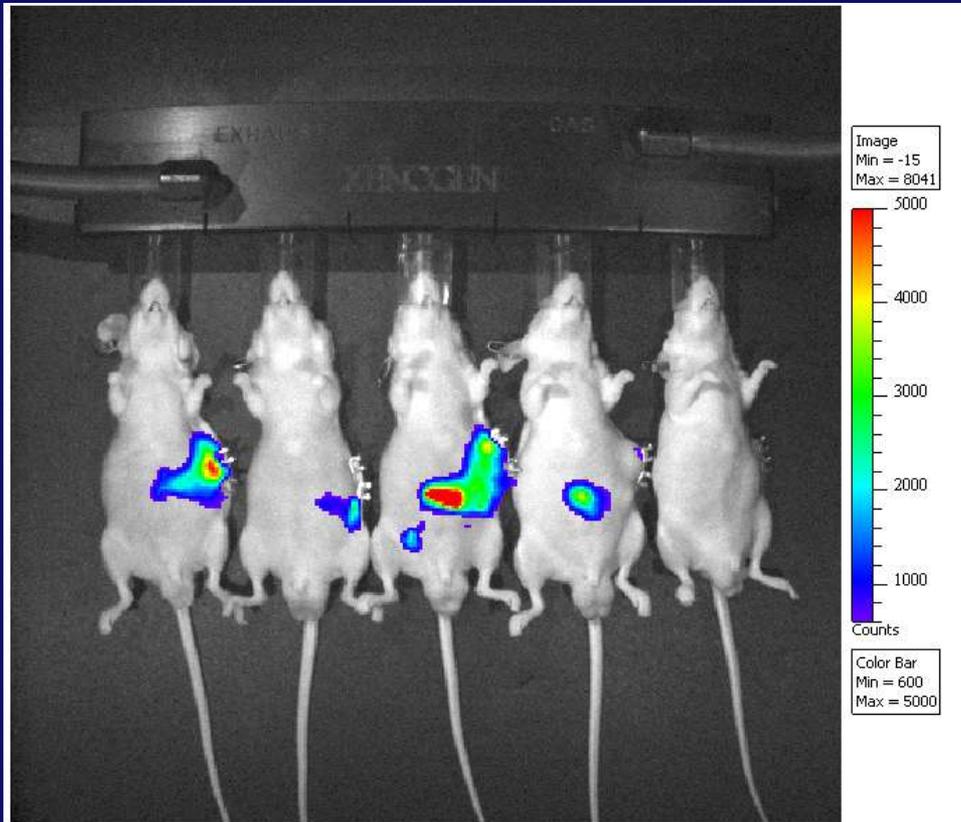
# Human Pancreatic Tumors increase the Incidence of Thrombosis in the IVC Stenosis Model



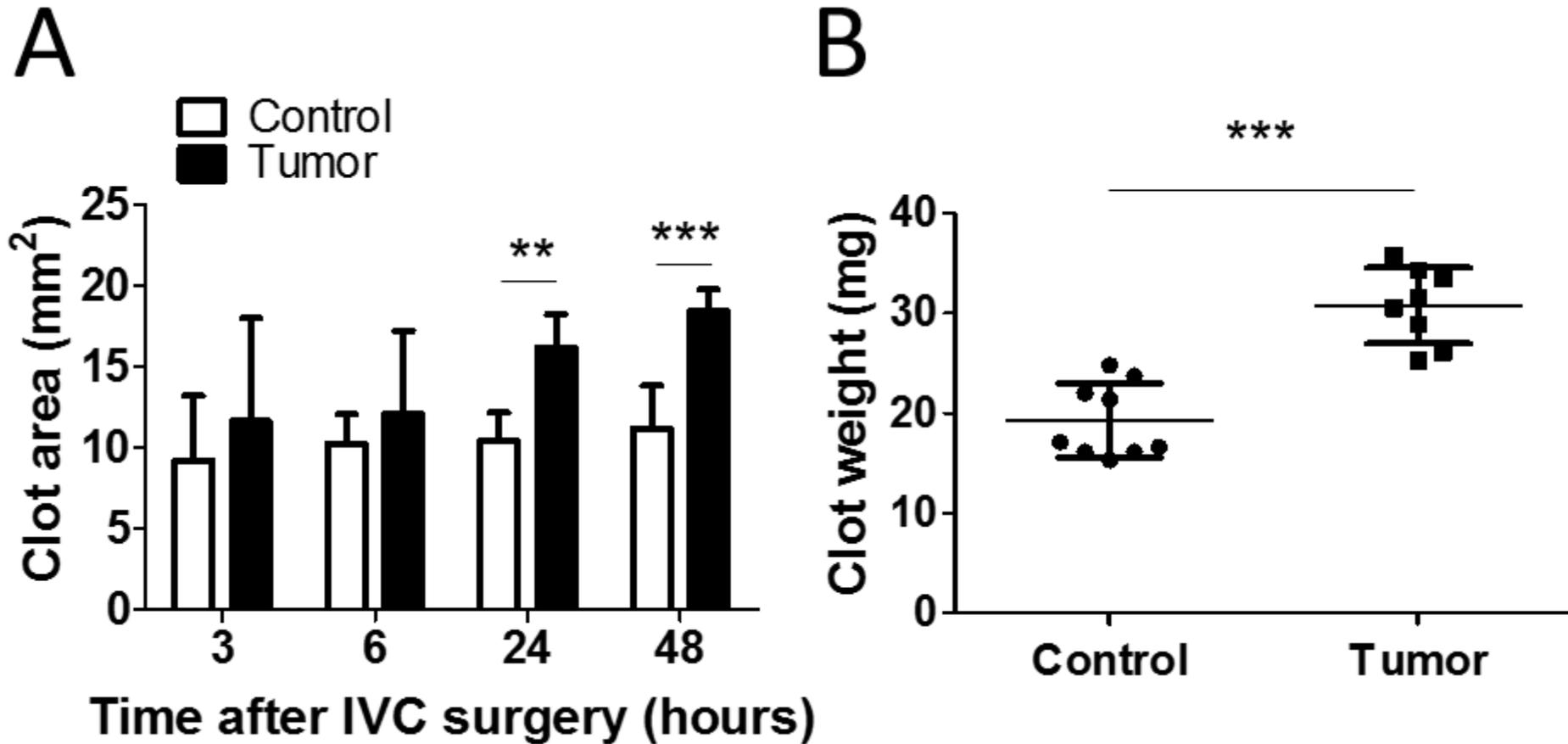
Thrombus Incidence

Time (h)	3	6	9	24
PBS	17%	33%	33%	33%
BxPc-3 Tumor	50%	50%	50%	50%

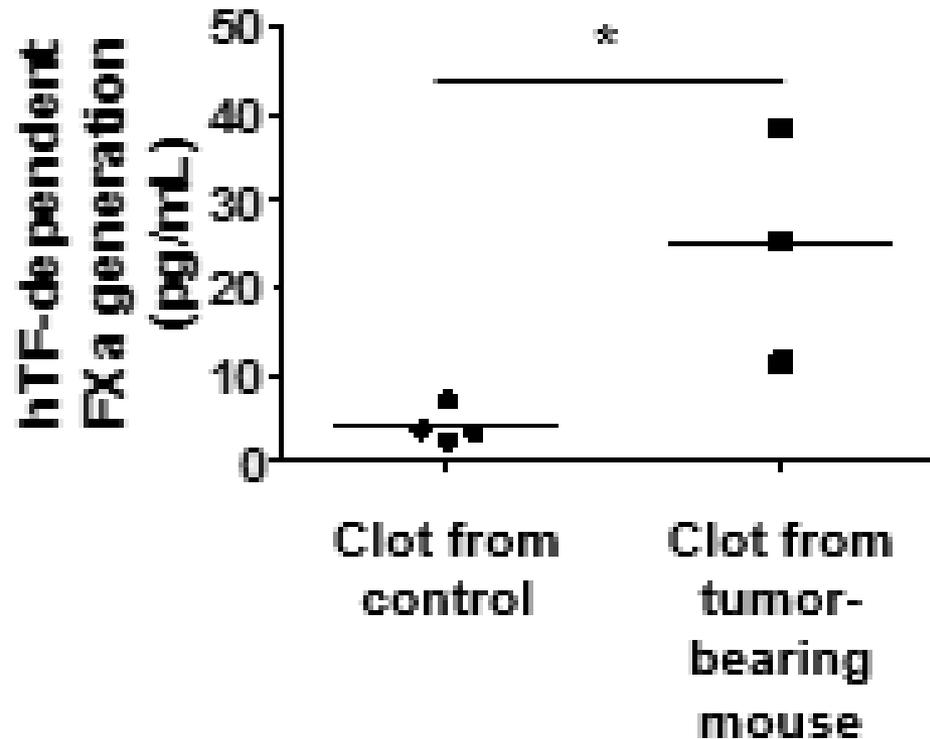
# Imaging of BxPc-3-luciferase Tumors in the Pancreas of Mice



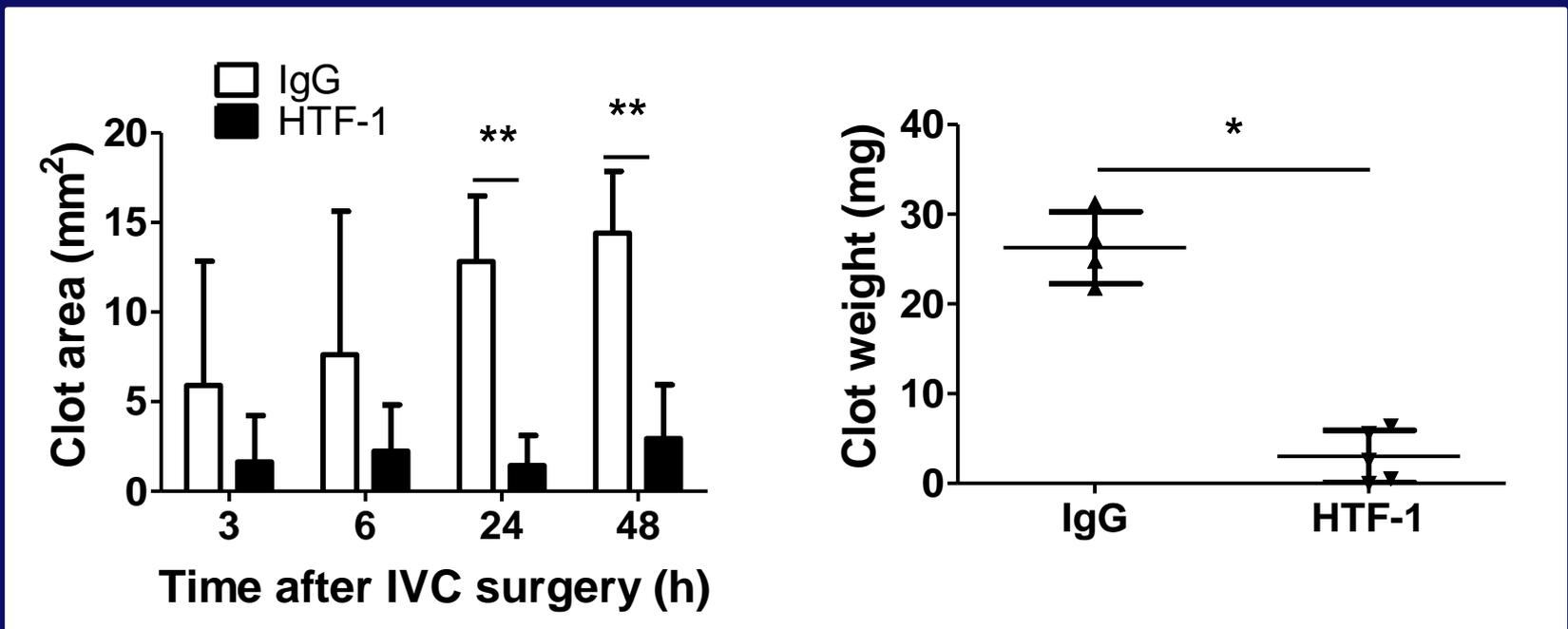
# Human Pancreatic Tumors Increase Clot Size in Nude Mice



# Clots from Tumor-bearing Mice Contain Human TF



# Inhibition of Human TF Reduces Clot Size in Tumor-bearing Mice



HTF-1 does not affect thrombosis in control mice

Hisada Y et al JTH 2017

# Conclusion

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**Tumor-derived TF<sup>+</sup> MVs  
enhance venous  
thrombosis in mice with  
pancreatic tumors**

# Clinical Studies

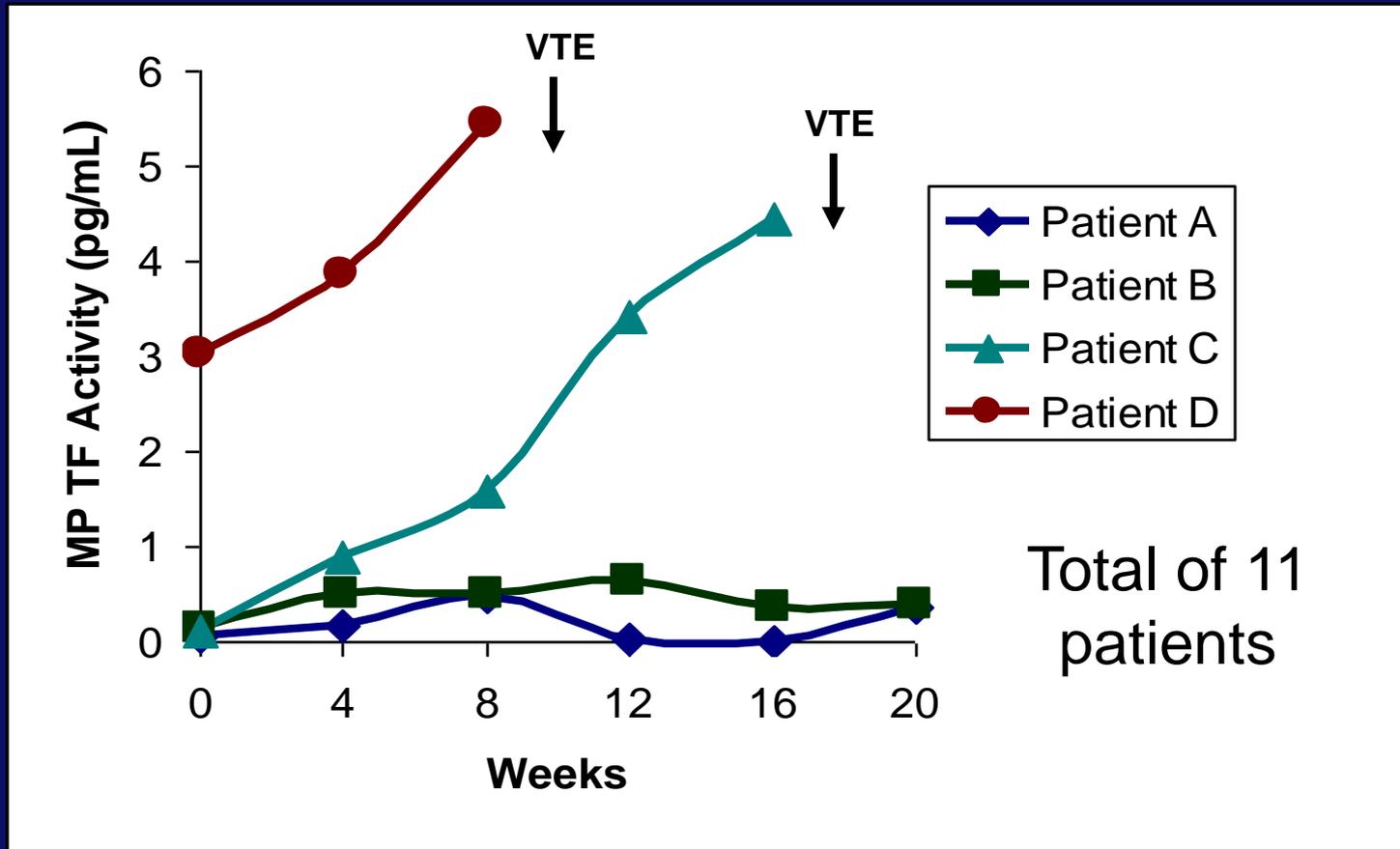
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## Hypothesis

**MV TF activity may be a useful biomarker to identify cancer patients at risk for venous thrombosis**

**Perform prospective studies**

# Levels of MV TF Activity Increase in Pancreatic Cancer Patients before VTE



# Association of MV TF Activity and VTE in Pancreatic Cancer Patients

---

Study	Patients	Time of Follow-up	Association between MV TF and VTE?
Khorana, 2008	2/9 patients developed VTE	5 months	Yes
Van Doornaal, 2012	3/13 patients developed VTE	6 months	Yes
Thaler, 2012	12/60 patients developed VTE	2 years	Weak
Bharthuar, 2013	*52/117 patients developed VTE	6 months	Yes
Van Es, submitted	9/89 patients developed VTE	6 months	Weak
Khorana, submitted	7/39 patients developed VTE	3 months	Yes
Ilich, In preparation	4/22 patients developed VTE	6 months	Yes

\*Pancreaticobiliary cancer patients

# Levels of MV TF Activity in 4 Cancers with a High Rate of VTE: Vienna CATS

	VTE	MV TF activity	Association with VTE
Pancreatic	12/60 (20%)	0.1 (0.04-0.19)	Yes
Stomach	6/43 (14%)	0.07 (0.0-0.17)	No
Colorectal	12/126 (10%)	0.05 (0.01-0.15)	No
Brain	19/119 (16%)	0.04 (0.0-0.08)	No

# Conclusion

---

**MV TF activity may be a useful biomarker to identify pancreatic cancer patients at risk for VTE**

# General Conclusions

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- TF provides a hemostatic envelope around blood vessels and at organ and body surfaces.
- TF expression provides additional hemostatic protection to vital organs, such as the heart, lung and brain.
- TF is expressed in atherosclerotic plaques and contributes to arterial thrombosis after plaque rupture.
- In pancreatic cancer, levels of tumor-derived circulating TF+ MVs are associated with VTE.

# Mackman Lab 2017



— TREC —  
K.G. JEBSEN THROMBOSIS  
RESEARCH AND EXPERTISE CENTER

TraCS

# Re-balancing the Clotting Cascade in Mice

---

Can we rescue the lethality of embryos lacking different anticoagulants with low levels of TF?

Cross =

$TFPI^{+/-}, mTF^{+/-}, hTF^{+}$  x  $TFPI^{+/-}, mTF^{+/-}, hTF^{+}$

Mice

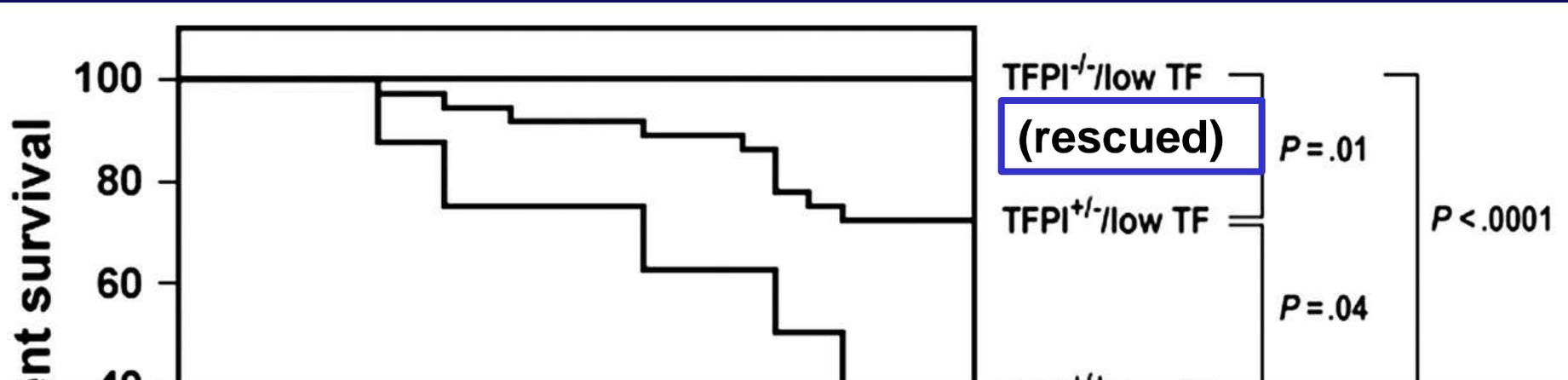
Low TF,  $TFPI^{+/+}$

Low TF,  $TFPI^{+/-}$

**1 slide**

Low TF,  $TFPI^{-/-}$  (rescued mice)

# Balancing TF and TFPI in Mice

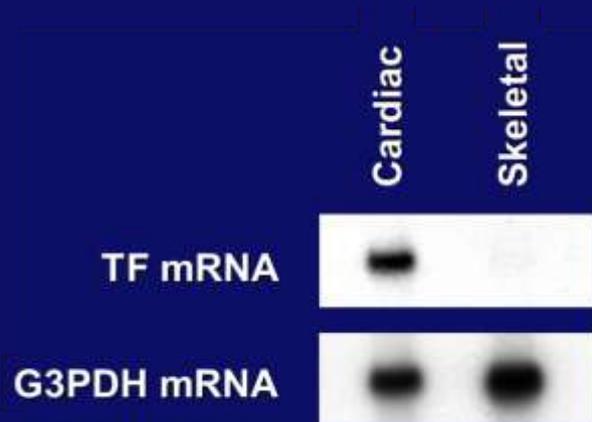


1/ TFPI<sup>-/-</sup> embryos are rescued by reducing the level of TF.

2/ Fatal bleeding in low TF mice is rescued by reducing the level of TFPI.

# TF Expression in Mouse and Human Cardiac and Skeletal Muscle

## Mouse



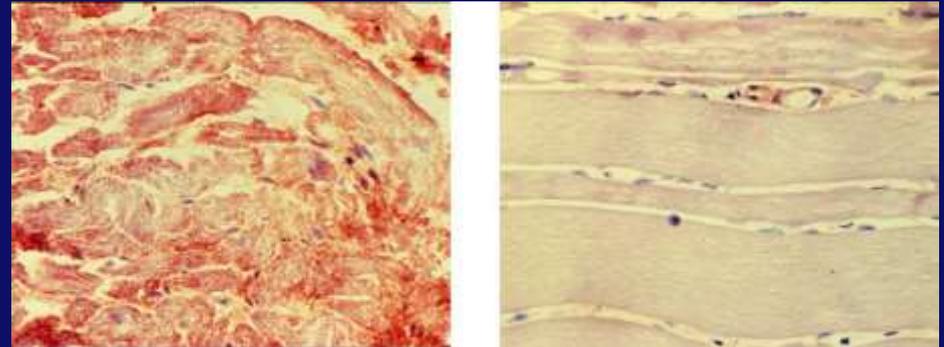
Mackman et al AJP 1993

**move**

## Human

Cardiac  
muscle

Skeletal  
muscle



Drake, Morrissey & Edgington (1989)  
A J P 134:1087-1097

from Jim Morrissey, University of Illinois

# Hemostatic Defects in Low TF Mice

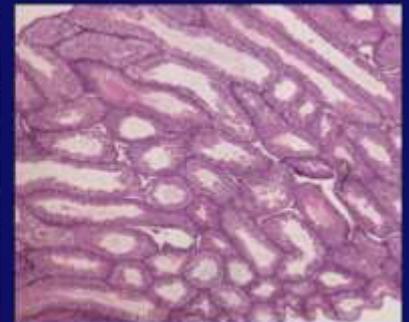
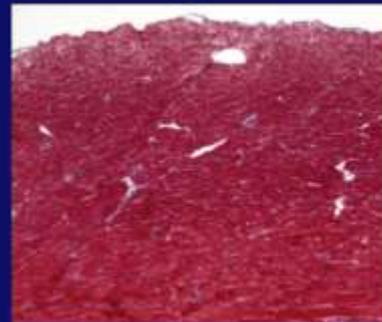
Placenta

Lung

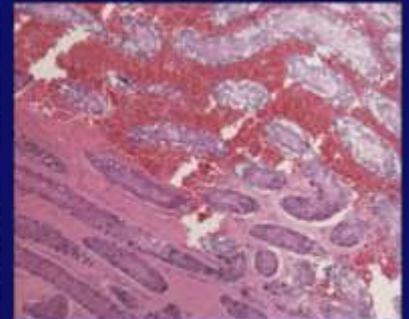
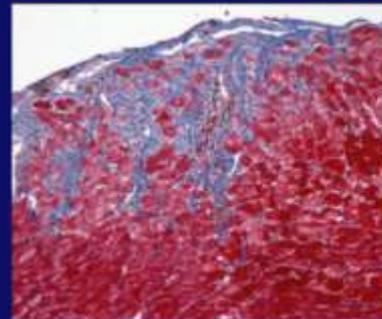
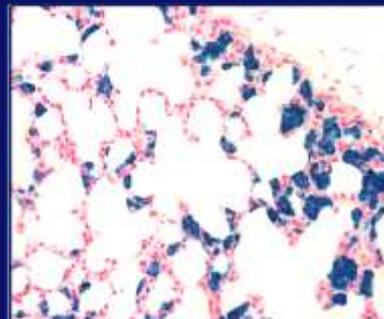
Heart

Testis

WT



Low  
TF



Erlich et al.

Pedersen et al.  
Blood 2005

Pawlinski et al.  
PNAS 2002

Mackman  
JTH 2007

**move**

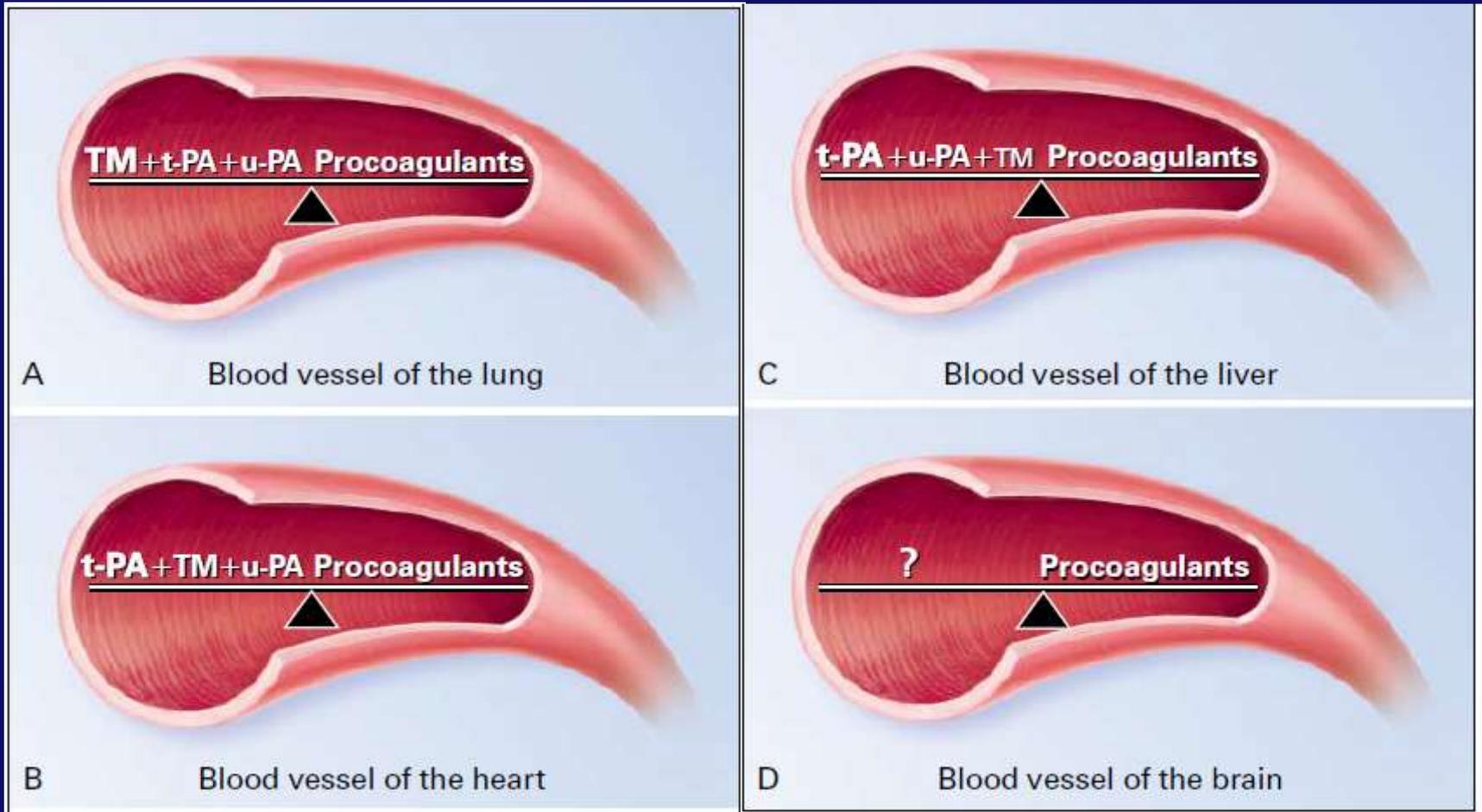
# Role of Uterine TF in Hemostasis During Pregnancy

Breeding strategy	Level of TF expression		Fatal post-partum hemorrhage
	Mother	Embryo	

Low levels of TF in the embryo leads to fatal post-partum hemorrhage, whereas low levels of TF in the mother and embryo leads to fatal mid-gestational hemorrhage.

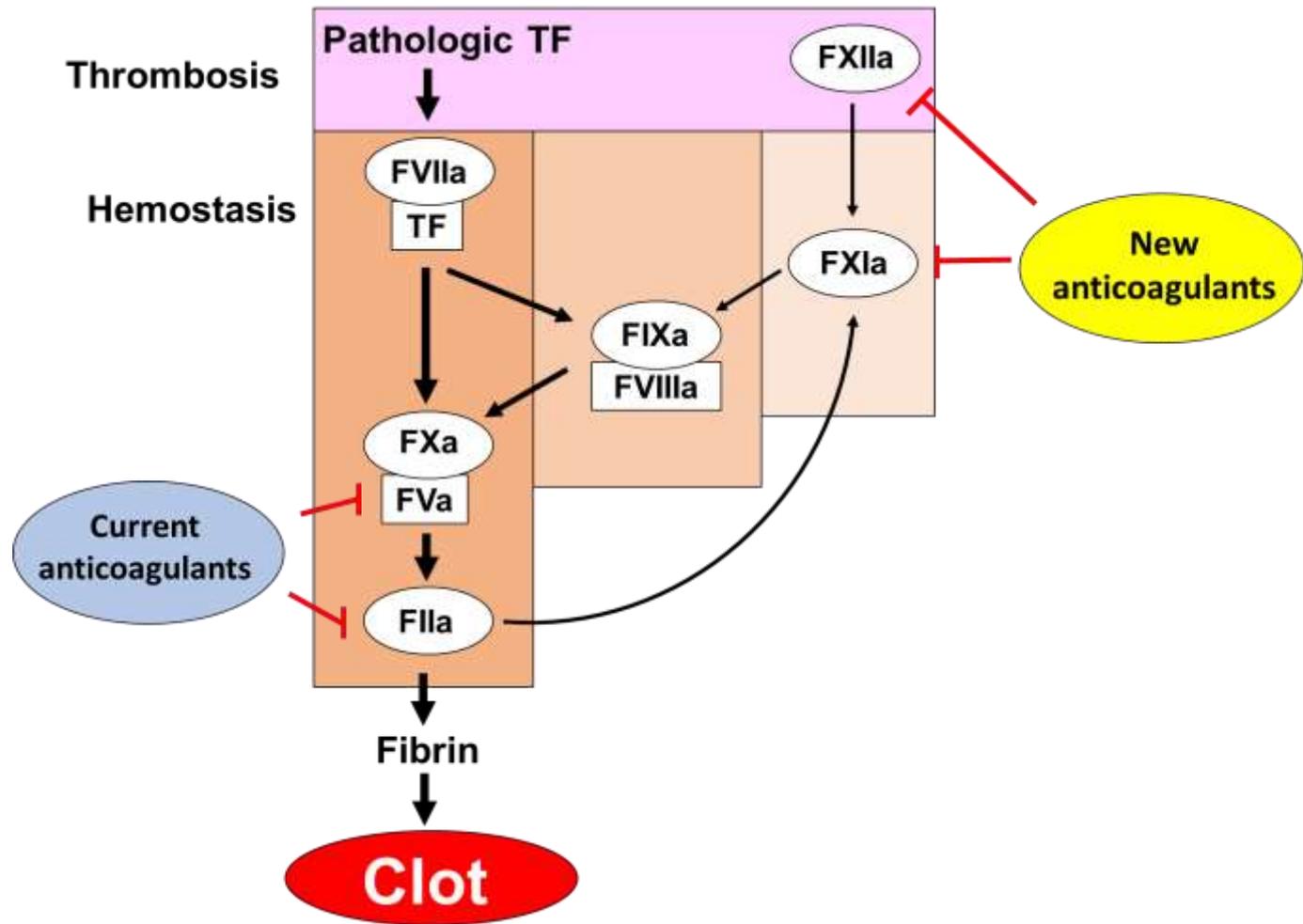
**F<sup>move</sup> IX<sup>-/-</sup> have normal pregnancies**

# Vascular Bed-specific Hemostasis



Aird and Rosenberg NEJM 1999

# Development of New Anticoagulant Drugs





## Factor XI Antisense Oligonucleotide for Prevention of Venous Thrombosis

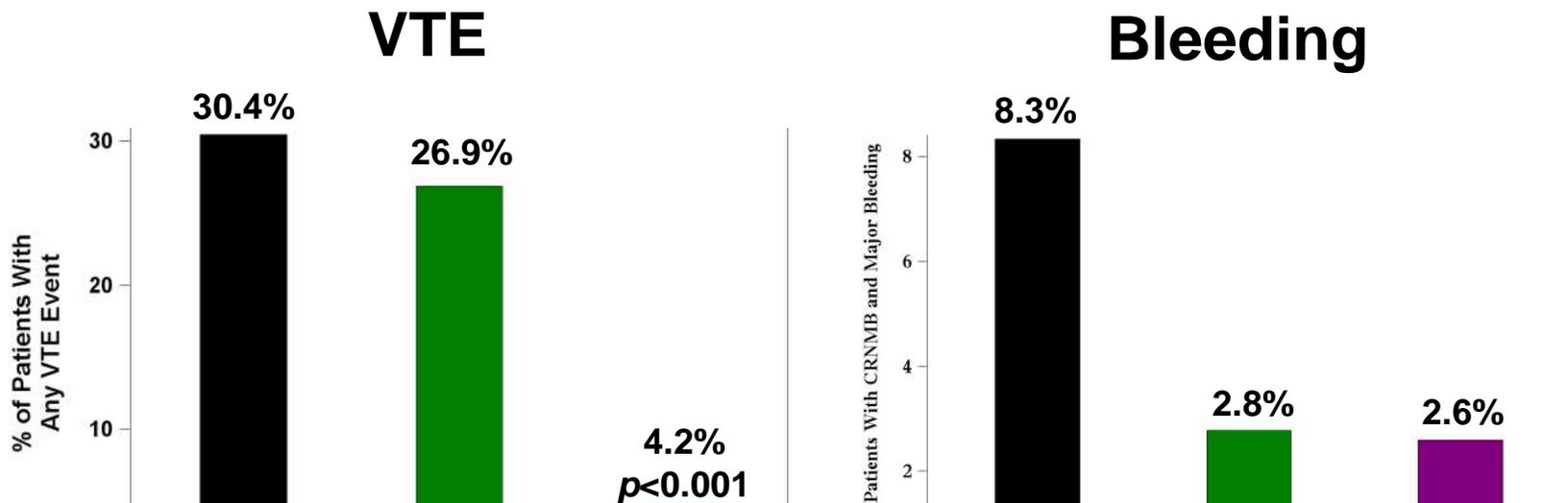
Harry R. Büller, M.D., Claudette Bethune, Ph.D., Sanjay Bhanot, M.D., Ph.D, David Gailani, M.D., Brett P. Monia, Ph.D., Gary E. Raskob, Ph.D., Annelise Segers, M.D., Peter Verhamme, M.D., and Jeffrey I. Weitz, M.D.

2015;372:232-240

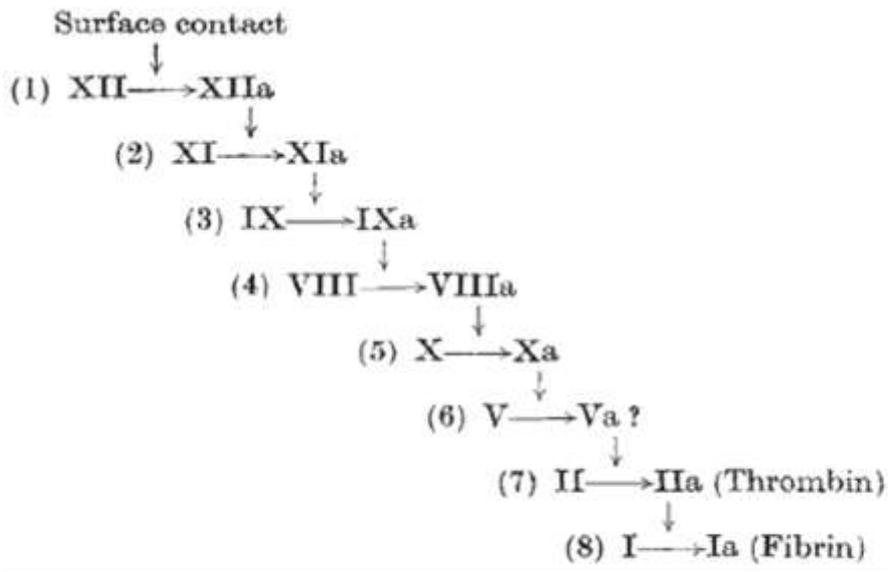
- Effect of decreased levels of FXI on VTE in pts undergoing total knee replacement
- Two doses of a FXI ASO were used: 200 mg ~41% FXI; 300 mg ~22% FXI.
- Enoxaparin was used as a control.

Slides from Dr. J. Weitz

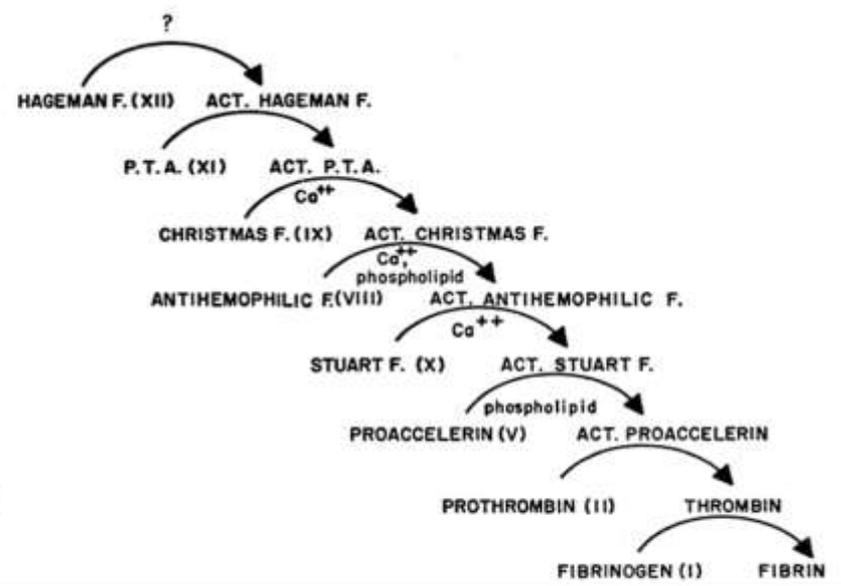
# Effect of Reducing FXI Levels on VTE and Bleeding after Total Knee Replacement



**FXI appears to be a promising target to reduce VTE without increasing bleeding**



Macfarlane Nature 1964



Davie Science 1964

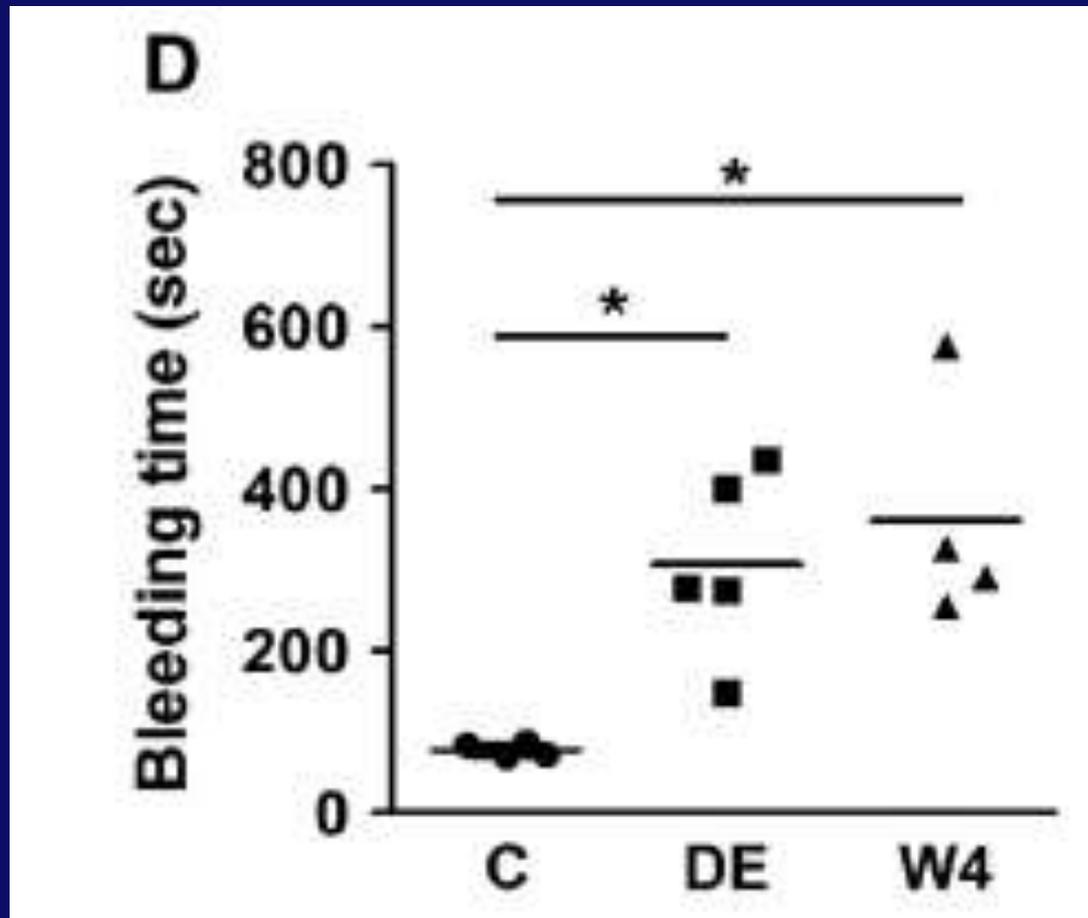
**Determine the effect of  
anticoagulants on alveolar  
hemorrhage in mice after  
influenza A infection**



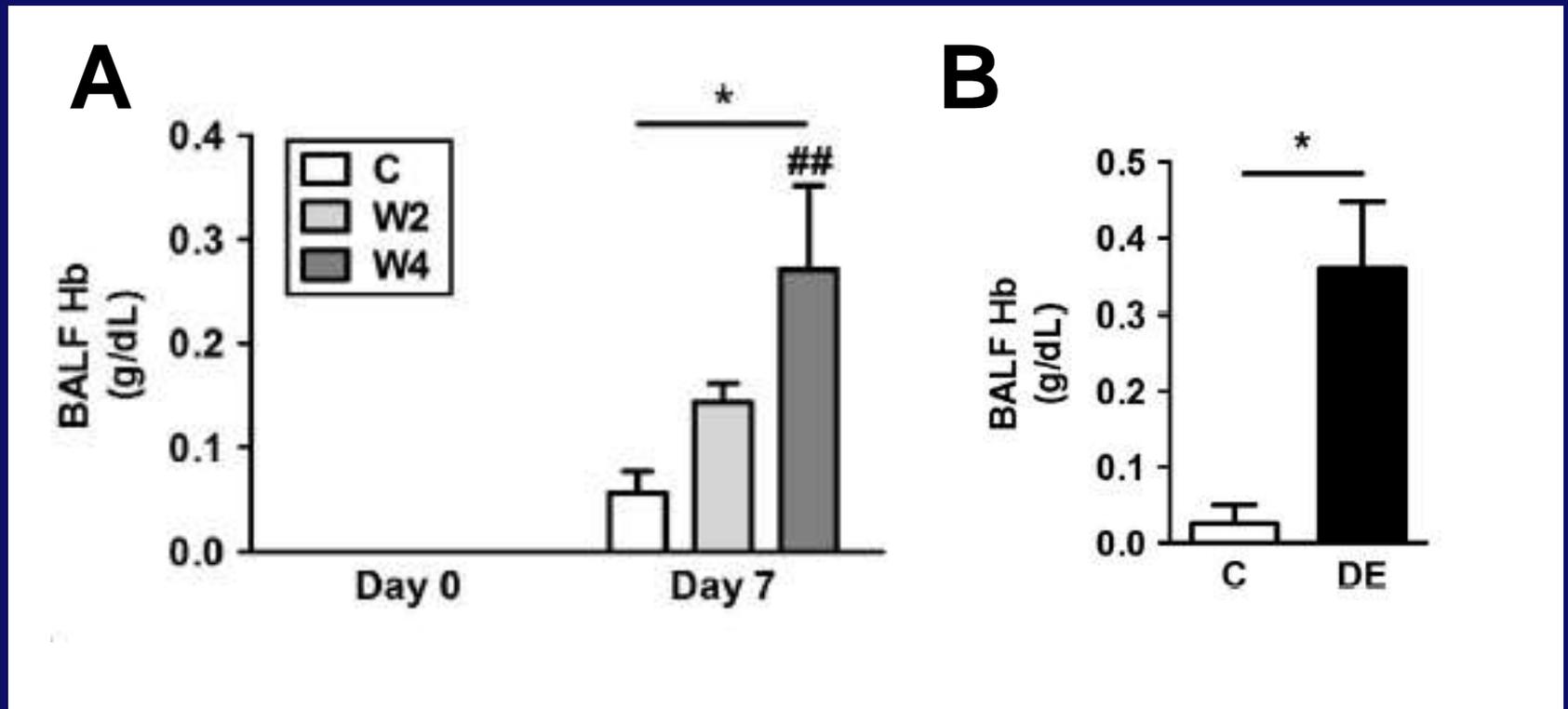
**Kohei Tatsumi**

# Anticoagulating Mice with Dabigatran or Warfarin

---



# Warfarin and Dabigatran Increase Lung Hemorrhage After H1N1 Infection



# Conclusion

---

**Anticoagulants may increase  
alveolar hemorrhage and  
morbidity in patients infected with  
influenza A**

**Use of anticoagulants in the US**

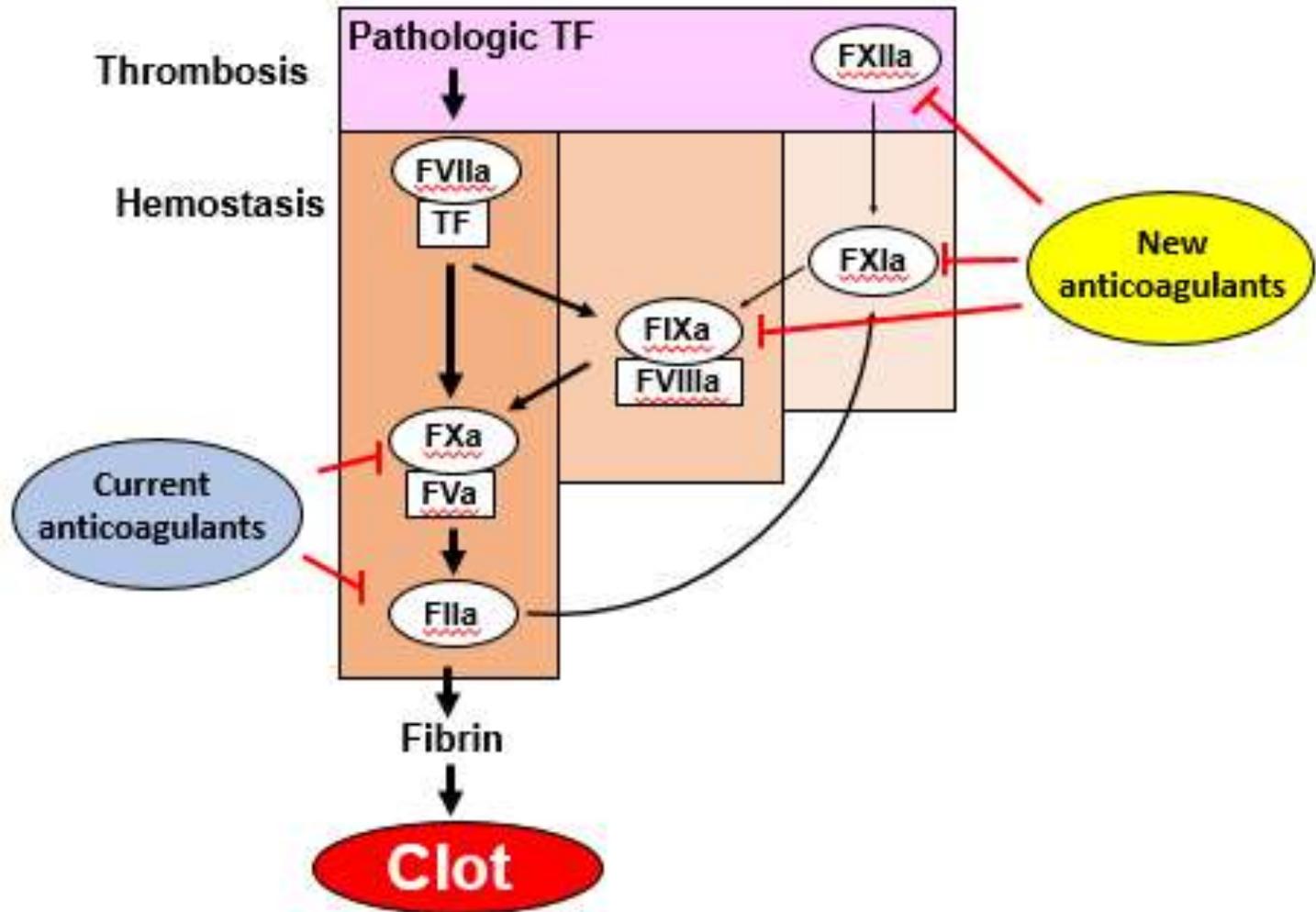
**<1% under 65**

**5% 65-74**

**10% >75**

**Medical Expenditure Panel Summary Oct 2009**

# Development of New Anticoagulant Drugs



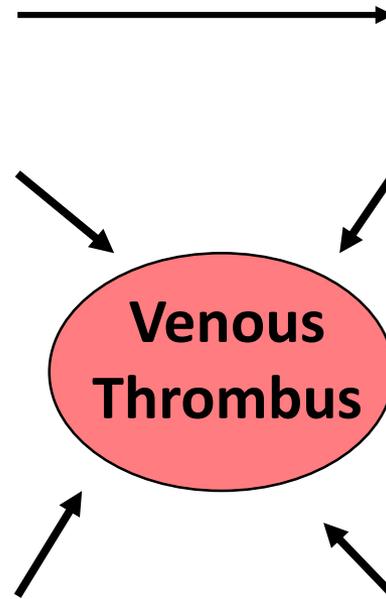
# Risk Factor for Cancer-associated Thrombosis

## Tumor characteristics

Site - high risk = pancreatic, brain, lung, ovarian, lymphoma, myeloma, kidney, stomach, bone  
- low risk = breast, prostate  
Stage - localized  
- metastatic

## Treatment

Chemotherapy  
Radiotherapy  
Surgery  
CVC  
Hormone therapy  
Erythropoiesis stimulating agents  
Anti-angiogenic agents



## Blood cells

Platelet count  
Leukocyte count

## Hemostatic System

Prothrombotic variants  
Anticoagulant deficiencies

## Patient characteristics

History of VTE  
Age  
Immobilization  
Obesity

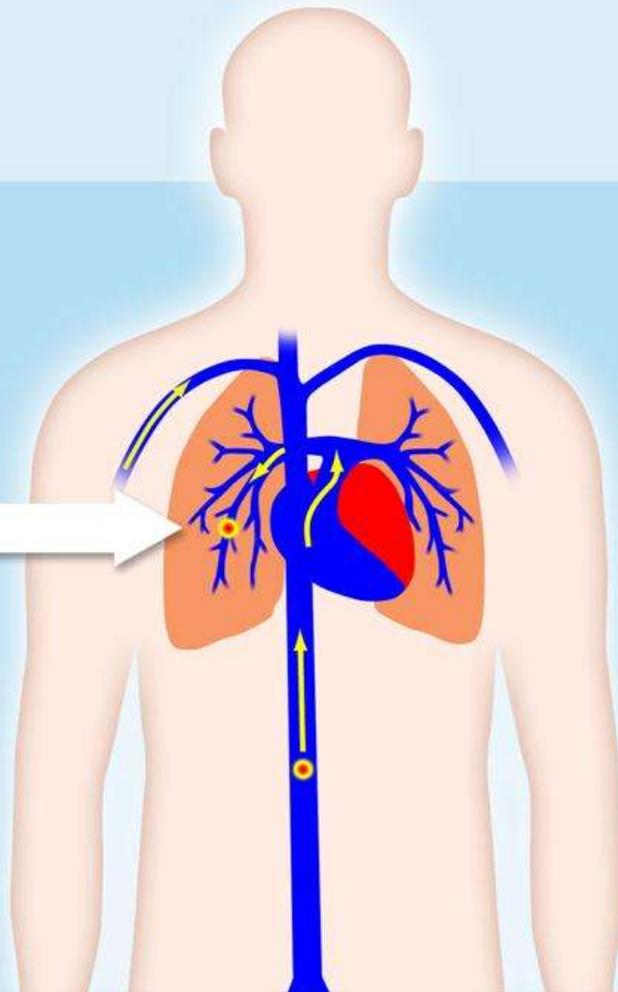
# Venous Clots

## Pulmonary embolism (=PE)

- shortness of breath
- chest pain
- cough
- bloody sputum

## Deep vein thrombosis (=DVT)

• swelling

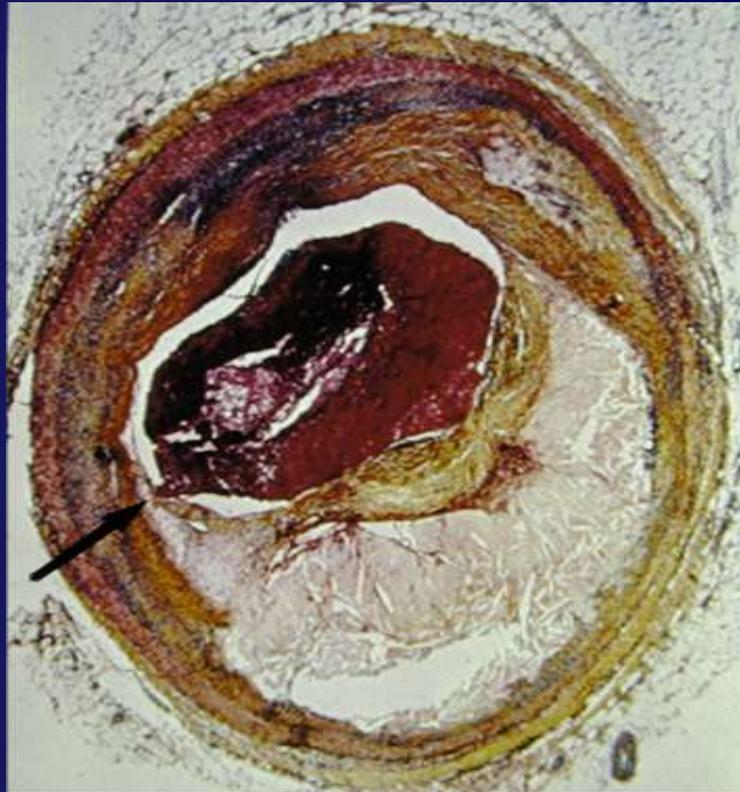


**Venous thromboembolism  
(VTE) = DVT + PE**

Slide from Dr. Stephan Moll [clotconnect.org](http://clotconnect.org)

# Arterial Thrombosis

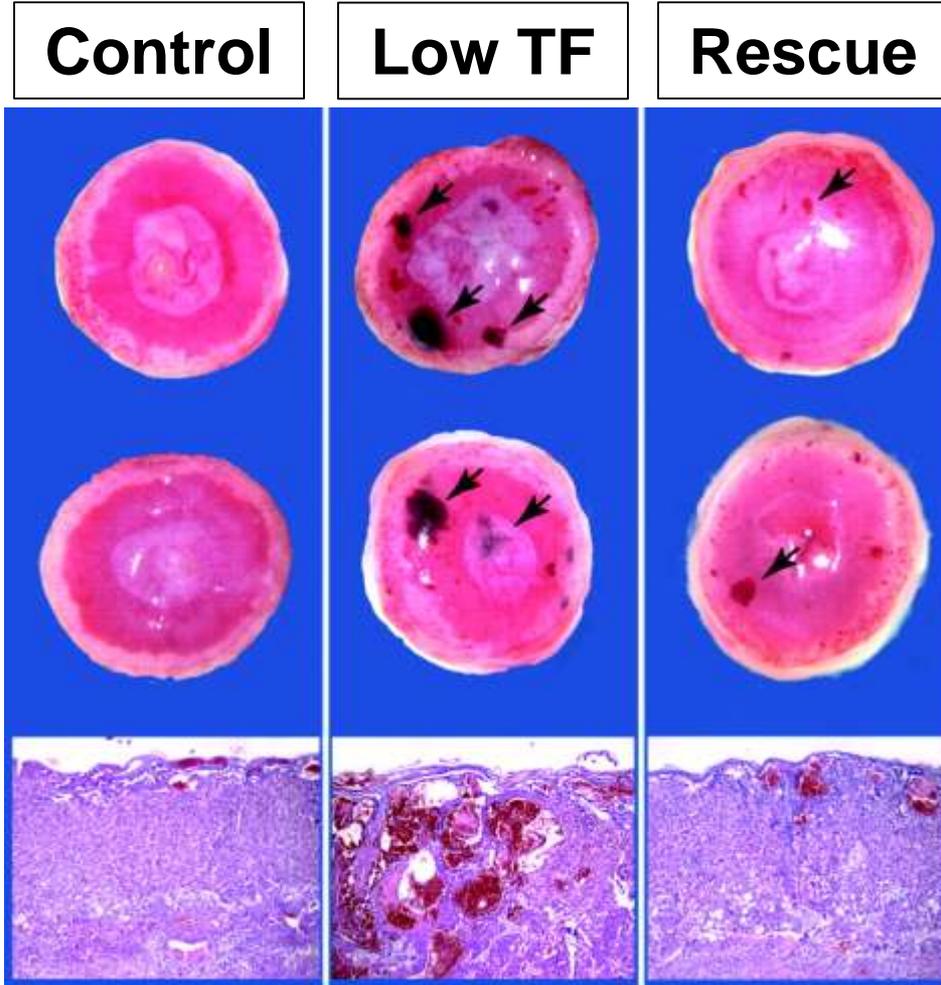
---



**Thrombosis in a human  
coronary artery**

**provided by Dr. Taubman**

# Reduced Blood Pools in “Rescued” Placentas



Pedersen B et al Blood 2005

# History of Tissue Factor-Protein

---

- **Purification of human TF** (Broze G et al JBC 1985).
- **Crystal structure** (Harlos K et al Nature 1994).

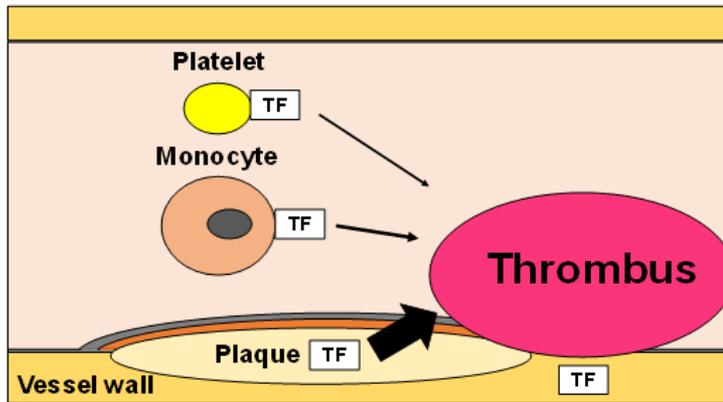
# History of Tissue Factor-Gene

---

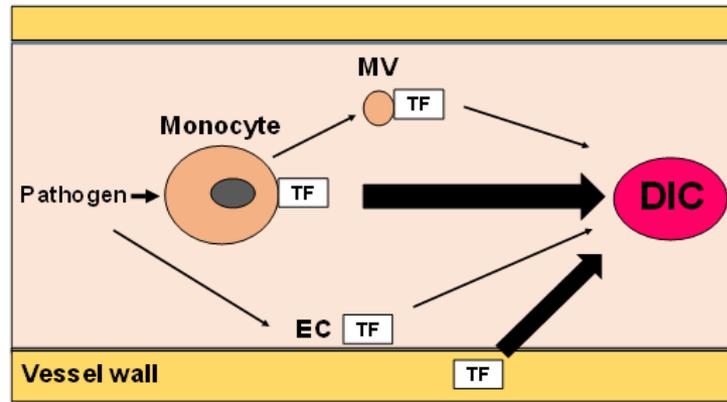
- **Human TF cDNA** (Spicer E et al PNAS 1987; Morrissey J et al Cell 1987; Scarpati E et al Biochem 1987; Fisher K et al Thromb Res 1987).
- **Human TF gene** (Mackman N et al Biochem 1989).
- **Promoter characterization** (Mackman N et al JEM 1991).
- **KO of the mouse TF gene** (Bugge T et al PNAS 1996; Carmeliet P et al Nature 1996; Toomey J et al Blood, 1996)

# Roles of TF in Thrombosis and Disseminated Intravascular Coagulation

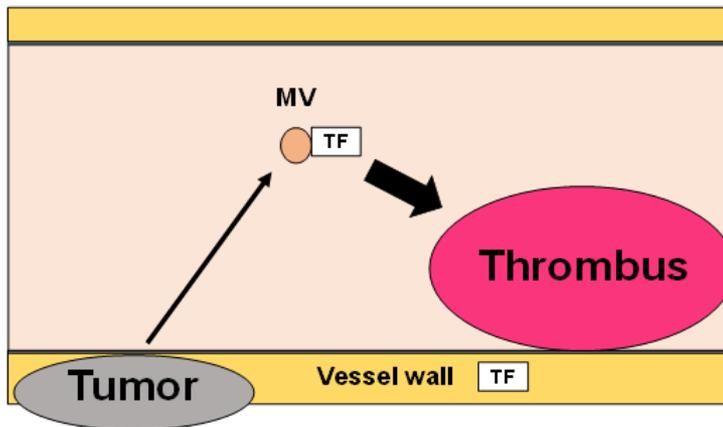
A. Atherosclerosis



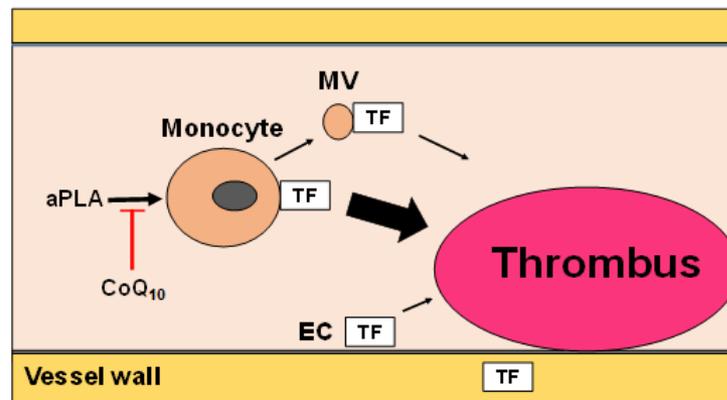
B. Endotoxemia/sepsis



C. Pancreatic cancer

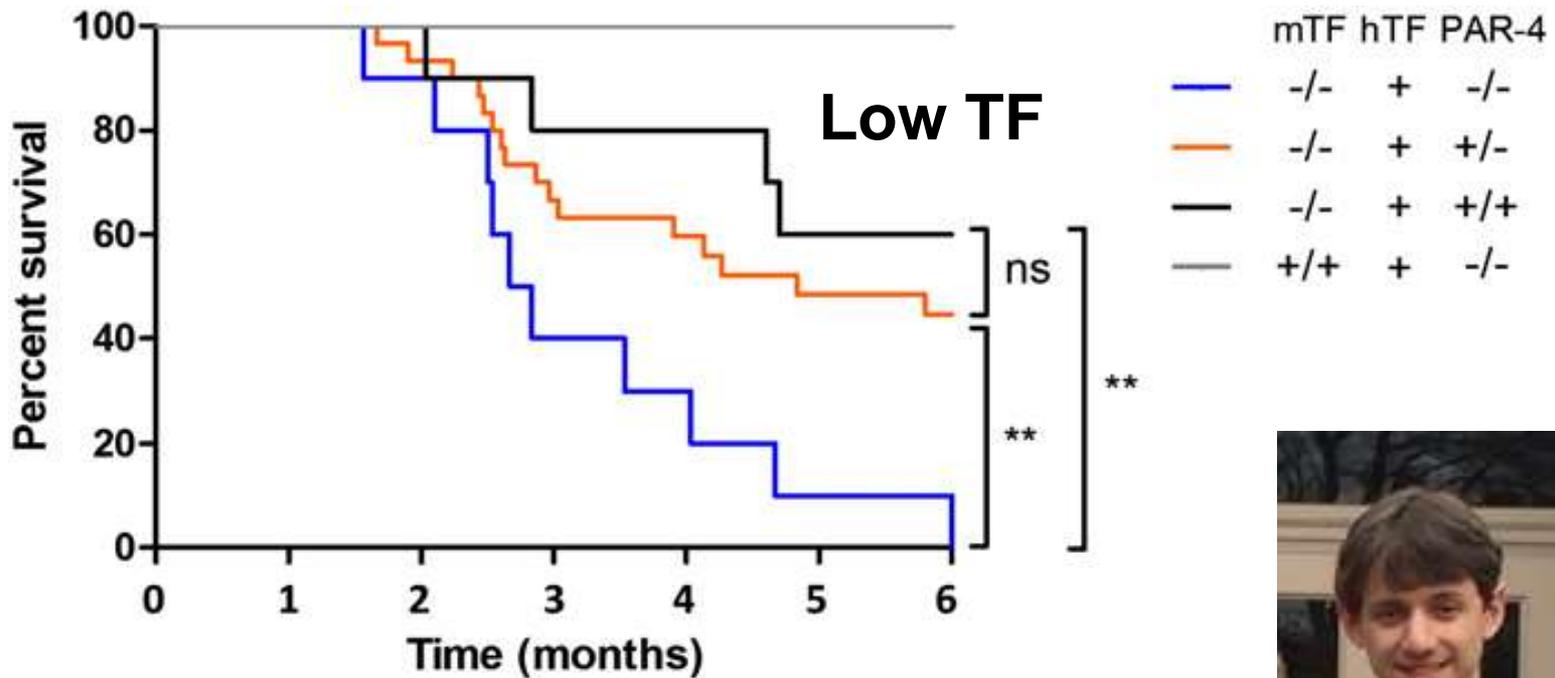


D. Antiphospholipid antibody syndrome

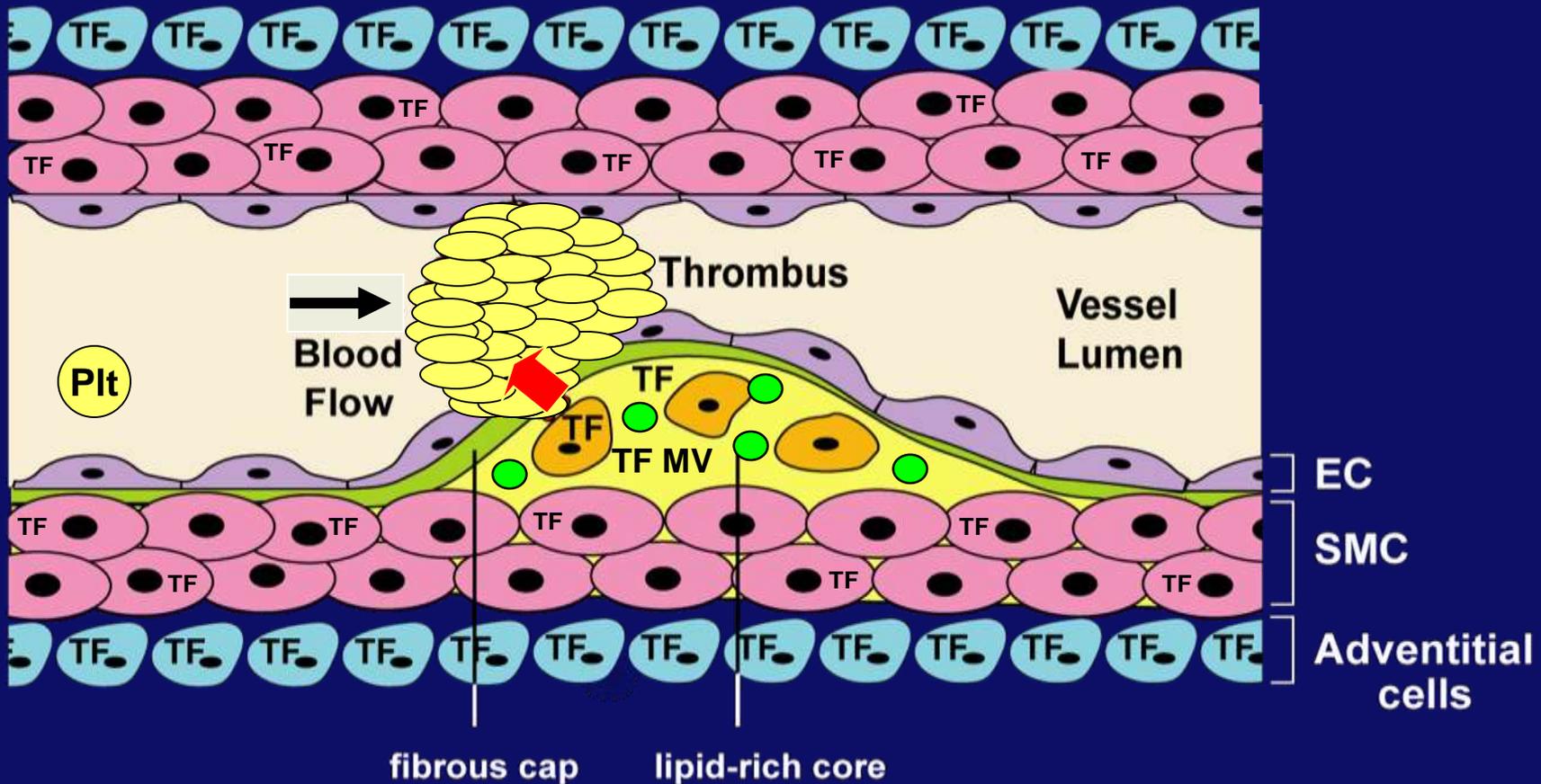


# Effect of Combining Low Levels of TF with PAR4 Deficiency

Expected numbers of low TF, PAR4 deficient mice at wean

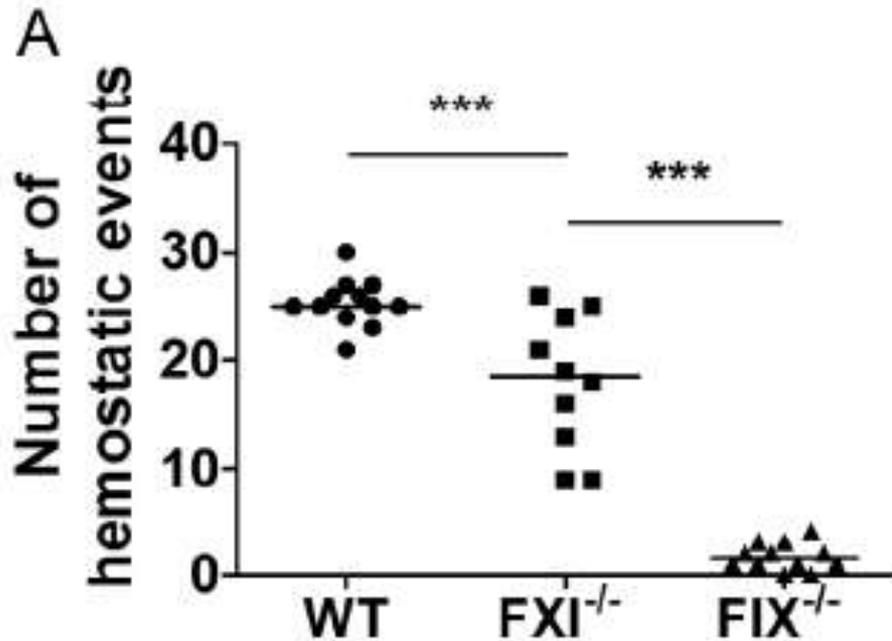


# Role of TF in Thrombosis After Rupture of an Atherosclerotic Plaque



Mackman ATVB 2004

# FXI<sup>-/-</sup> Mice Exhibit Increased Bleeding in the Saphenous Vein Model



# Effect of Combining Low Levels of TF with FXI Deficiency on Blood Pools in the Placenta

---



mTF<sup>+/-</sup>, FXI<sup>+/+</sup>



mTF<sup>-/-</sup>, FXI<sup>+/-</sup>

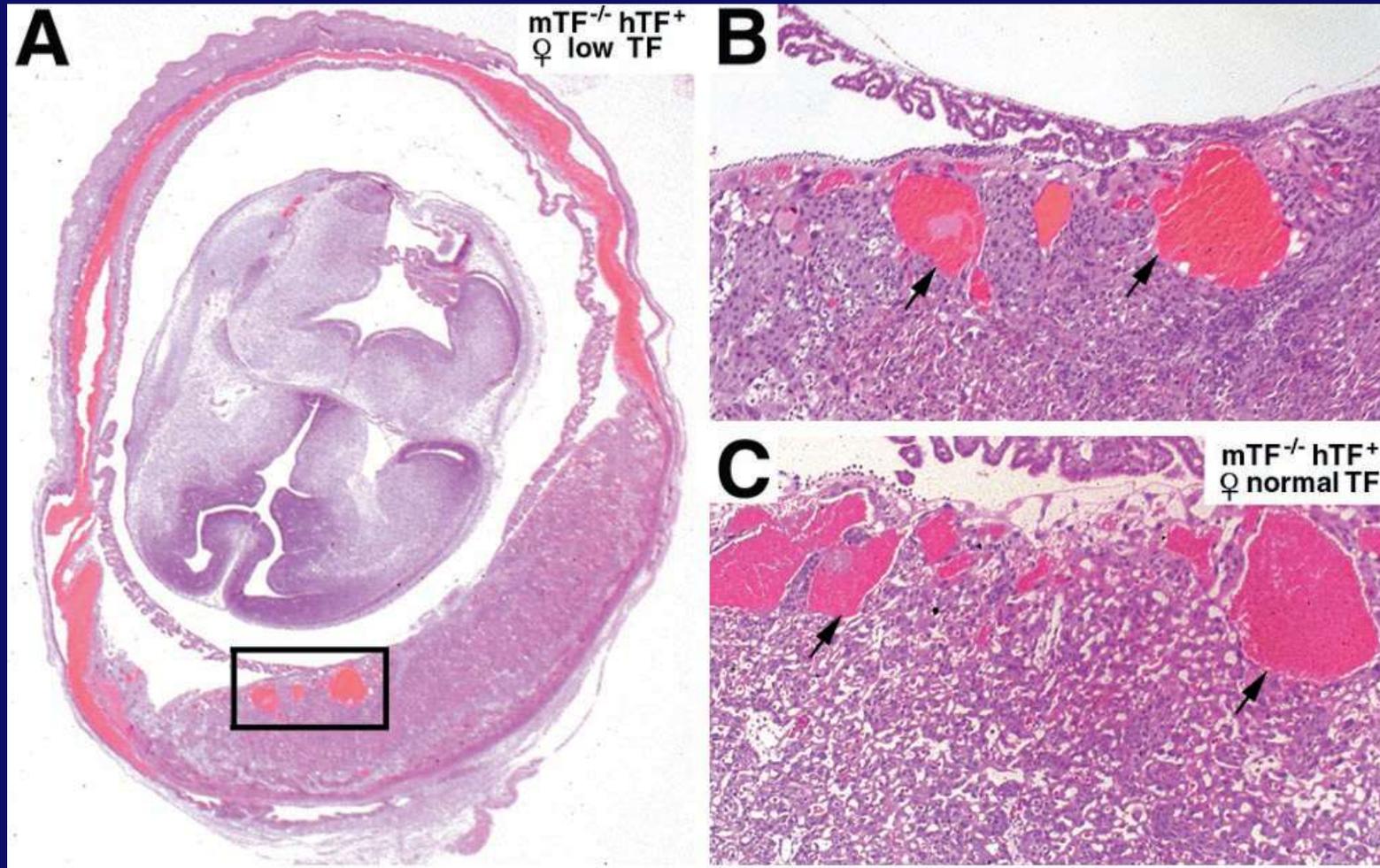


mTF<sup>-/-</sup>, FXI<sup>-/-</sup>

Grover unpublished

**Are the blood pools in low TF  
placentas due to a hemostatic  
defect?**

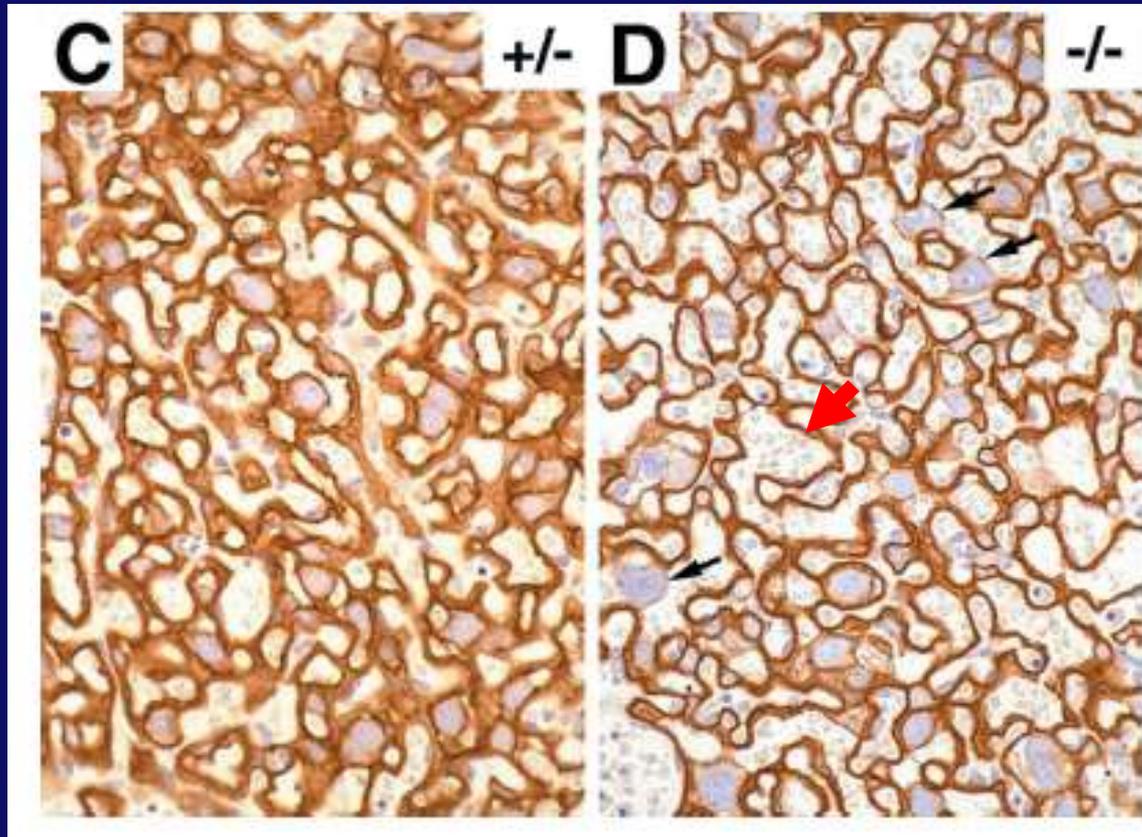
# Low TF Placentas Contain Blood Pools Regardless of the Level of TF in the Mother



Erlich et al PNAS 1999

# Cytokeratin Staining in Placenta

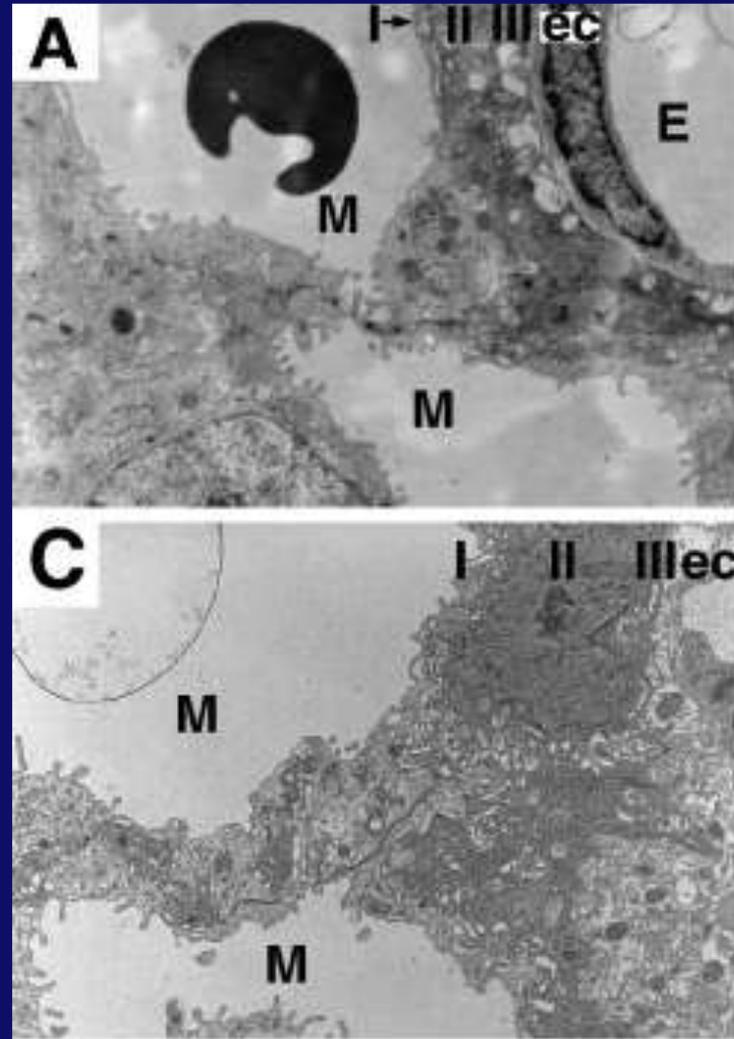
---



Reduced cyokeratin staining in layer I trophoblasts-  
multiple attachments to adjacent trabeculae  
Larger maternal lacunae in low TF placentas  
Erlich et al PNAS 1999

# Ultrastructural of the Placenta

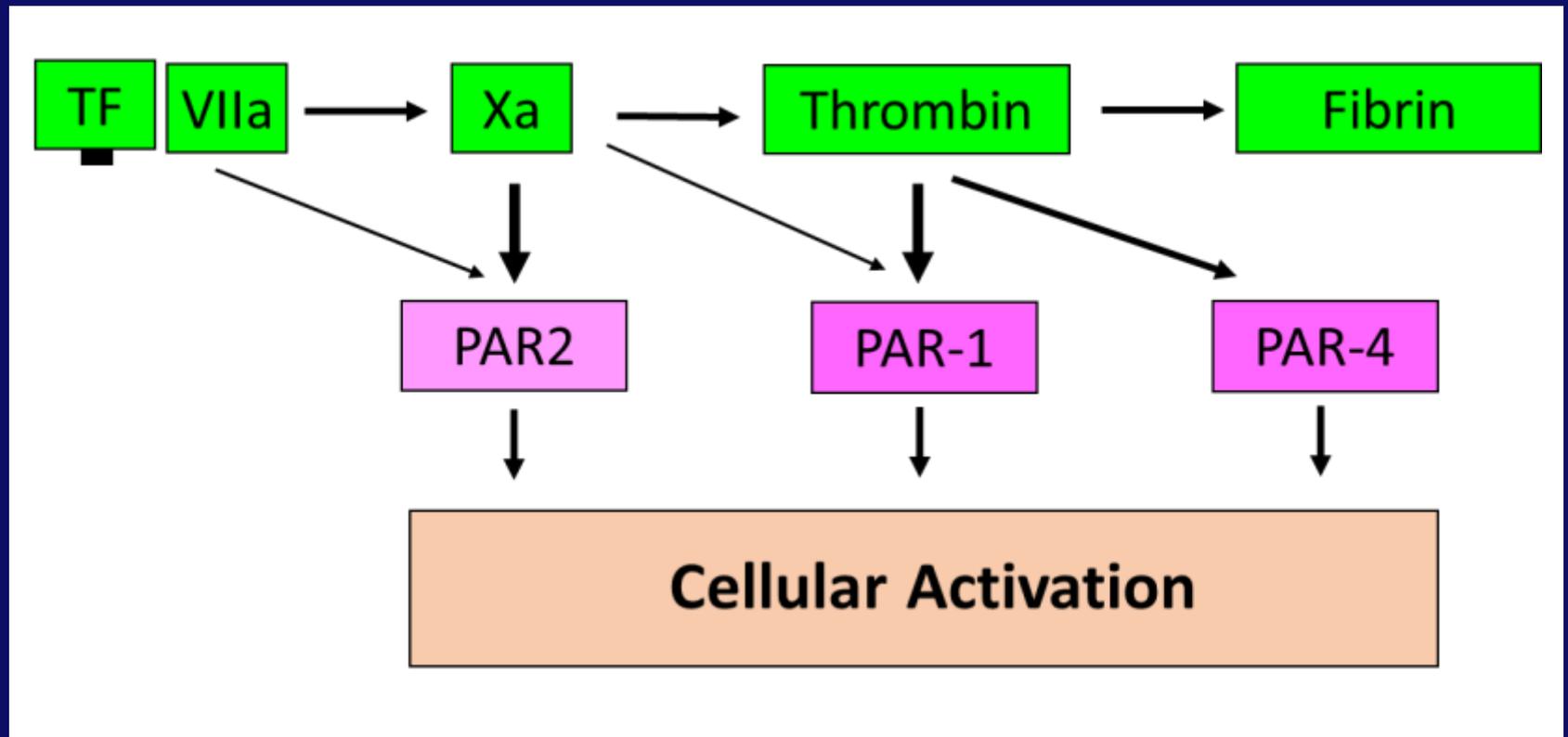
Reduced areas of contact between layer I trophoblasts that subdivide maternal lacunae



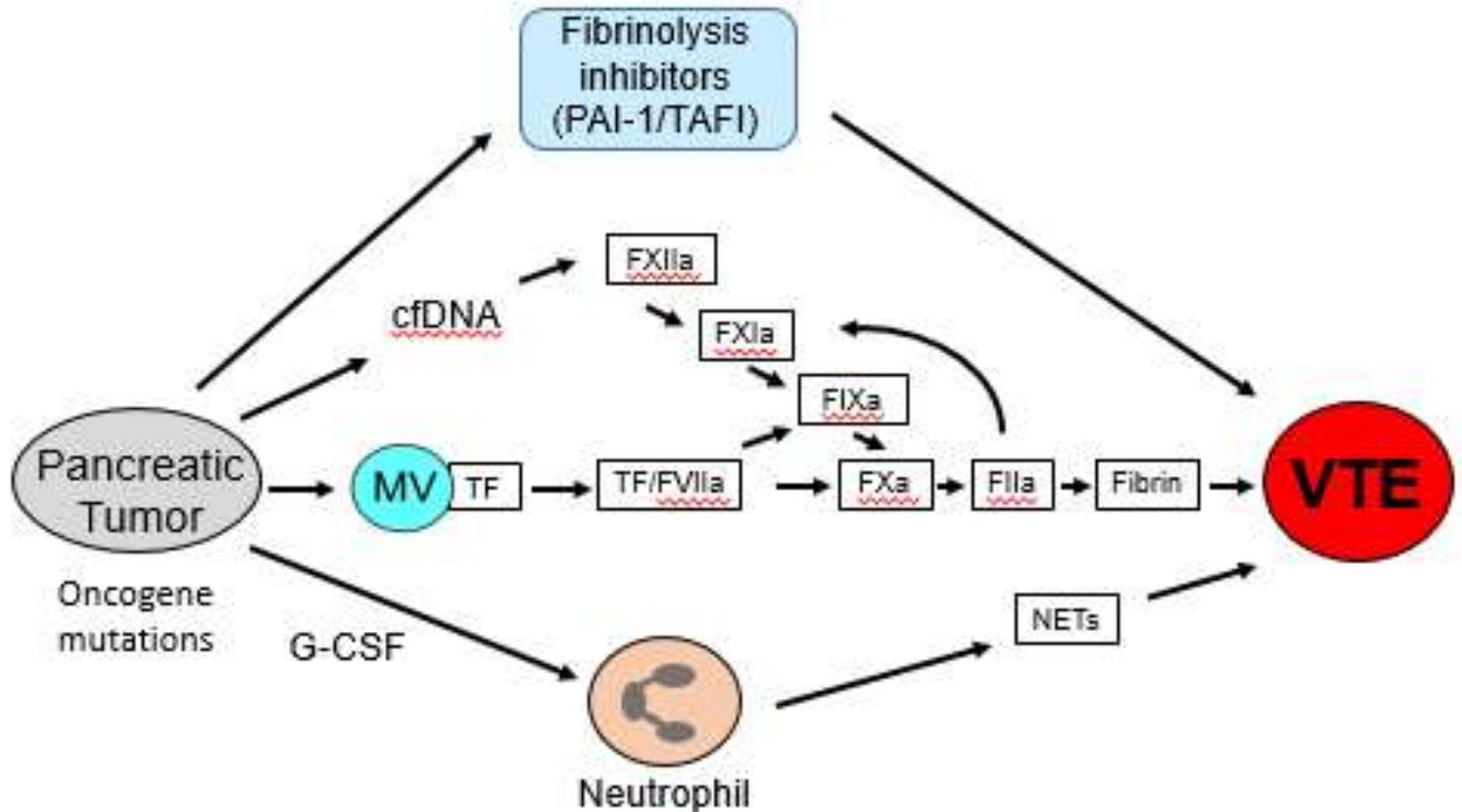
Erlich et al PNAS 1999

# Coagulation Proteases Activate PARs

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# Pathways that Increase VTE in Pancreatic Cancer Patients



# Effect of Crossing Low TF Mice with Mice Deficient in Genes of the Intrinsic Pathway

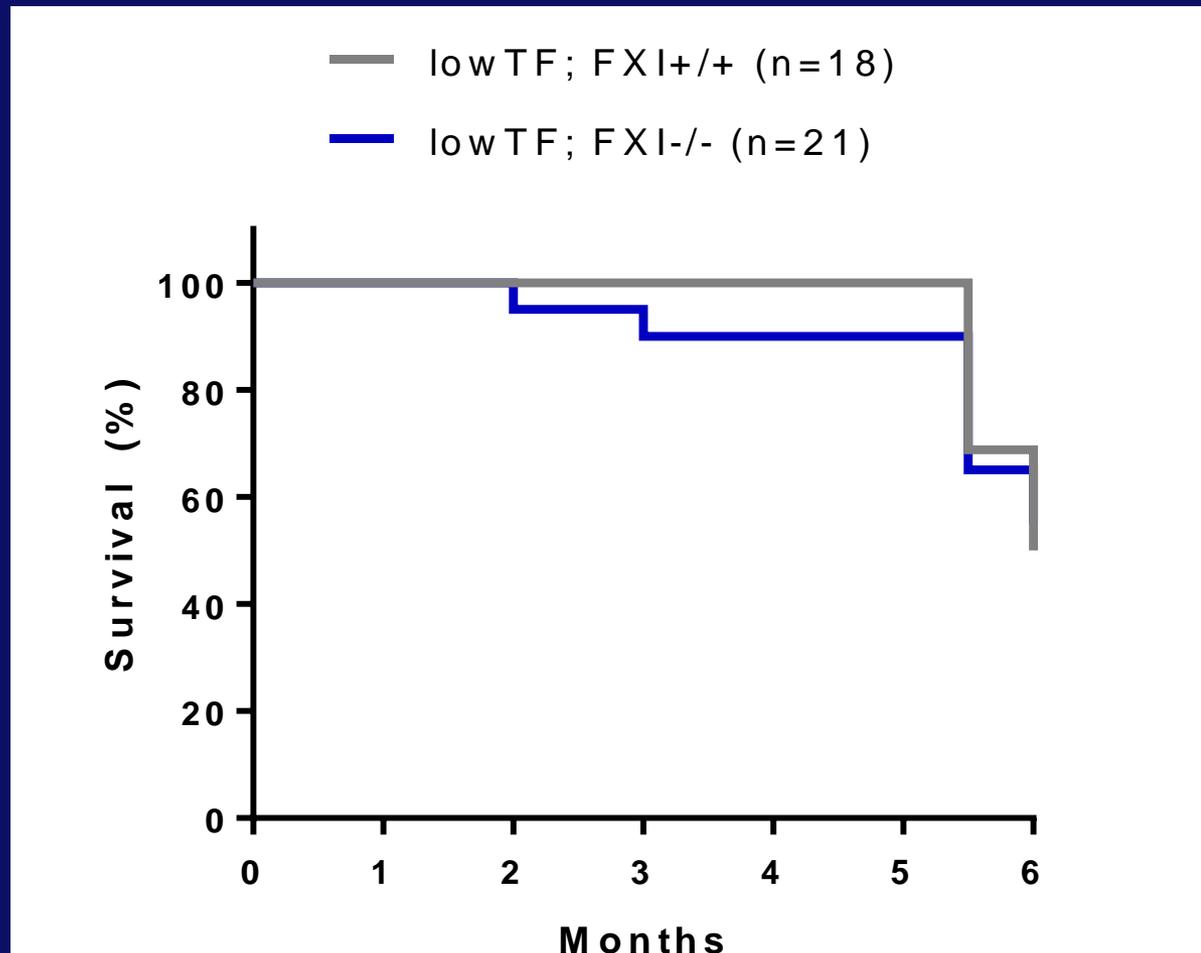
Low Tissue Factor and...	Survival	Low Tissue Factor and...	Survival	
FXII Deficiency	<b>Yes</b>	FXII Deficiency	<b>Yes</b>	
FXI Deficiency	<b>No</b>	FXI Deficiency	<b>Yes*</b>	<b>Loop 2</b>
FIX Deficiency	<b>No</b>	FIX Deficiency	<b>No</b>	<b>Loop 1</b>

Spronk H et al ISTH 2009

Mackman N 2017

\*increased placental blood pools and maternal death.

# Survival of Low TF Mice without FXI<sup>-/-</sup> Mice



Jamsa A et al unpublished data

# Ongoing Studies

---

- Determine the cause of death of mTF<sup>+/-</sup>, hTF<sup>+</sup>, FXI<sup>+/-</sup> mothers carrying Low TF, FXI<sup>-/-</sup> embryos. Placental hemorrhage?
- Determine when Low TF, FIX<sup>-/-</sup> embryos die.

# Effect of Crossing Two Prohemorrhagic Mice

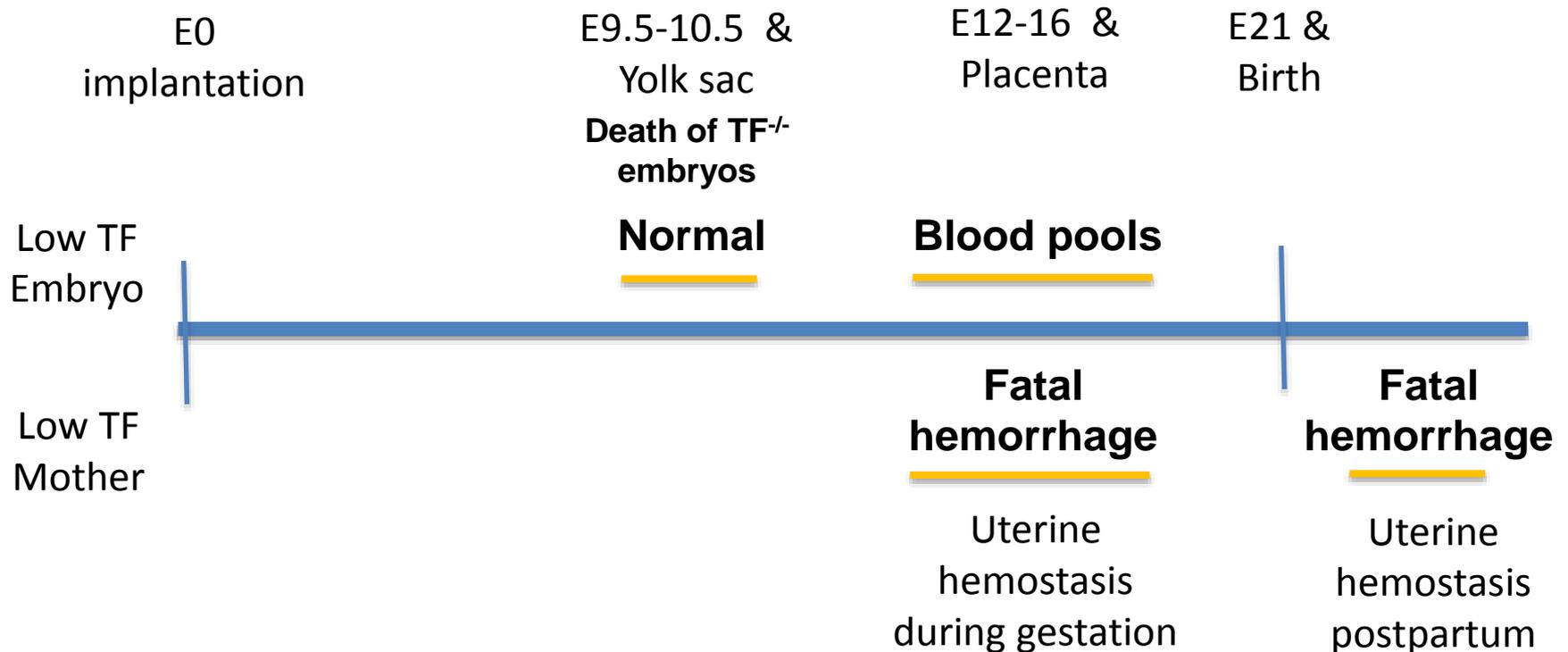
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Can we exacerbate the bleeding in Low TF mice by adding another prohemorrhagic genetic defect?

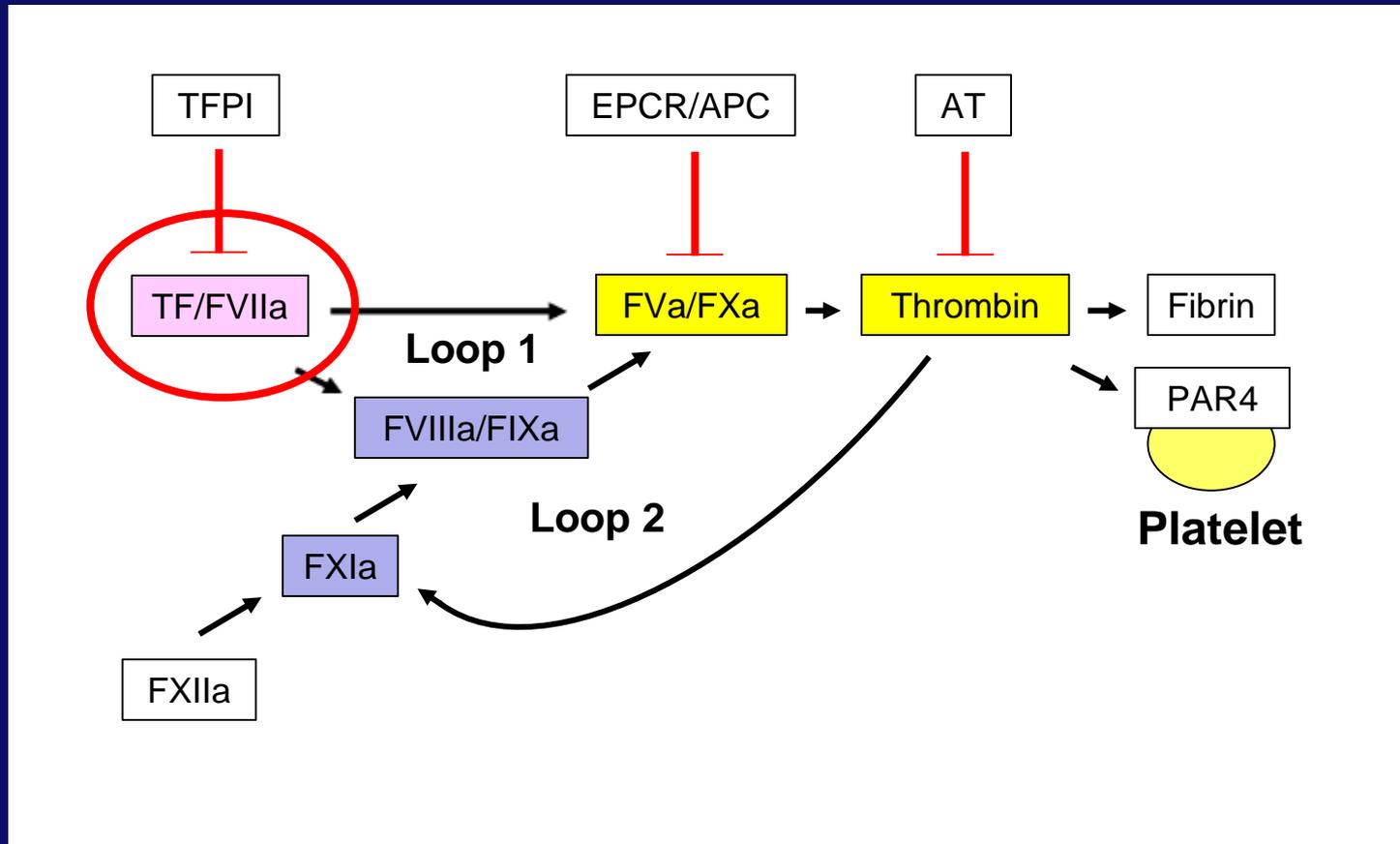
Combine Low TF with PAR-4 deficiency (no thrombin activation of platelets).

Bode & Mackman Thromb Res 2016

# Summary of the Hemostatic Defects in Low TF Placentas and Mothers



# Simplified Version of the Clotting Cascade



# Effect of Inactivation of the TFPI Gene in Mice

---

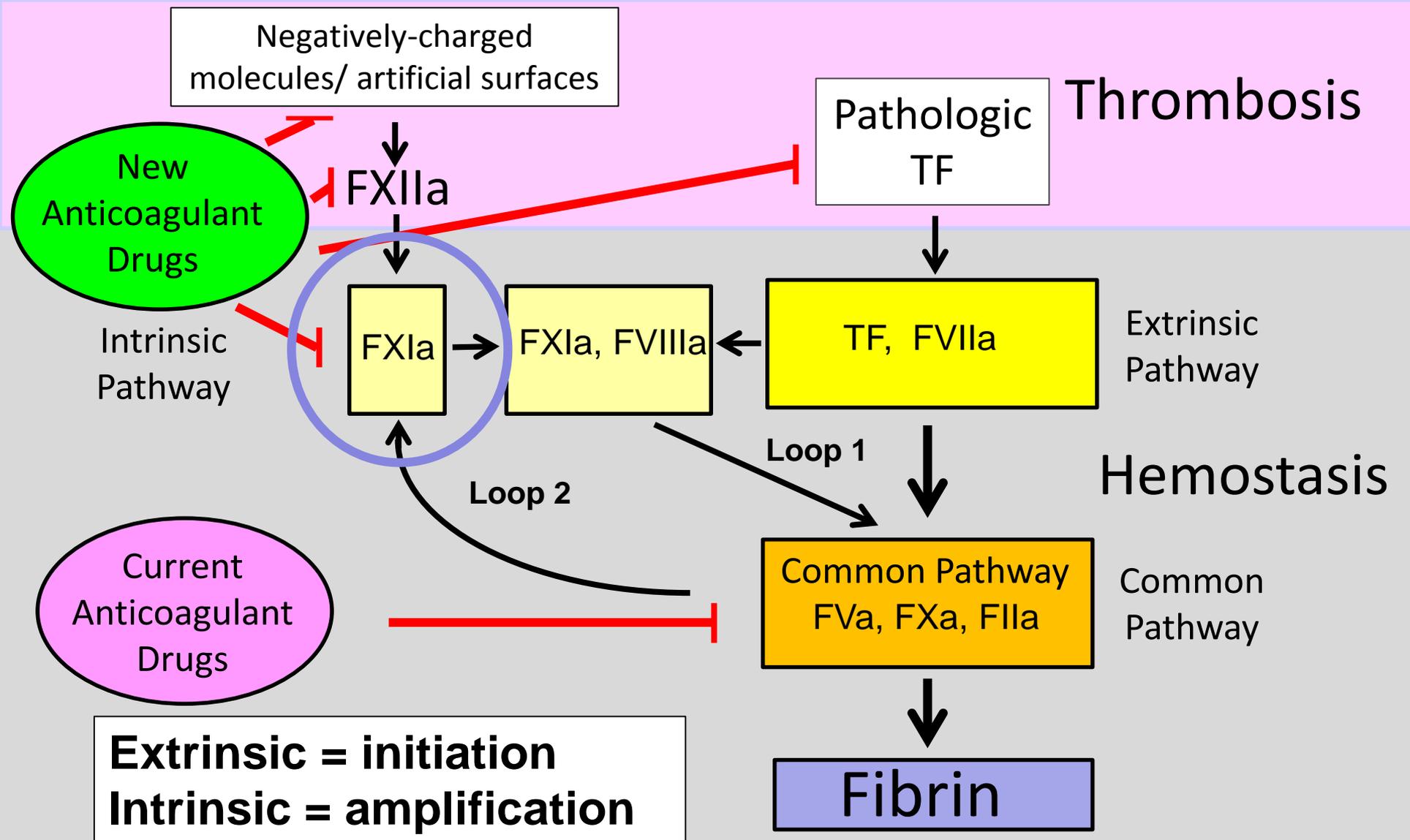
TFPI null embryos die during embryonic development



**TFPI is essential for survival**

Huang Z et al Blood 1997

# Development of New Anticoagulant Drugs





## Factor XI Antisense Oligonucleotide for Prevention of Venous Thrombosis

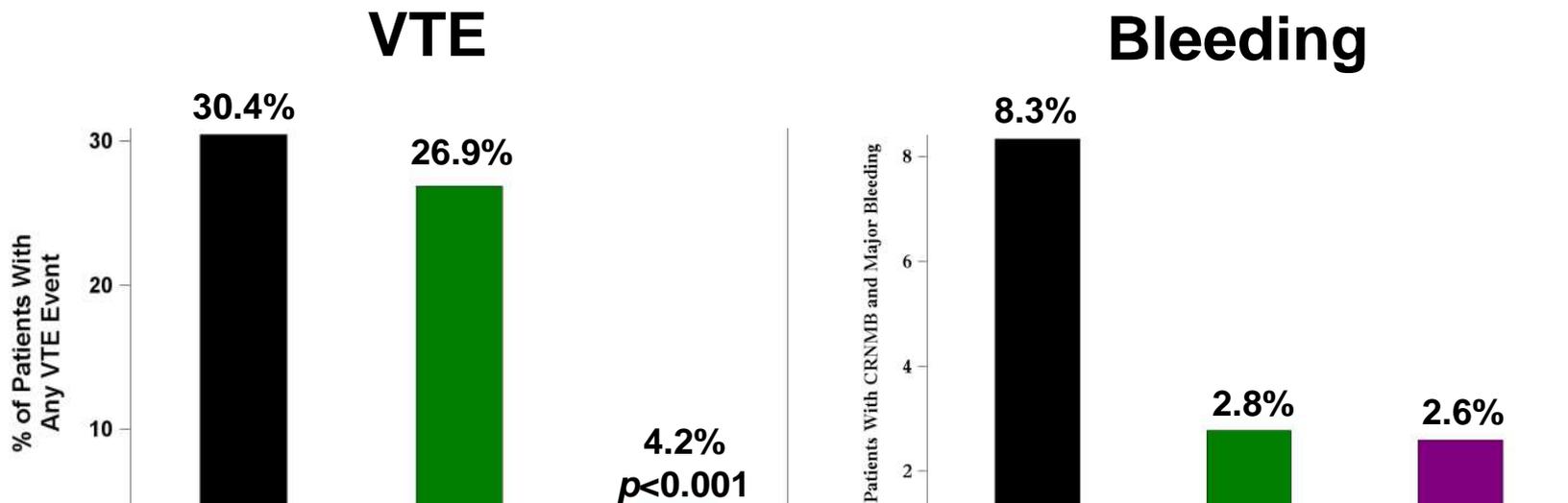
Harry R. Büller, M.D., Claudette Bethune, Ph.D., Sanjay Bhanot, M.D., Ph.D, David Gailani, M.D., Brett P. Monia, Ph.D., Gary E. Raskob, Ph.D., Annelise Segers, M.D., Peter Verhamme, M.D., and Jeffrey I. Weitz, M.D.

2015;372:232-240

- Effect of decreased levels of FXI on VTE in pts undergoing total knee replacement
- Two doses of a FXI ASO were used: 200 mg ~41% FXI; 300 mg ~22% FXI.
- Enoxaparin was used as a control.

Slides from Dr. J. Weitz

# Effect of Reducing FXI Levels on VTE and Bleeding after Total Knee Replacement

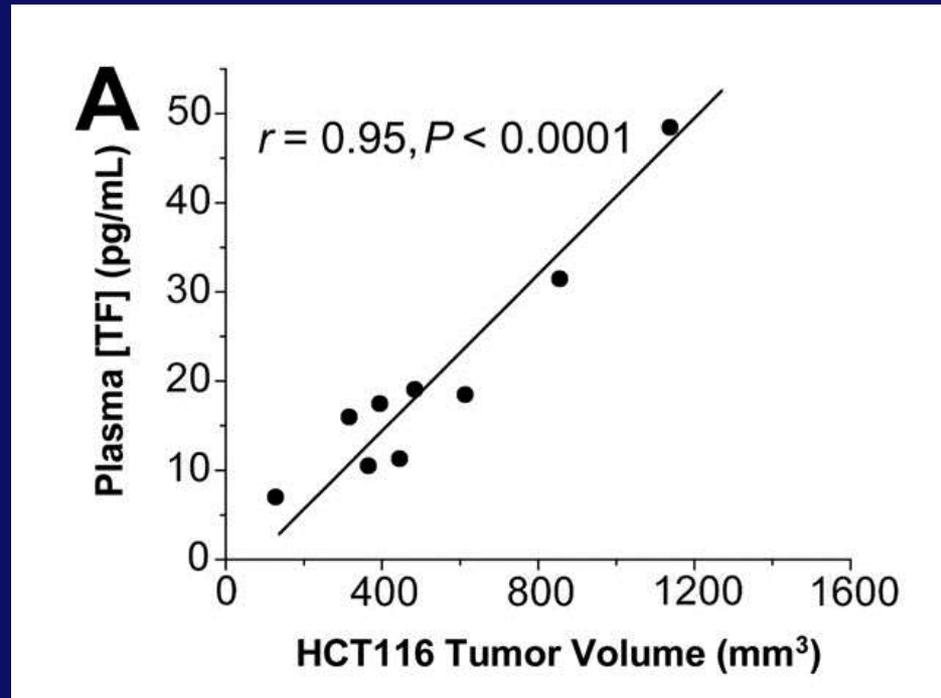


**FXI appears to be a promising target to reduce VTE without increasing bleeding**

**Do human tumors grown  
in mice release TF+ MV  
into the blood?**

**Move**

# Tumors Release Human TF into the Circulation in SCID Mice containing Human Tumors



**Move**

human colorectal cell line

Yu et al Blood 2005

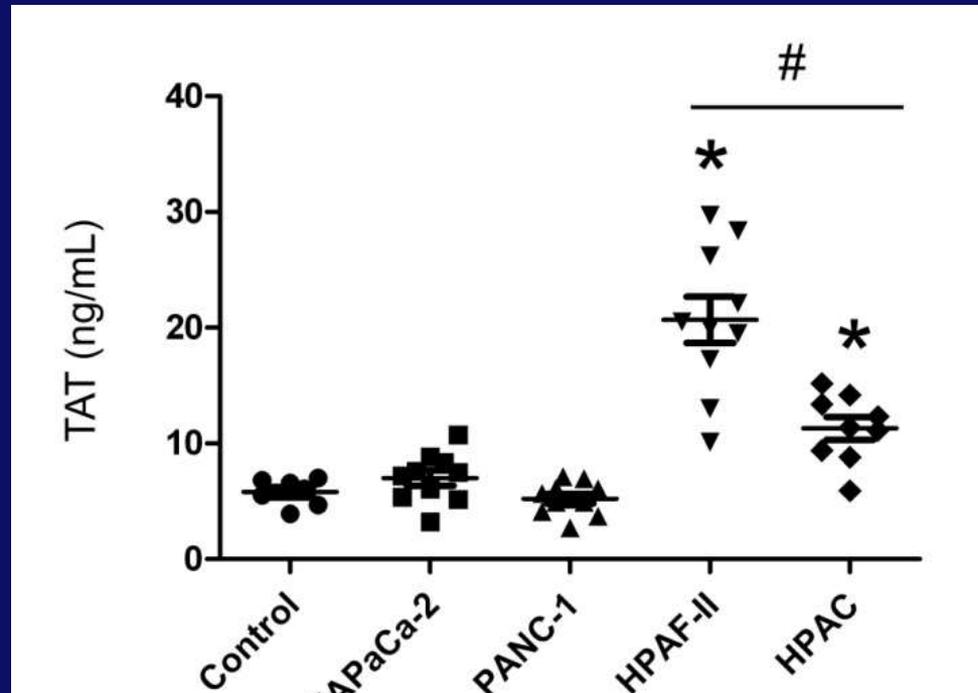
**Do TF<sup>+</sup> human pancreatic  
tumors grown in mice  
activate coagulation?**

**Move**

# Activation of Coagulation in Nude Mice containing Human Pancreatic Tumors



Jianguo Wang



An anti-human TF Ab abolishes the activation of coagulation in tumor-bearing mice

Move



# Effect of Inactivation of the TF Gene in Mice

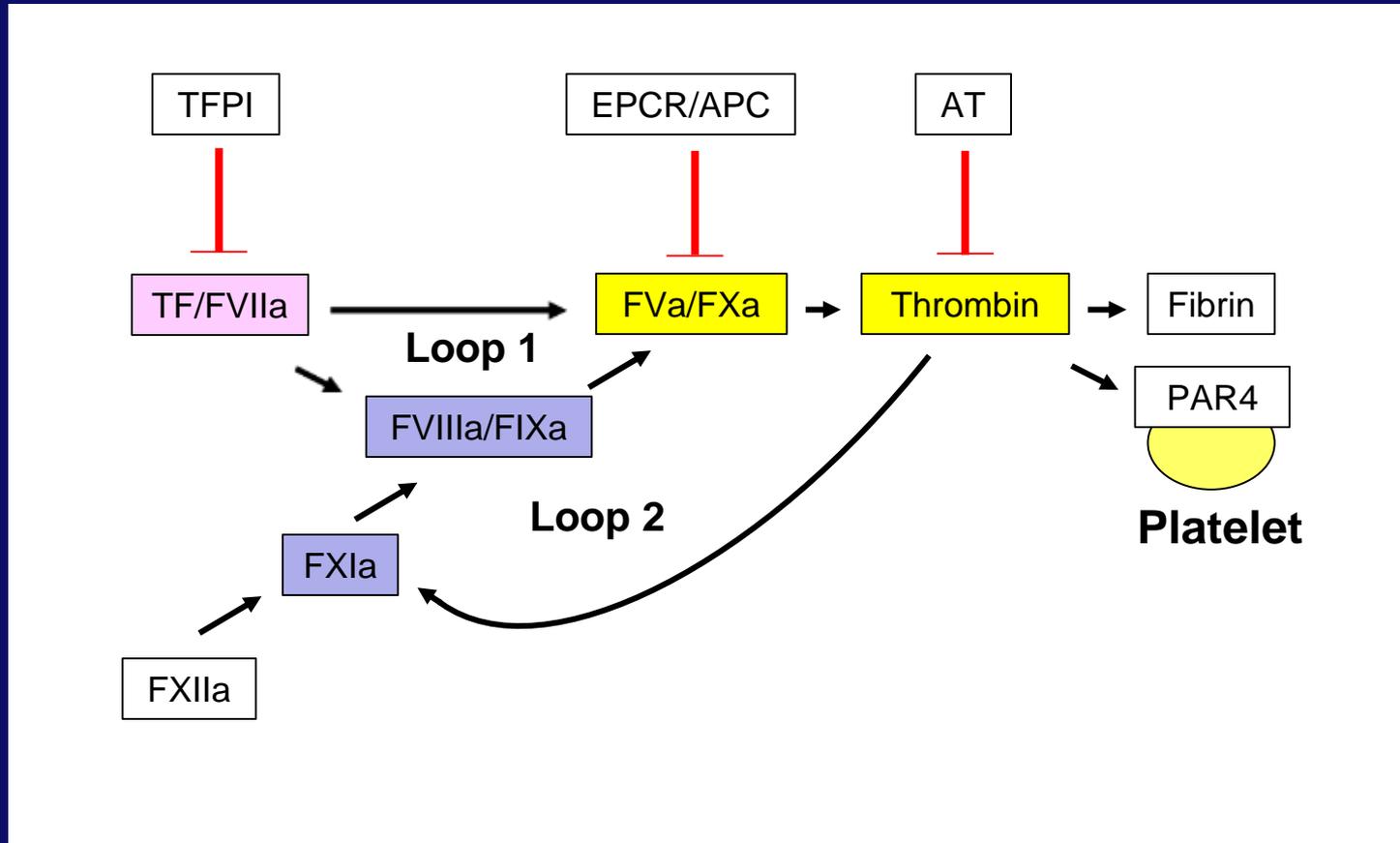
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TF null embryos die during embryonic development

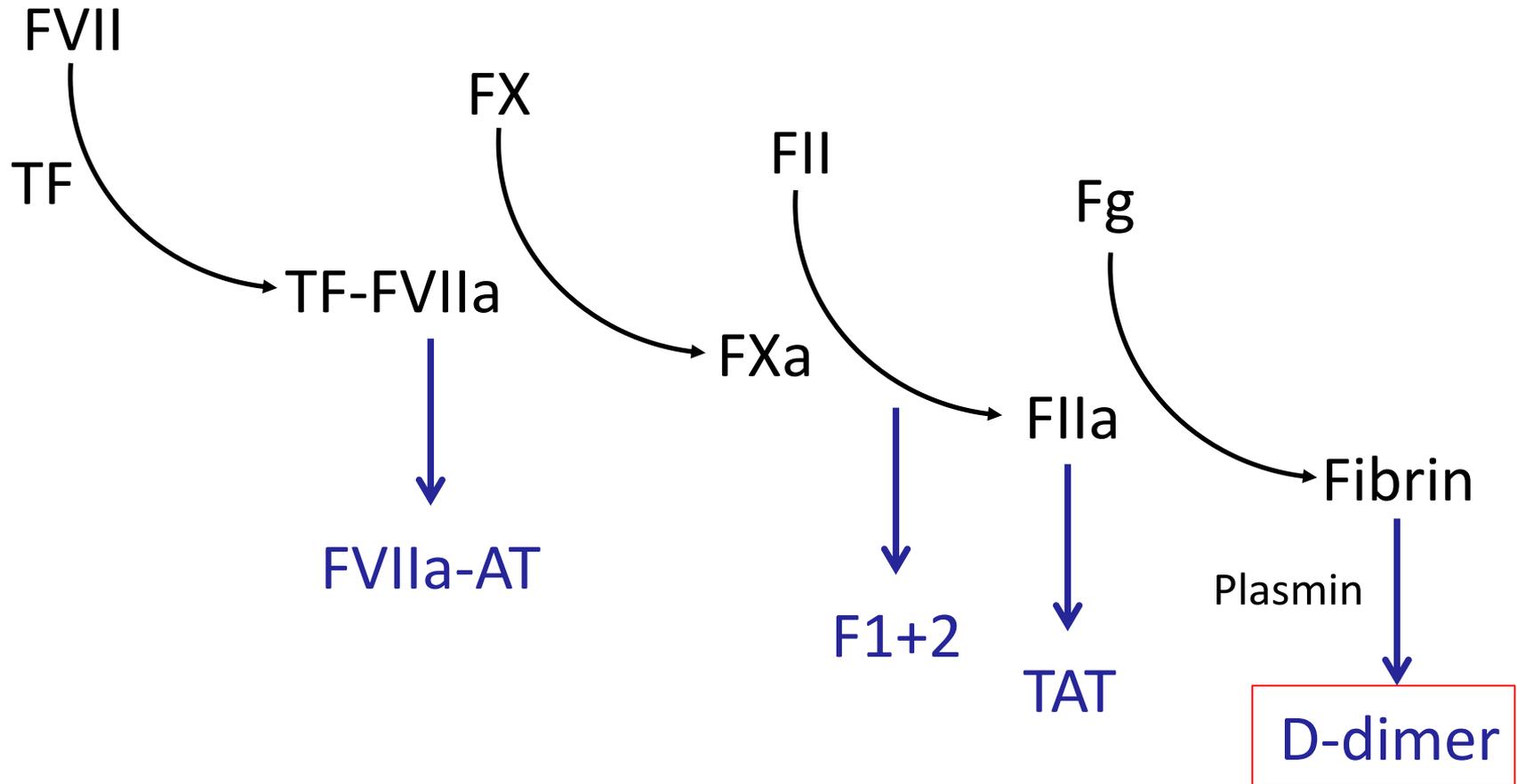
- Carmeliet, Mackman et al *Nature* 1996
- Bugge et al *PNAS* 1996
- Toomey et al *Blood* 1996

**TF are essential for survival**

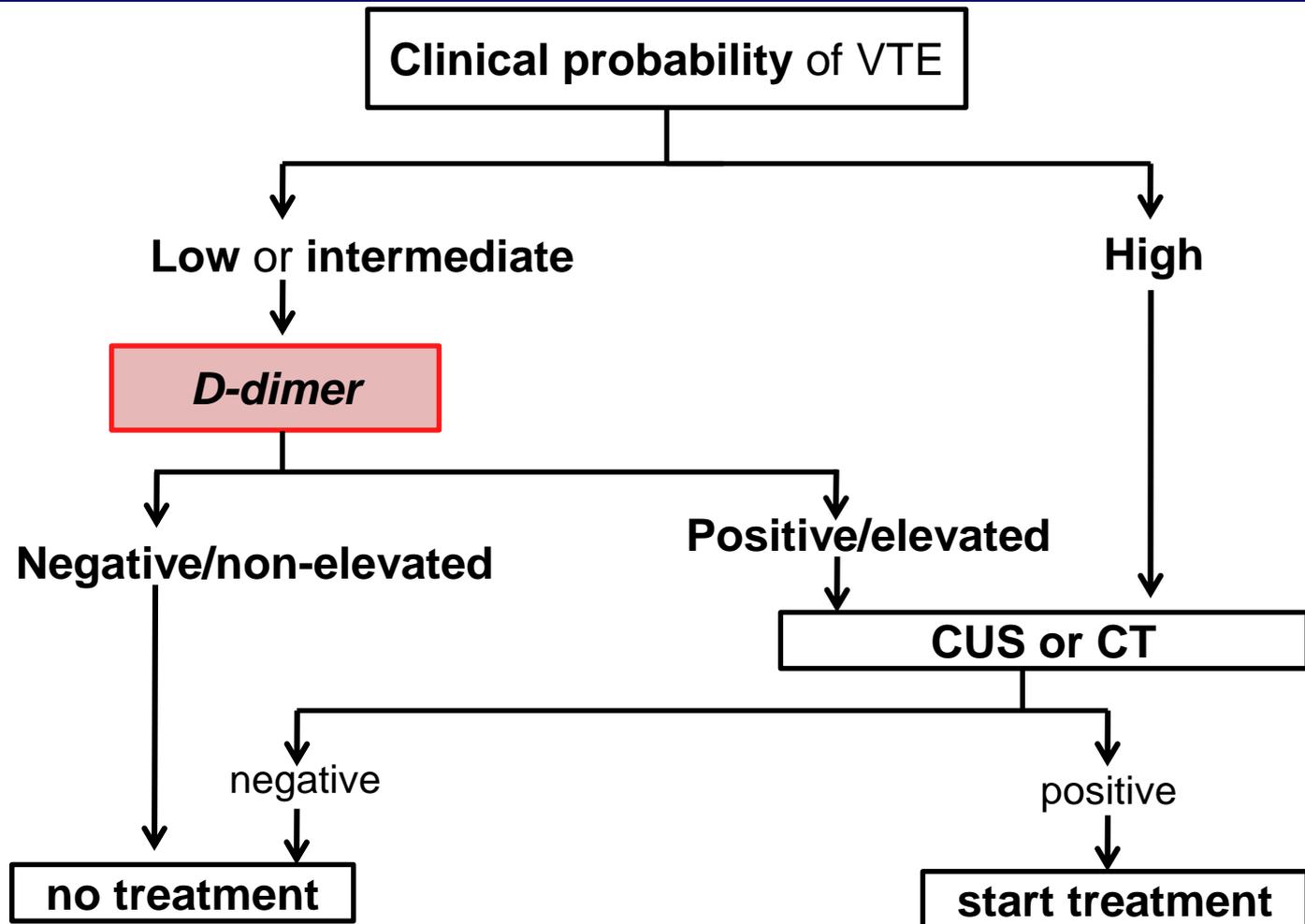
# Simplified Version of the Clotting Cascade



# Biomarkers Associated with Activation of the Clotting Cascade



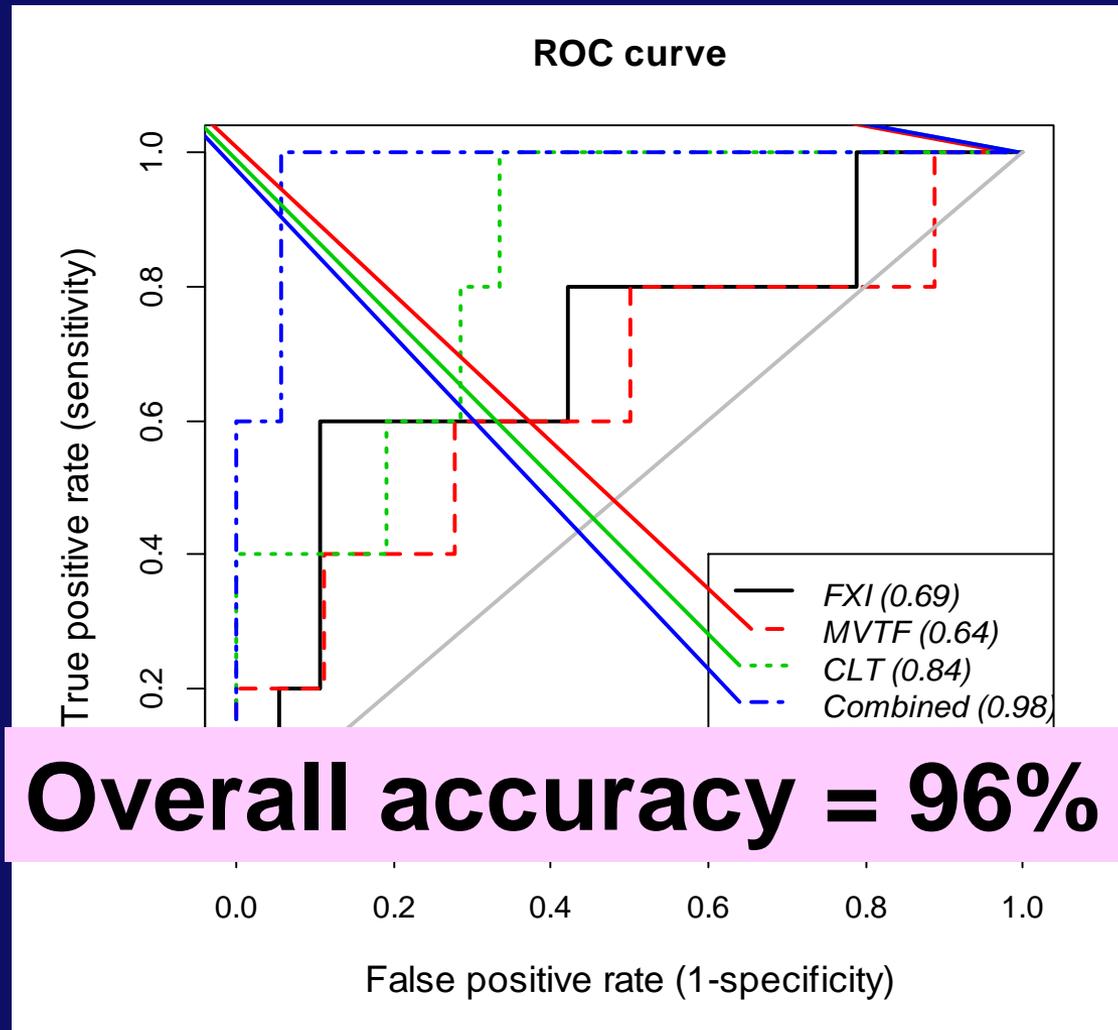
# Measurement of D-dimer in the Diagnostic Work-up of Suspected Venous Thromboembolism (VTE)



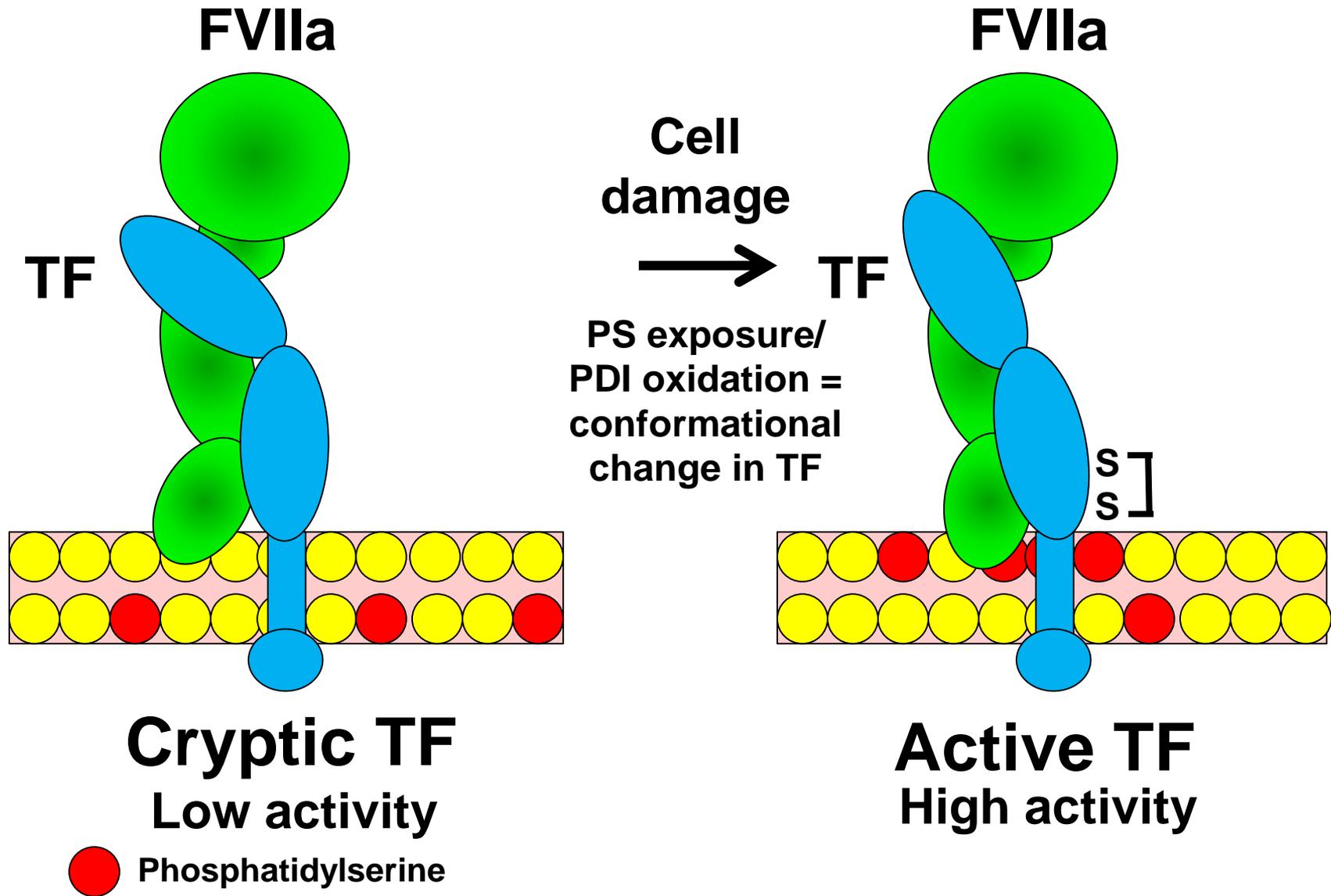
# Biomarkers for Predicting VTE in Pancreatic Cancer Patients



Data from 26  
pancreatic  
cancer pts (5  
developed VTE)

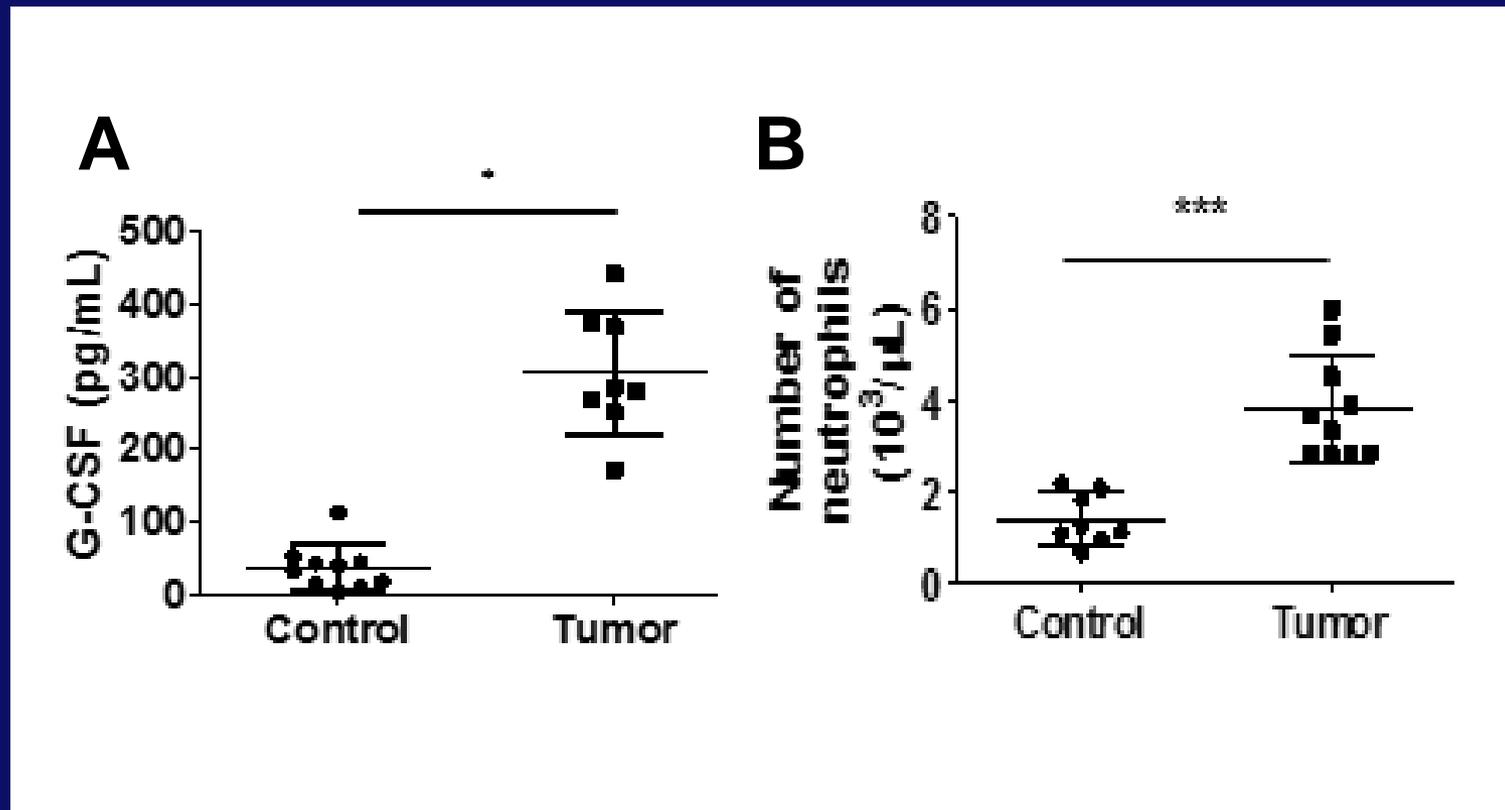


# Regulation of TF Activity by PS



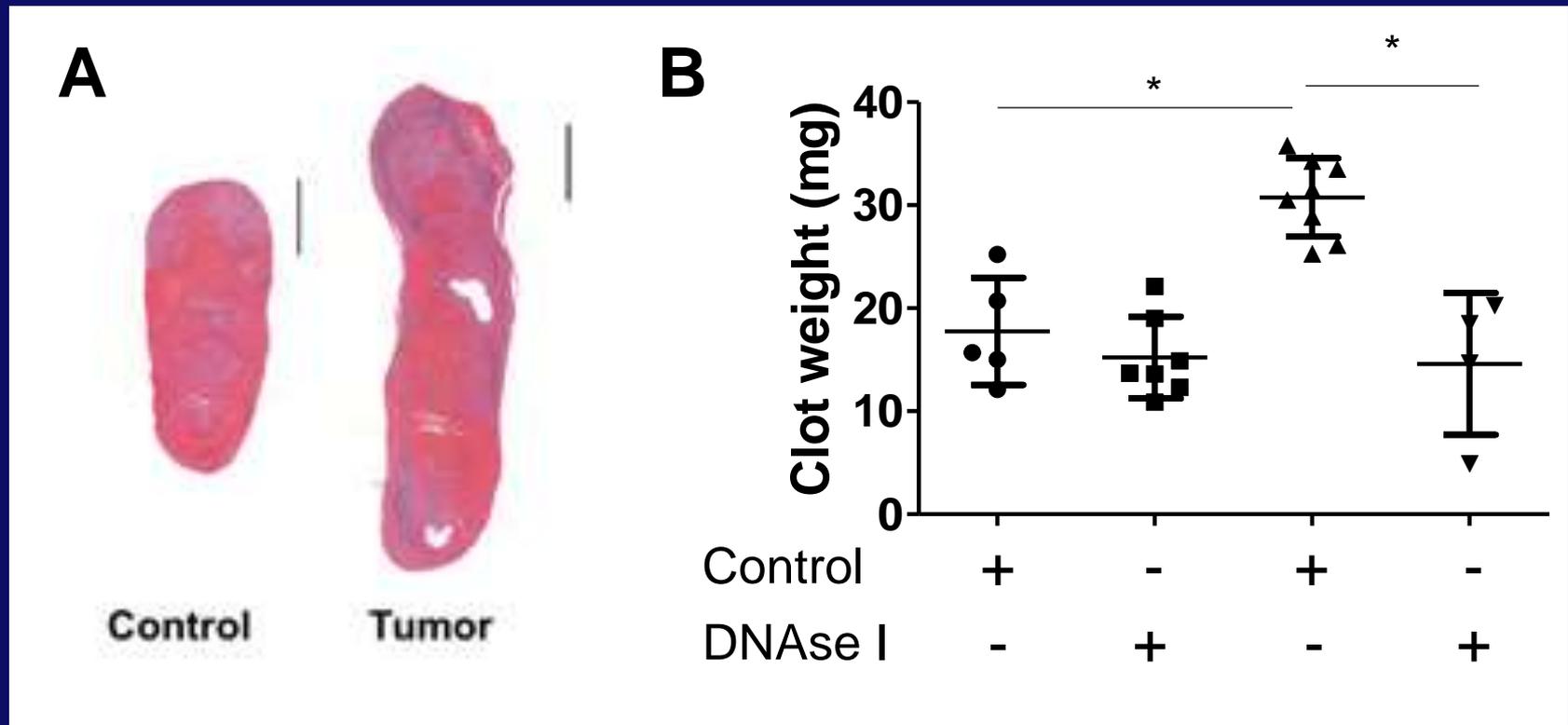


# Tumor-bearing Mice have Increased Levels of G-CSF and Neutrophils



Hisada Y et al JTH 2017; unpublished data

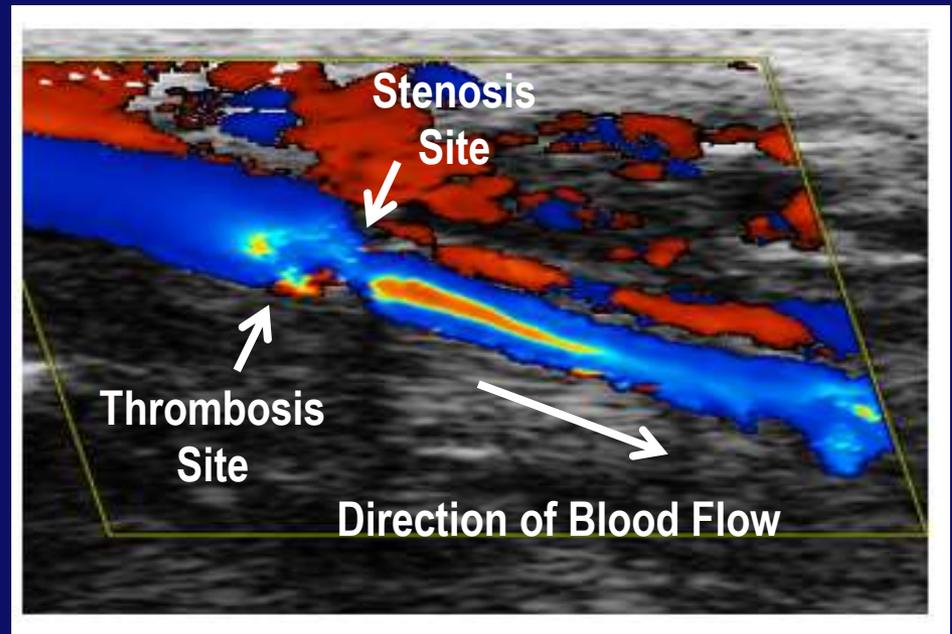
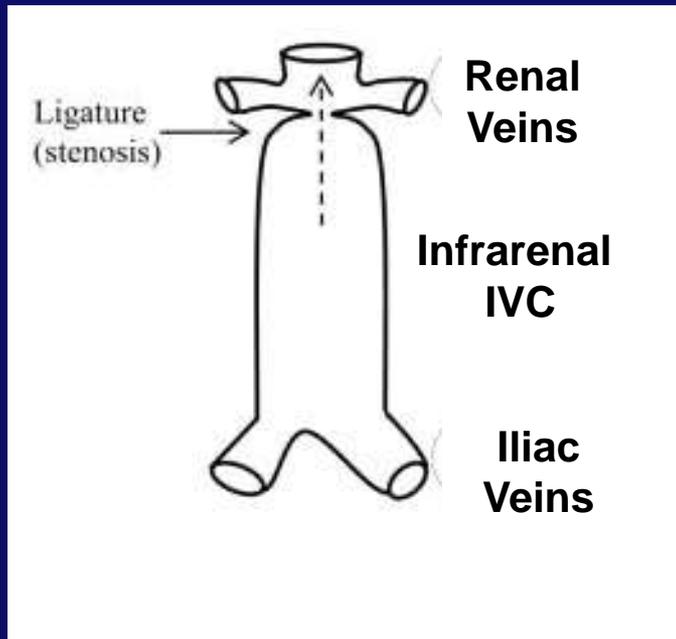
# Effect of DNase 1 Treatment on Clot Size In Tumor-bearing Mice



Hisada Y et al JTH 2017; unpublished data

# Mouse Model of Venous Thrombosis: Inferior Vena Cava Stenosis or Stasis

## Color Doppler

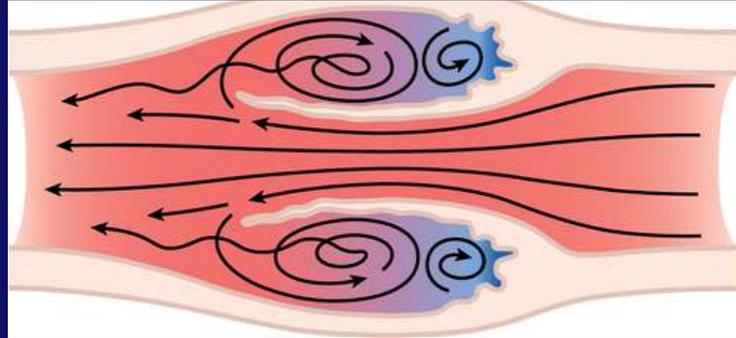


**Can perform kinetic studies on thrombus formation**

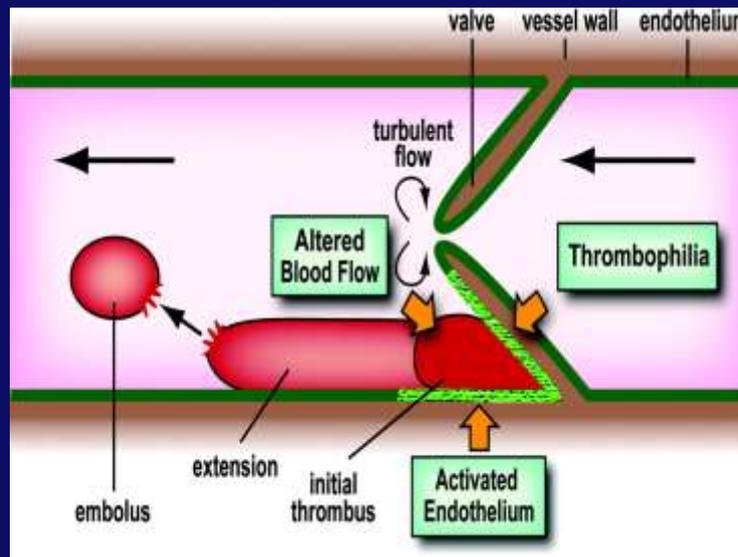
Brill A et al. Blood 2011; von Brühl M et al. J Exp Med 2012; Geddings J et al. JTH 2014

# Formation of a Deep Vein Thrombus

Hypoxia in valve pockets



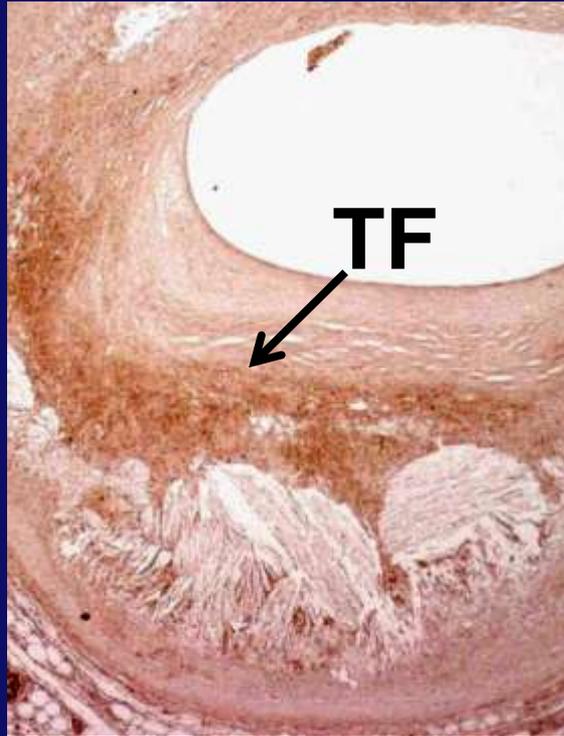
Bovill Ann Rev Physiol 2011



Moll and Mackman ATVB 2008

# High Levels of TF are Present in Atherosclerotic Plaques

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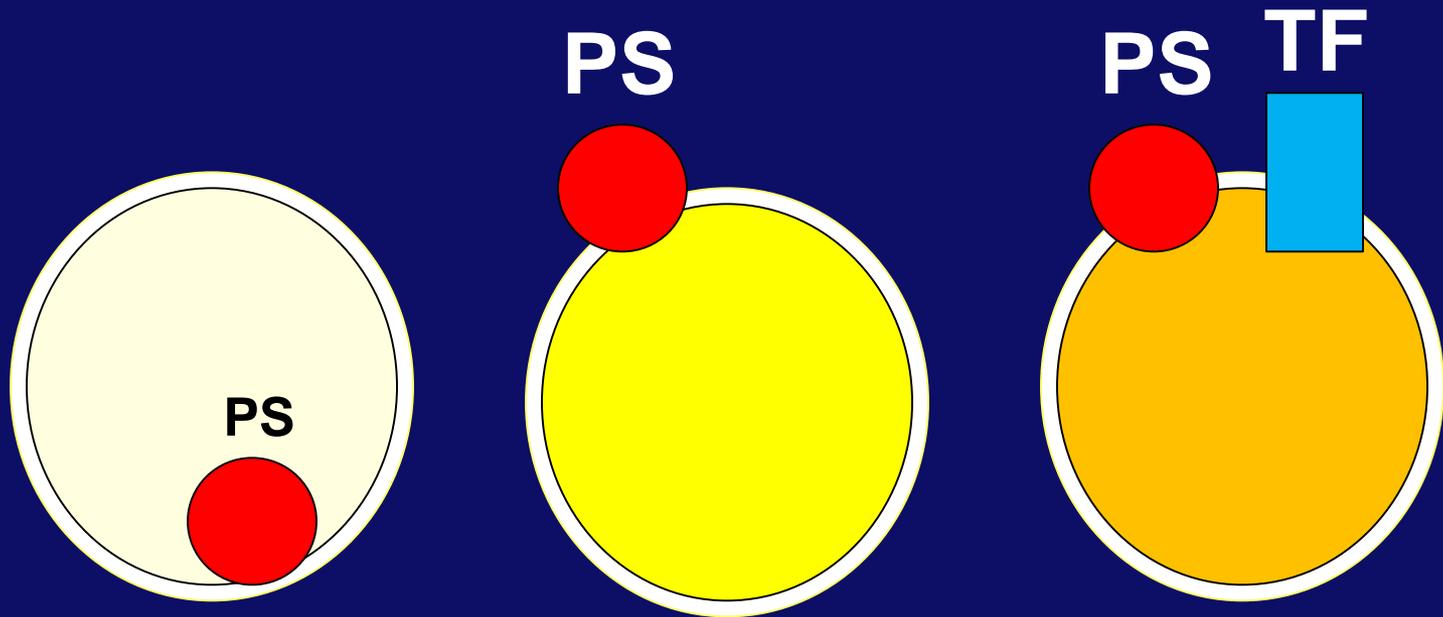


**“Thrombotic” TF**

Slide provided by Dr. Y. Asada

# Types of Procoagulant EVs

---



Procoagulant  
activity

+

++

+++

Origin

Plt/RBC

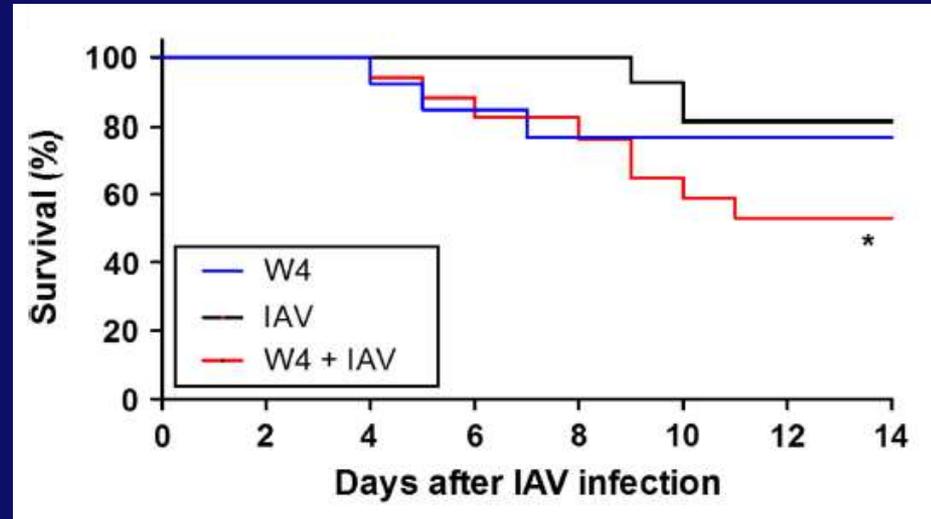
Plt/RBC

Mo/TC

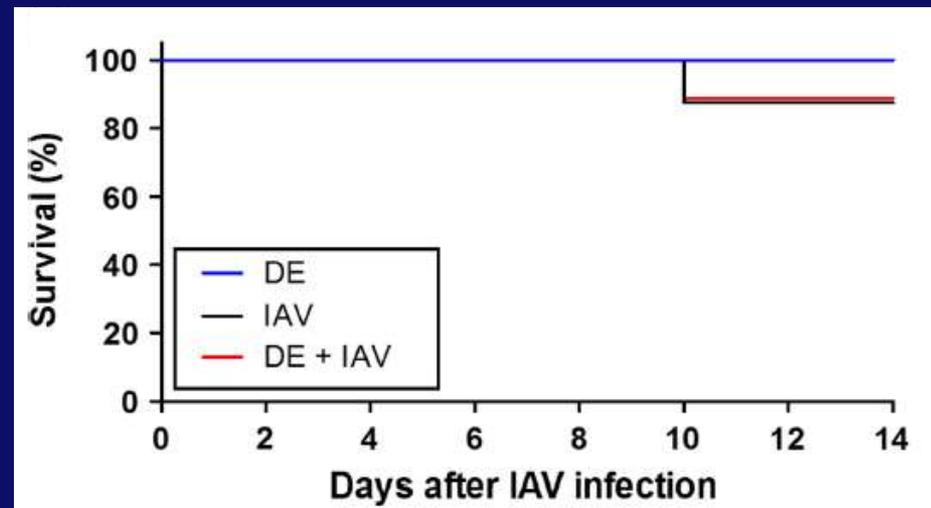
Plt, platelet; RBC, red blood cell, Mo, monocyte; TC, tumor cell

# Effect of Warfarin and Dabigatran on the Survival of Mice Infected with H1N1

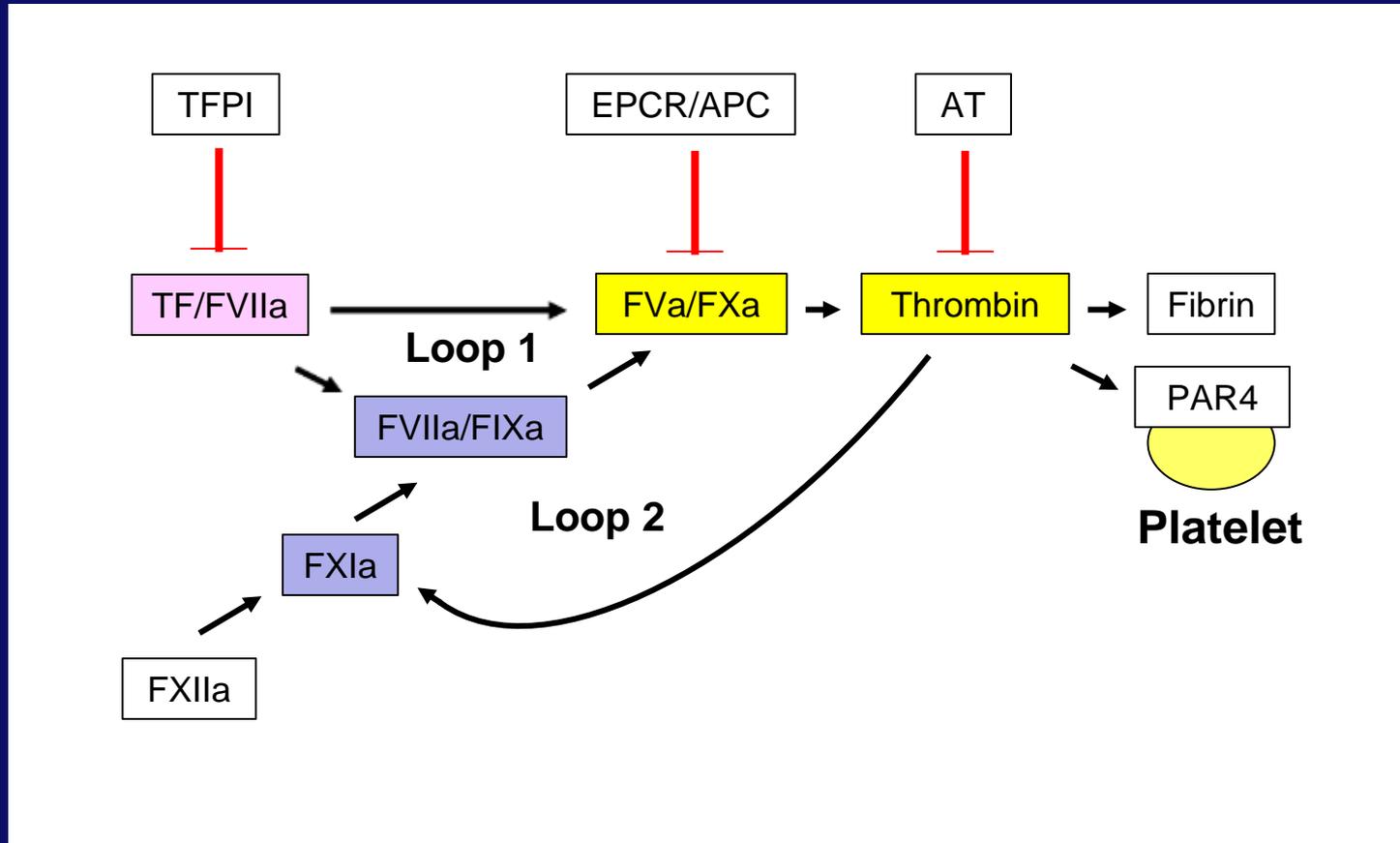
Warfarin



Dabigatran



# Simplified Version of the Clotting Cascade



# Clinical Characteristics and Biomarkers that are Predictive of VTE in Pancreatic Cancer

Patient	Death	VTE	Elevated WBC	Elevated Hbg	D-dimer (ng/mL)	MV-TF activity (pg/mL)	PAI-1 (ng/mL)	Clot lysis time (min)
6	n	n	n	n	0.9	0.1	9.25	2.18
7	n	n	n	n	-	0.08	-	-
8	n	n	n	n	-	0.32	-	-
9	n	n	n	n	1.93	0.16	-	1.97
12	n	n	n	n	0.74	0.06	1.58	3.52
14	n	n	n	n	0.74	0.16	0.32	2.07
15	n	n	n	n	1.12	0.01	15.46	3.17
20	y	y	y	y	36.42	1.23	31.78	23.97
22	n	n	n	n	2.24	0.39	3.97	2.18
27	y	n	y	y	12.34	0.94	19.33	5.92
35	n	n	n	n	0.85	-	0.06	2.15
38	n	n	n	n	0.76	0.03	2.24	2.47
40	n	n	n	n	0.96	0.02	2.07	1.62

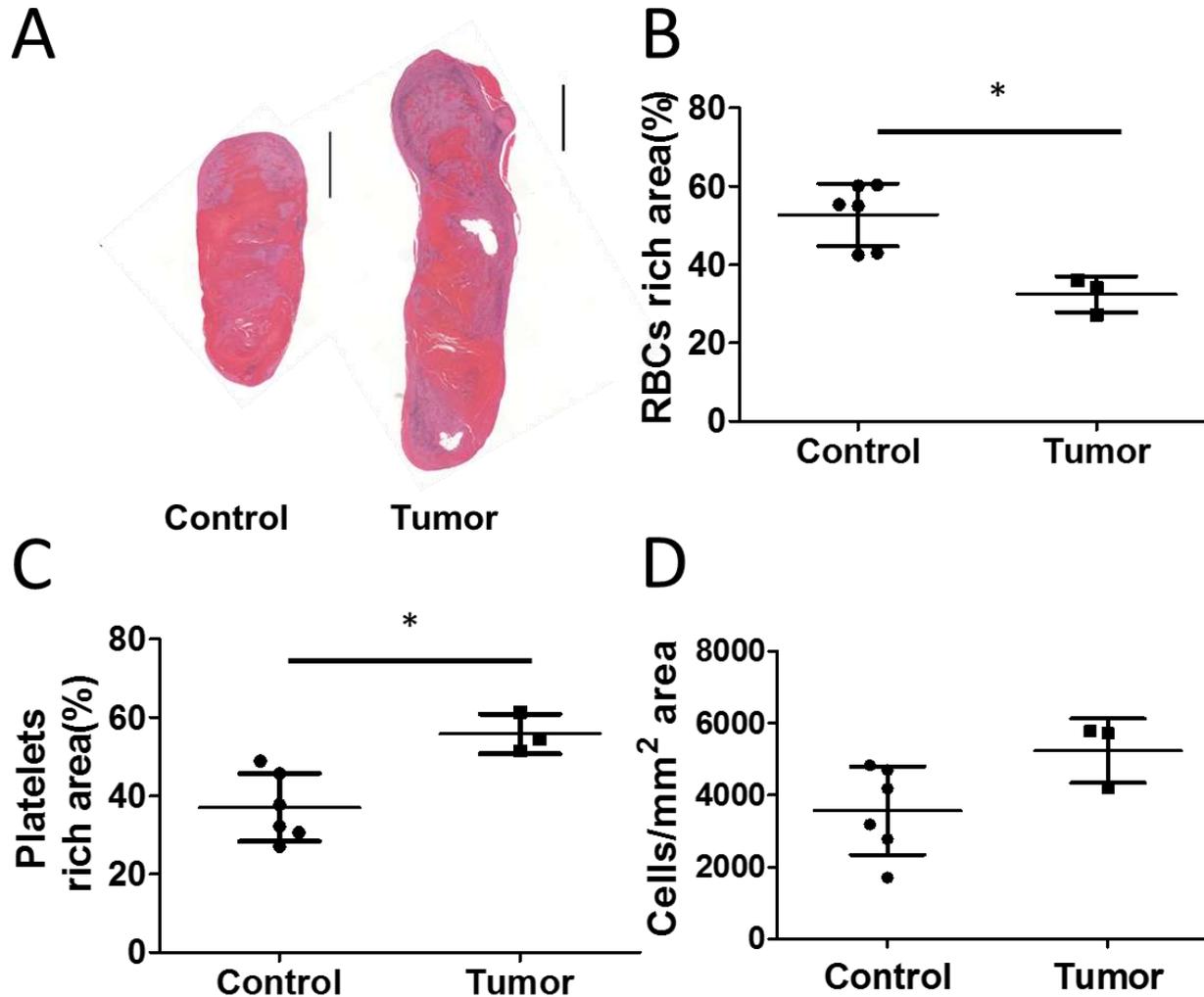
Kasthuri R et al in prep

# General Conclusions

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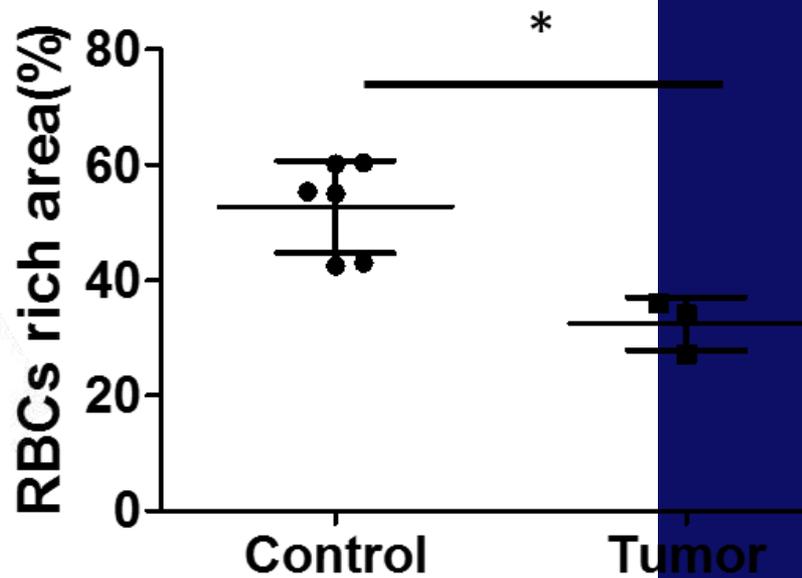
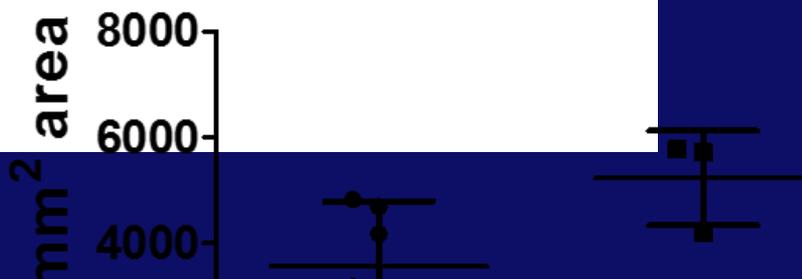
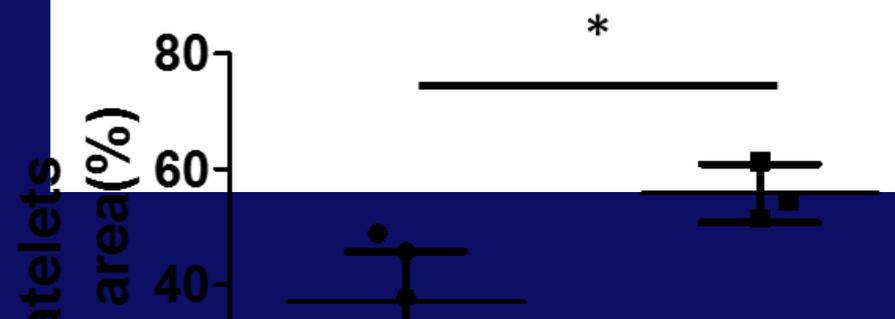
# Clots from Tumor-bearing Mice have more Platelets/Inflammatory Cells



**A**

Control

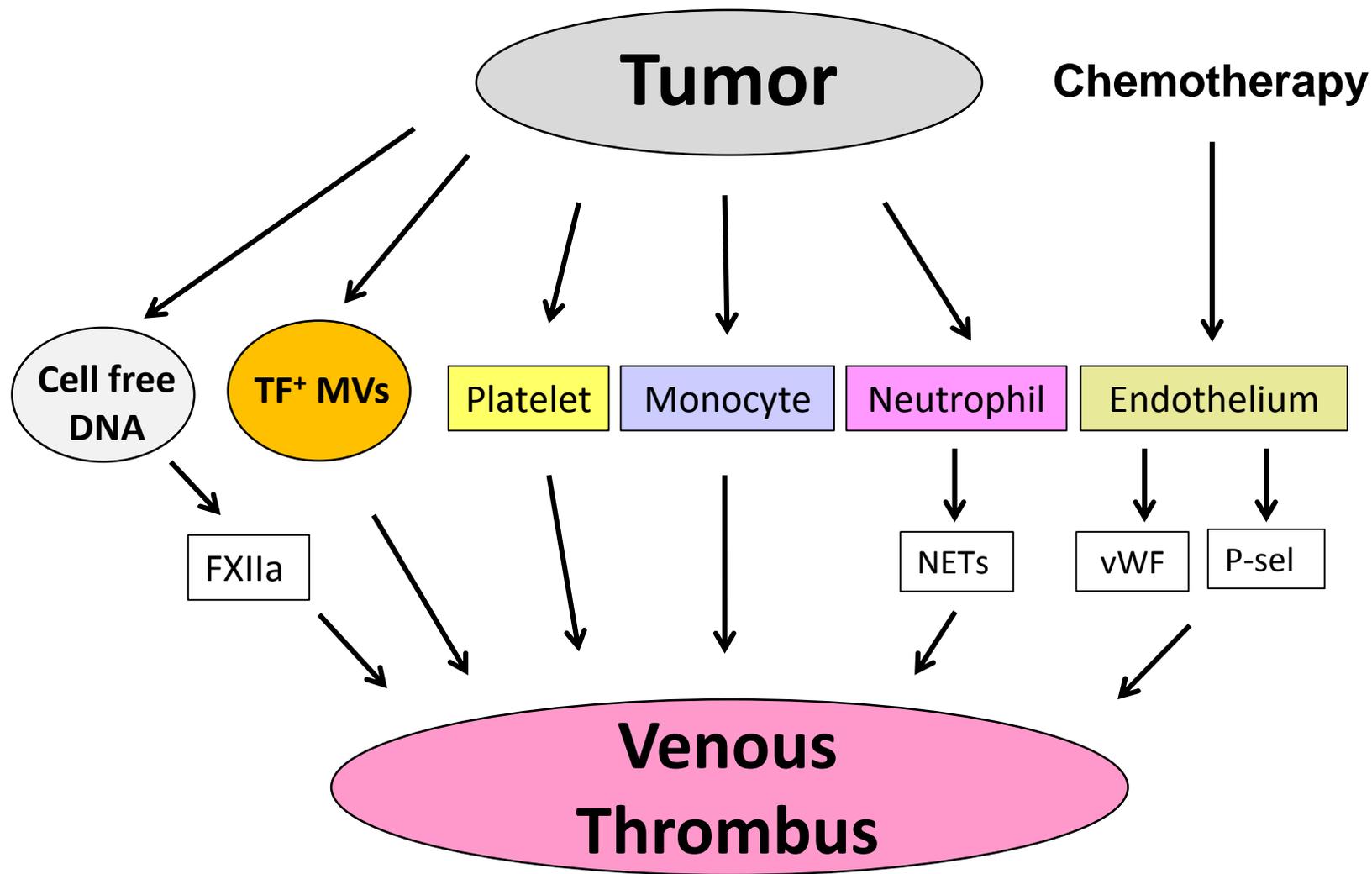
Tumor

**B****D****C**

splenocyte

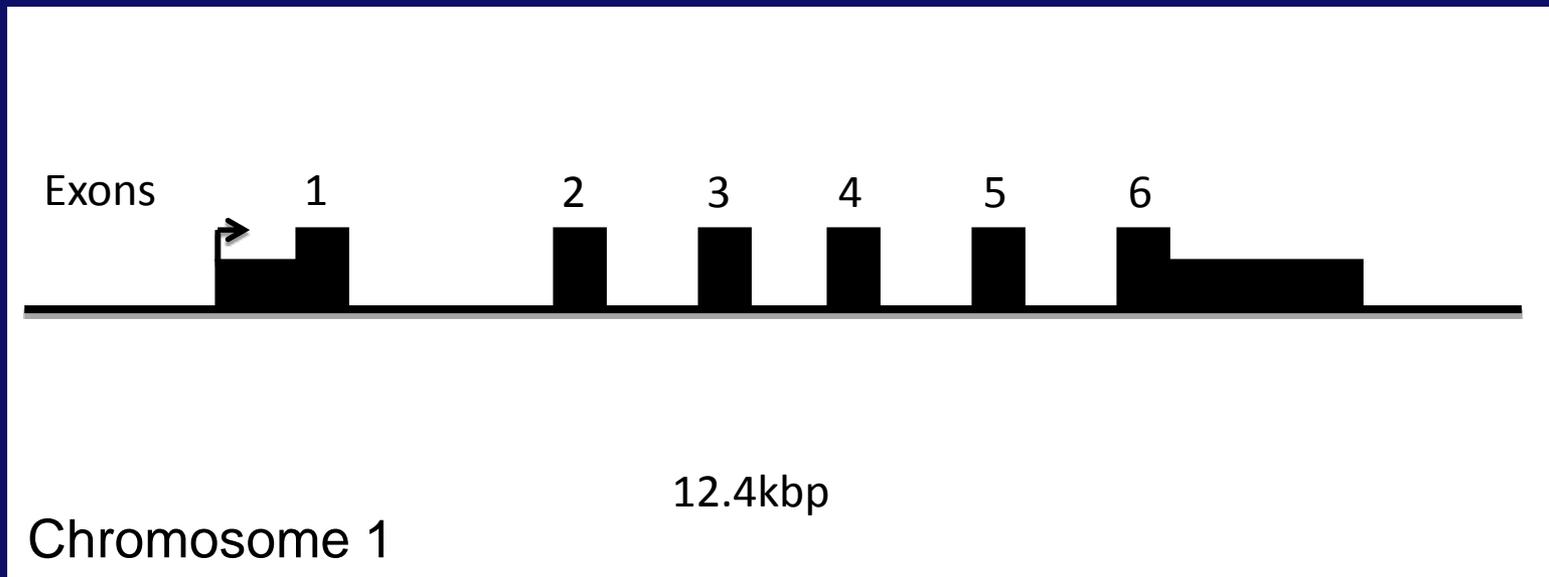
area(%)

# Possible Mechanisms of VTE in Cancer Patients



# The Human Tissue Factor Gene

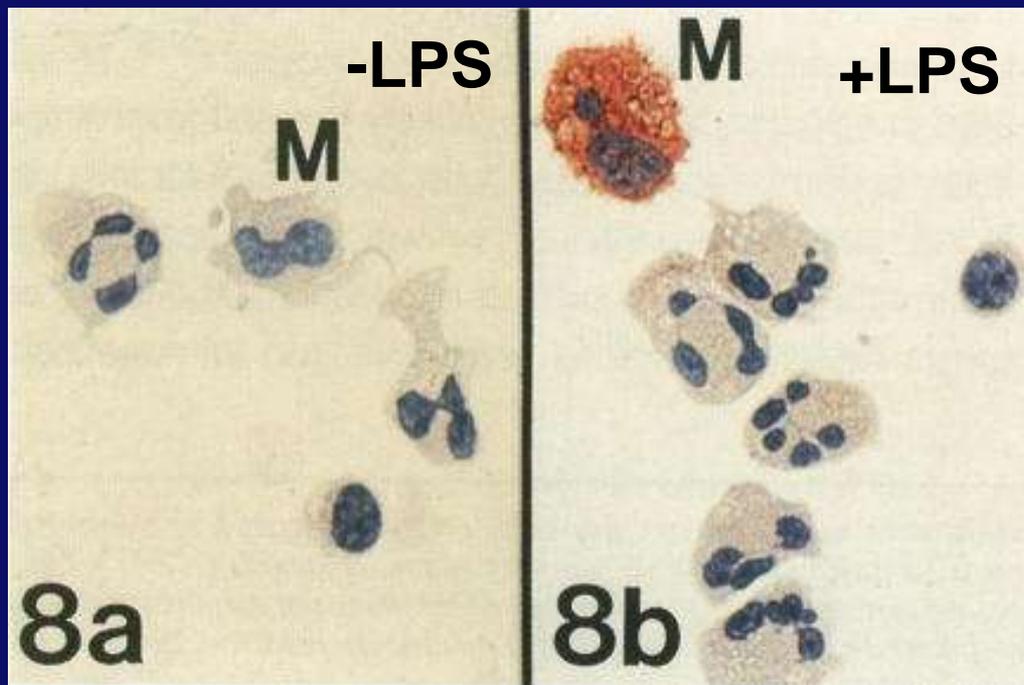
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Mackman et al Biochemistry 1989

# LPS Induction of TF Expression in Monocytes

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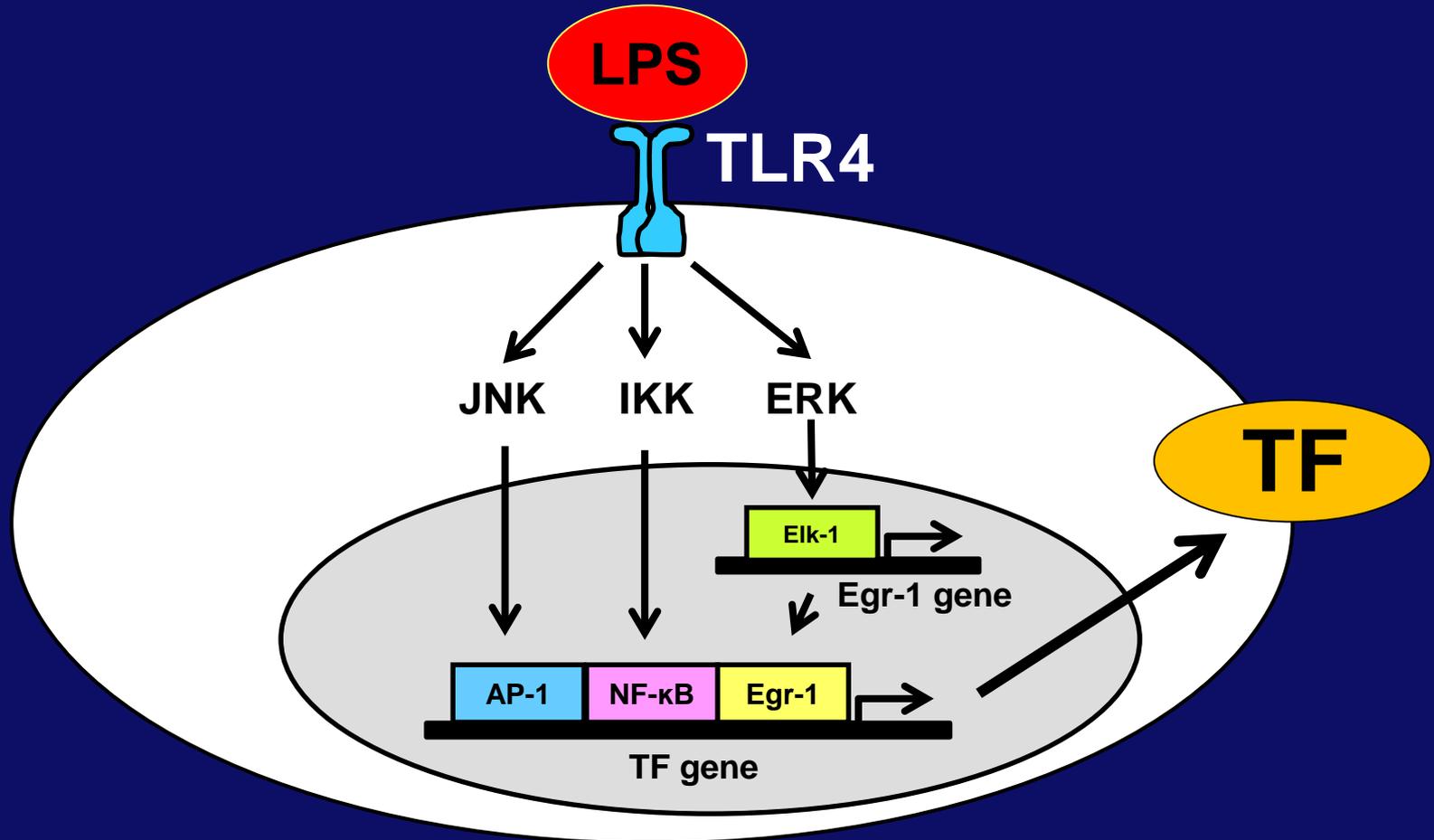


Host Defense

6 hrs

Drake et al AJP 1989

# LPS Induction of the TF Gene in Monocytic Cells

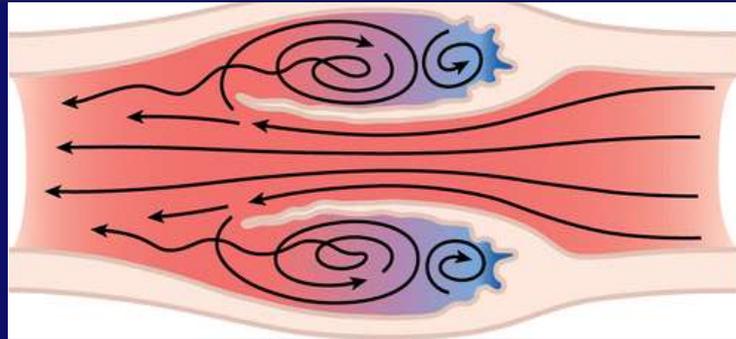


Mackman et al, JEM 1991; Mackman JBC 1994; Parry and Mackman JBC 1994; Oeth et al, MCB 1994; Guha et al, Blood 2002

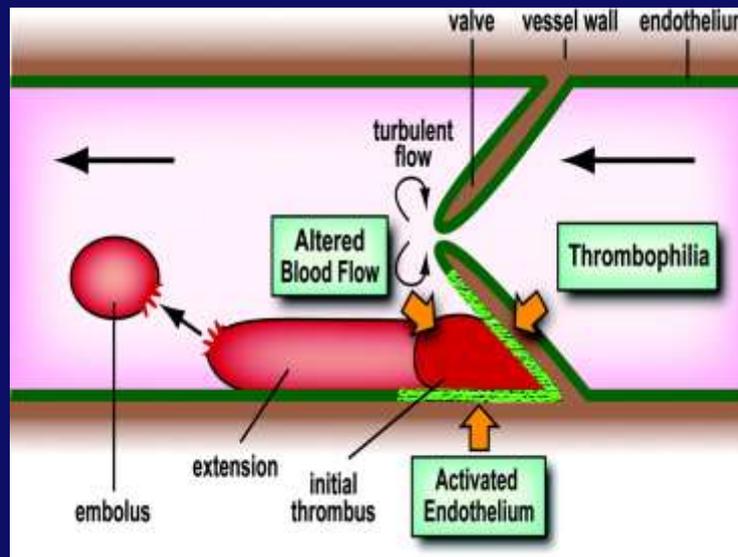


# Formation of a Deep Vein Thrombus

Hypoxia in valve pockets

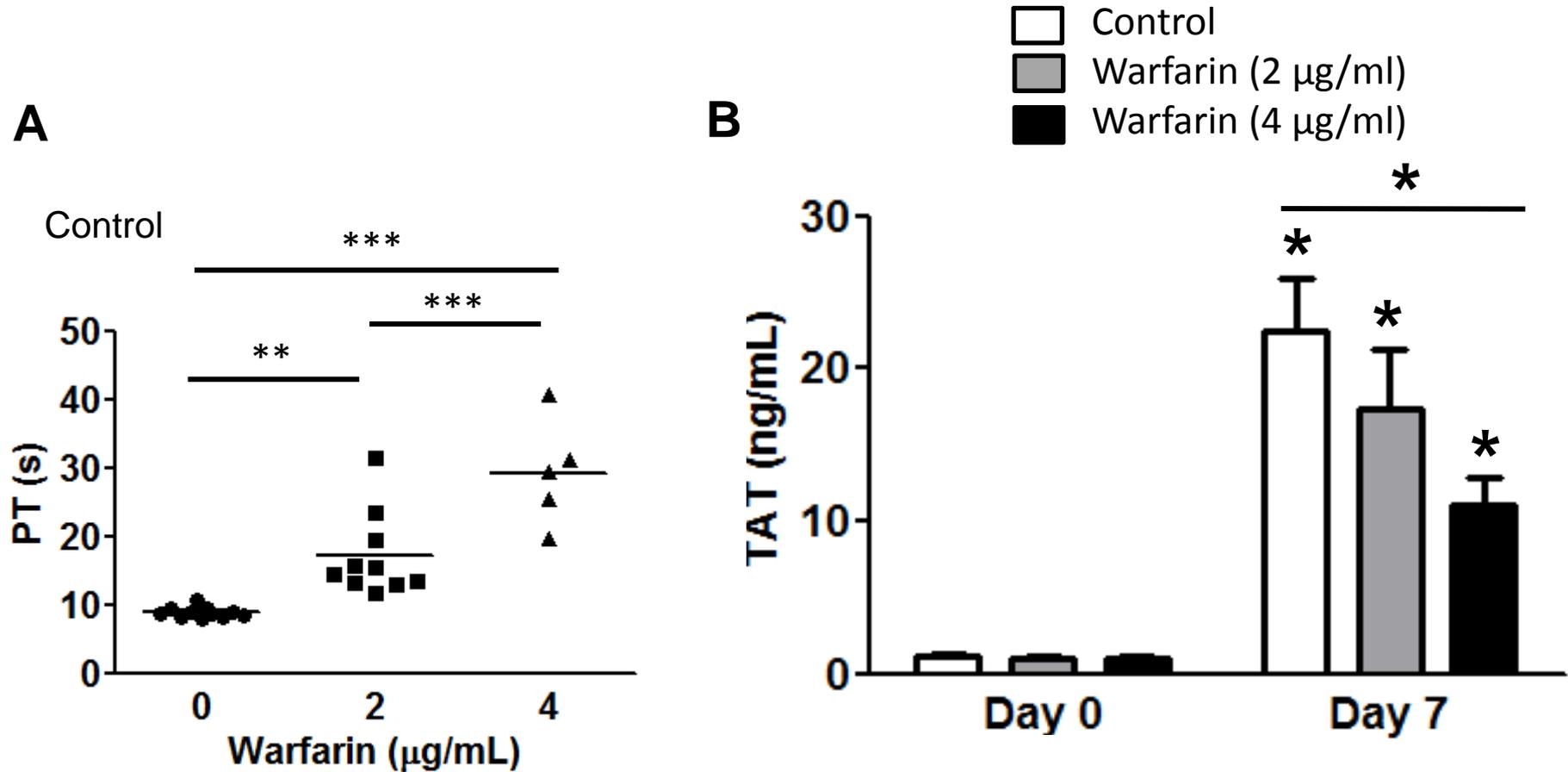


Bovill Ann Rev Physiol 2011

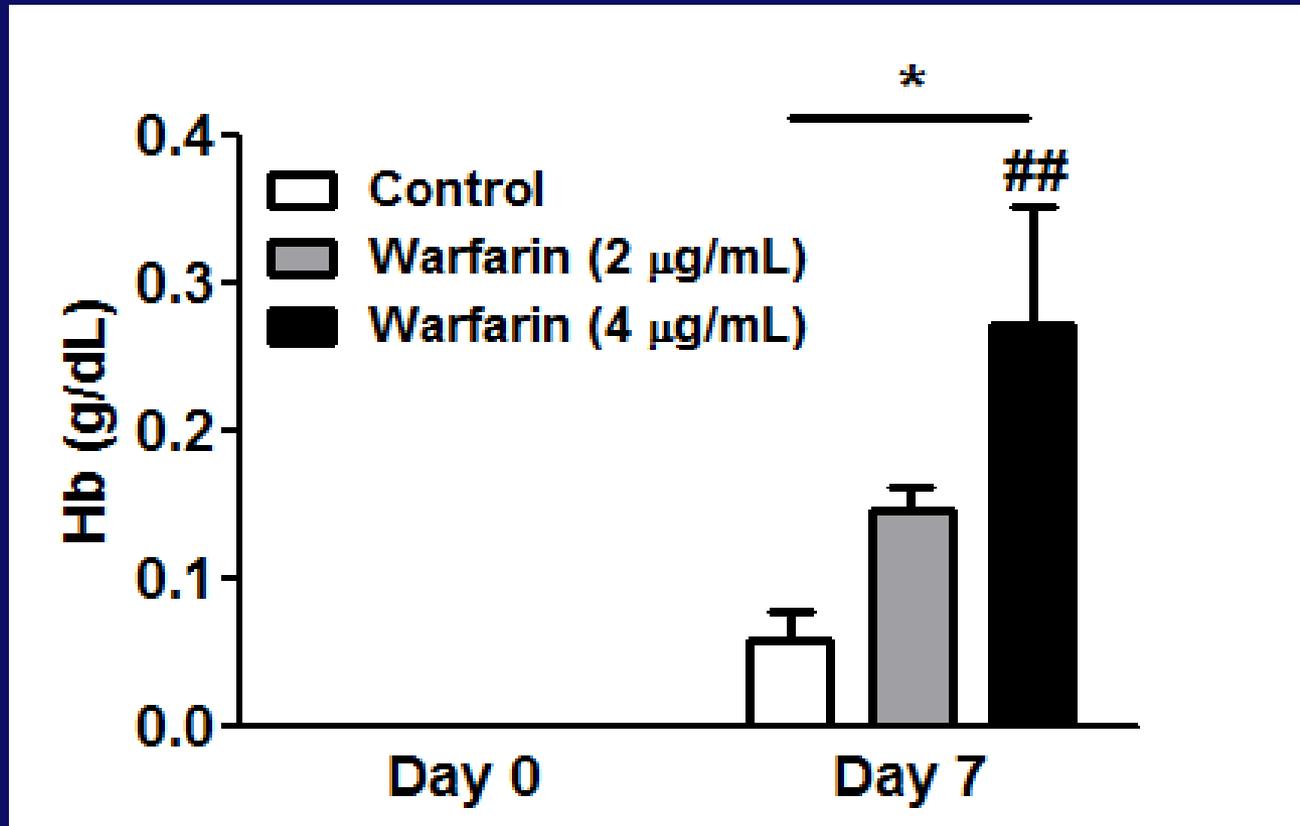


Moll and Mackman ATVB 2008

# Anticoagulating Mice with Warfarin



# Warfarin Increases Lung Hemorrhage After IAV Infection



# TF Expression in Pancreatic Cancer and VTE

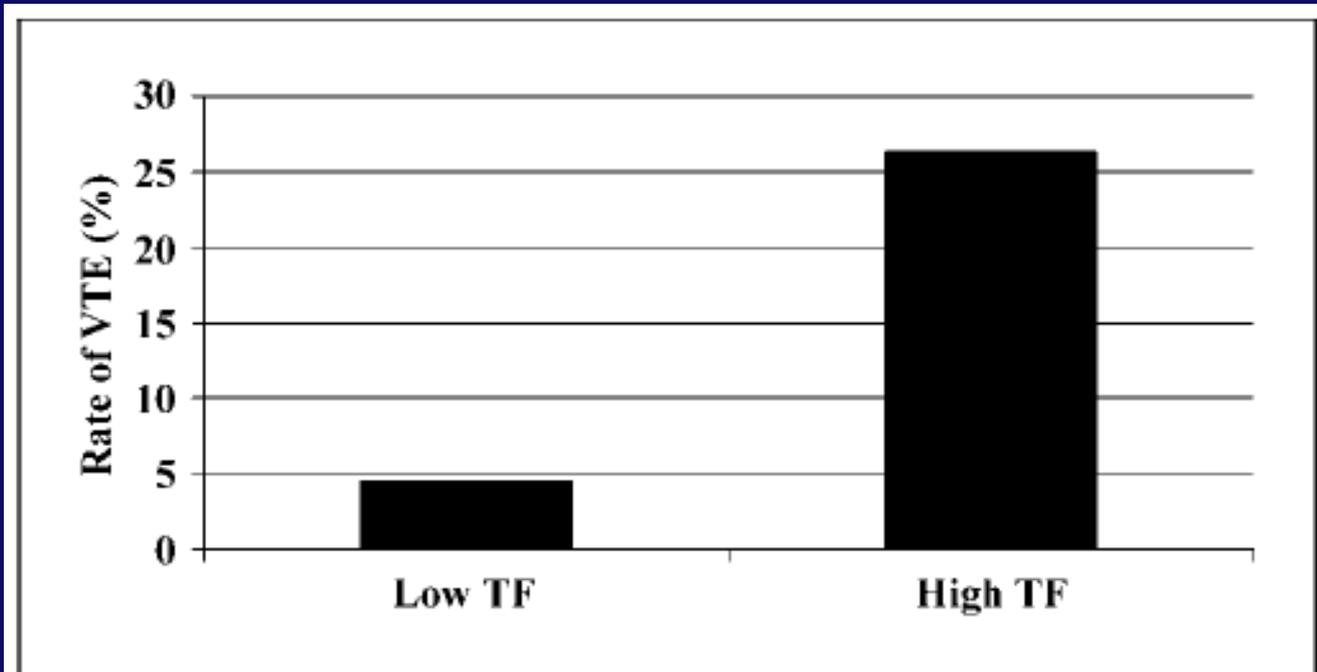
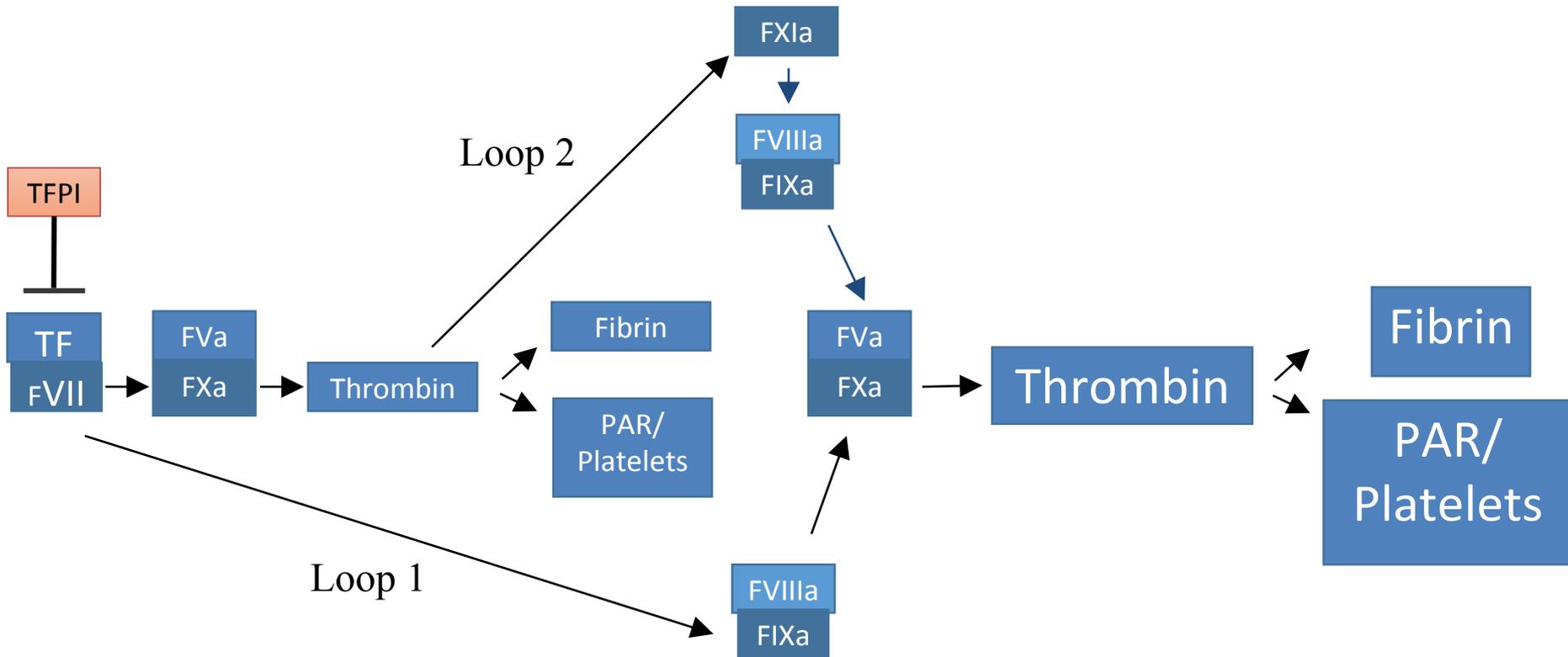


Fig. 2. TF expression and symptomatic venous thromboembolism (VTE) in pancreatic cancer. Pancreatic cancer patients with low TF expression had a venous thromboembolism rate of 4.5%, and this was elevated 4-fold to 26.3% in patients with high TF expression ( $P = 0.04$ ).

# Amplification Loops in the Clotting Cascade

Initiation

Propagation



# Effect of Crossing Two Prohemorrhagic Mice

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Can we exacerbate the bleeding in Low TF placentas and mice by adding another prohemorrhagic genetic defect?

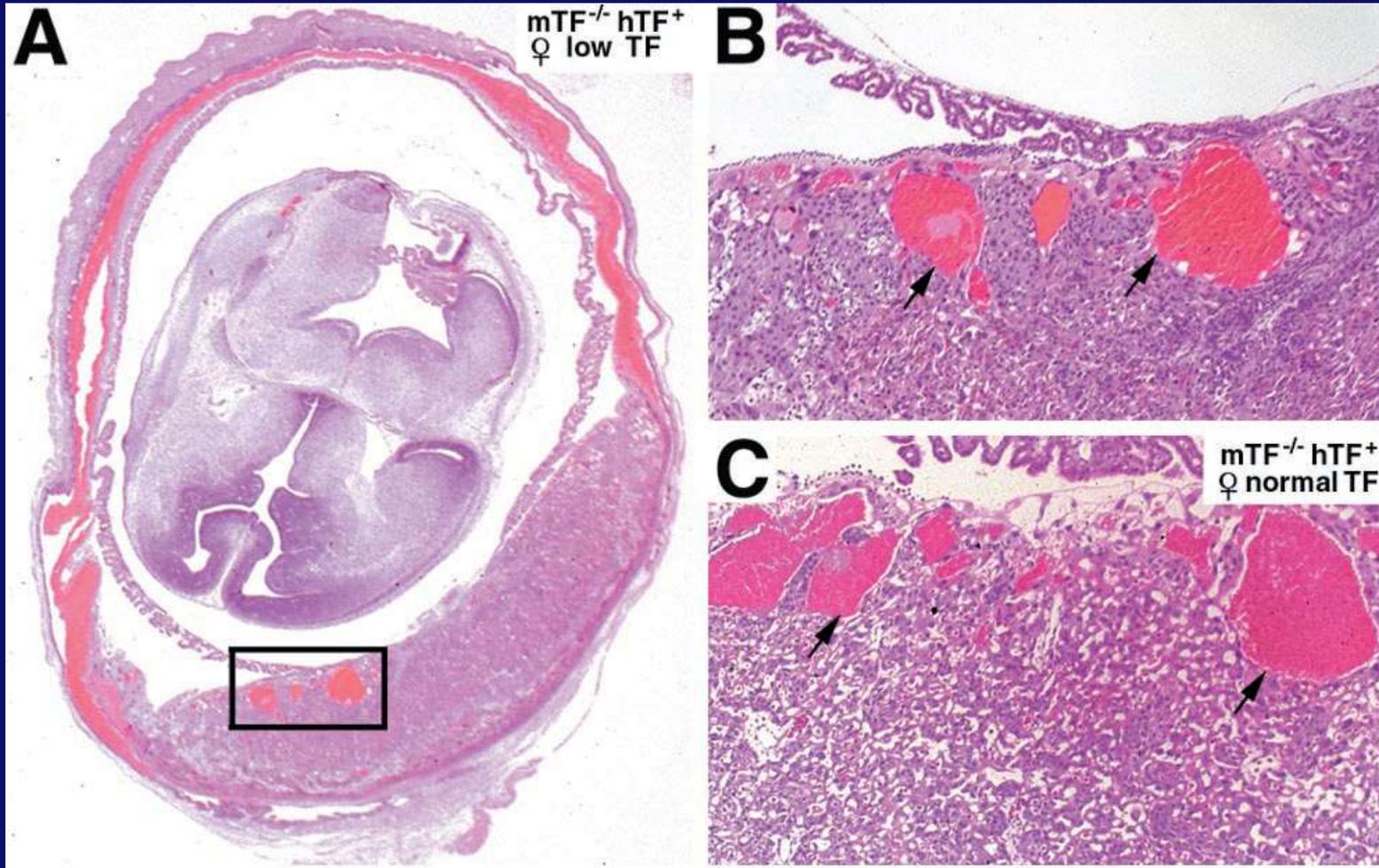
Low TF x PAR4<sup>-/-</sup> - increased fatal lung hemorrhage Bode & Mackman Thromb Res 2016

Low TF x FIX<sup>-/-</sup> - embryonic death.

Low TF x FXI<sup>-/-</sup> - expected number of mice at wean but increased placental blood pools and maternal death.

Low TF x FXII<sup>-/-</sup> - expected number of mice at wean.

# Low TF Placentas Contain Blood Pools in Low TF and Control Mothers



Erlich et al PNAS 1999

# Roles of TF in the Uterus and Placenta During Gestation and Post-Partum

Breeding strategy	Level of TF expression		Fatal mid-gest hemorrhage	Fatal post-partum hemorrhage
	Mother	Embryo		
♂ $-/-, h^+$ X ♀ $-/-, h^+$	1%	1%	17/41 (42%)	4/22 (18%)
♂ $+/+$ X ♀ $-/-, h^+$	1%	50%	0/43 (0%)	6/43 (14%)

**FVIII<sup>-/-</sup> and FIX<sup>-/-</sup> have normal pregnancies**

# Effect of Inactivation of the TFPI Gene in Mice

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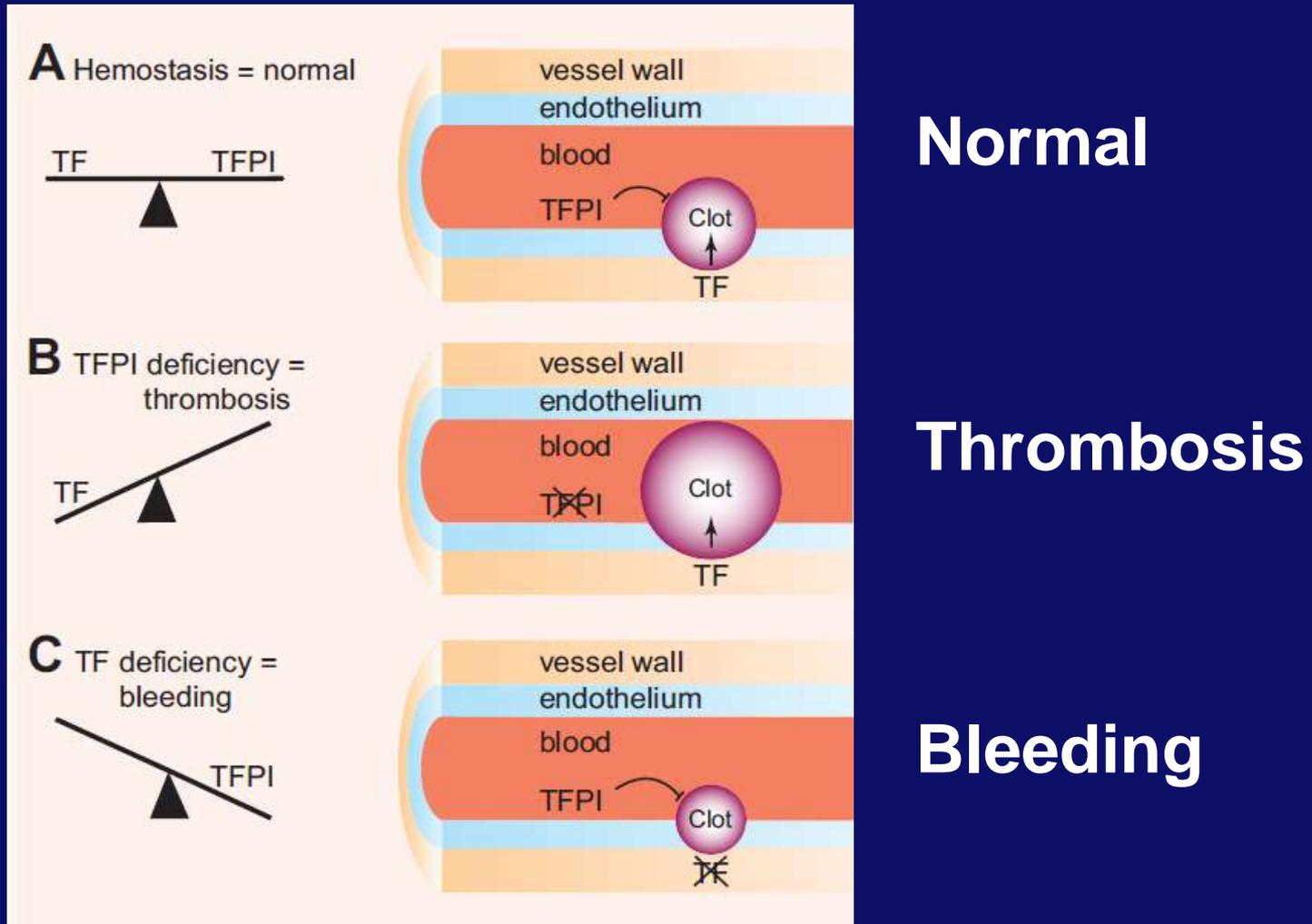
TFPI null embryos die during embryonic development



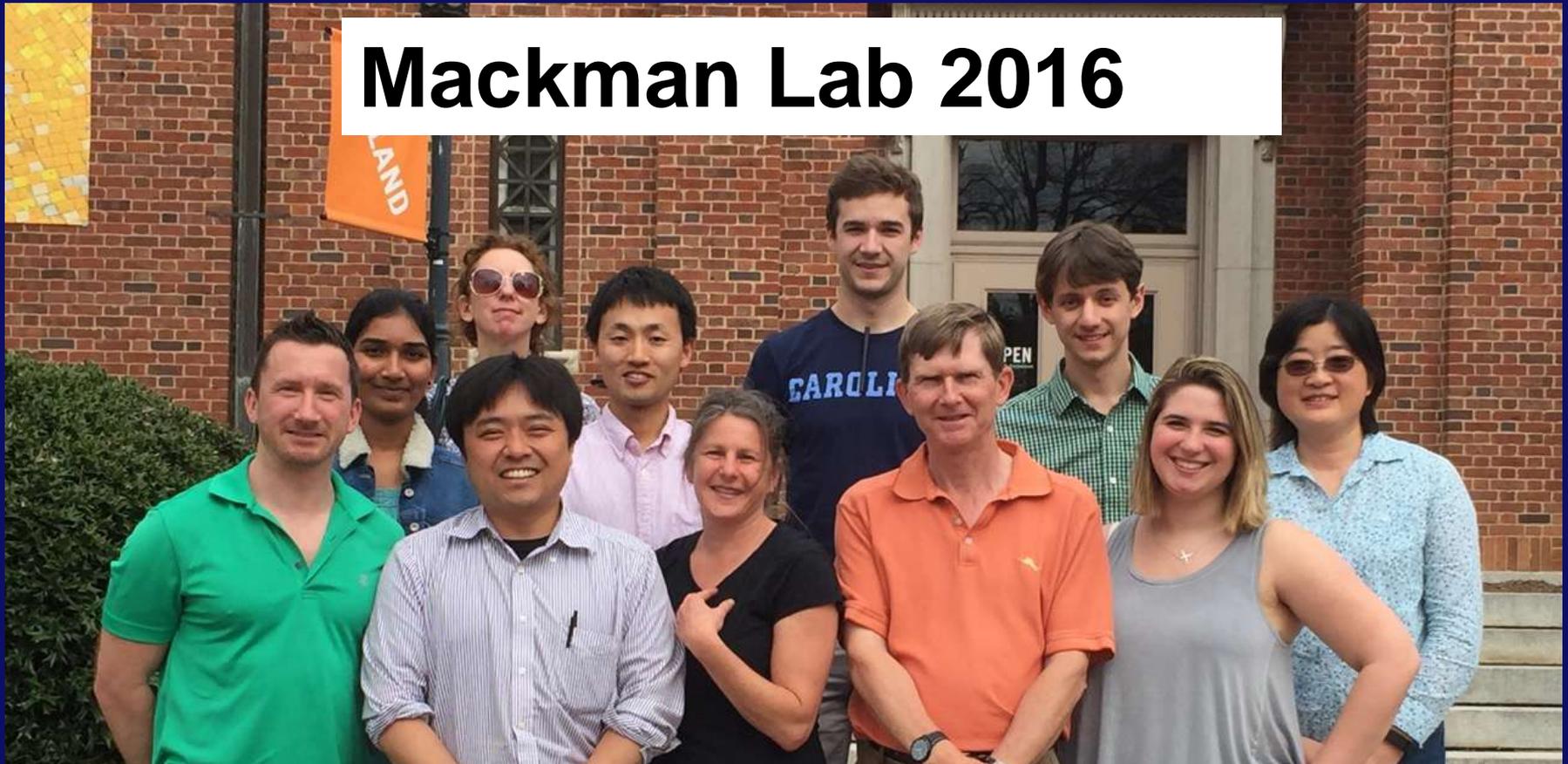
**TFPI is essential for survival**

Huang Z et al Blood 1997

# Balancing the TF Pathway



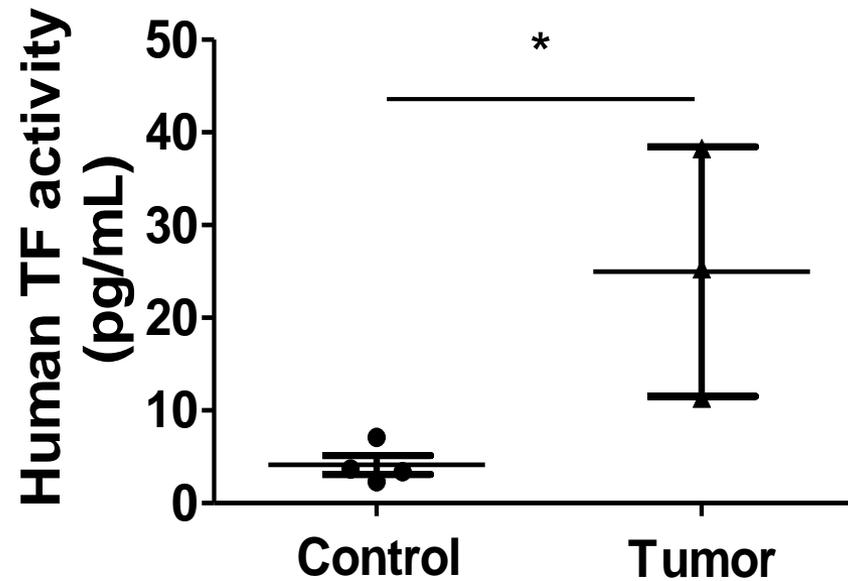
# Mackman Lab 2016



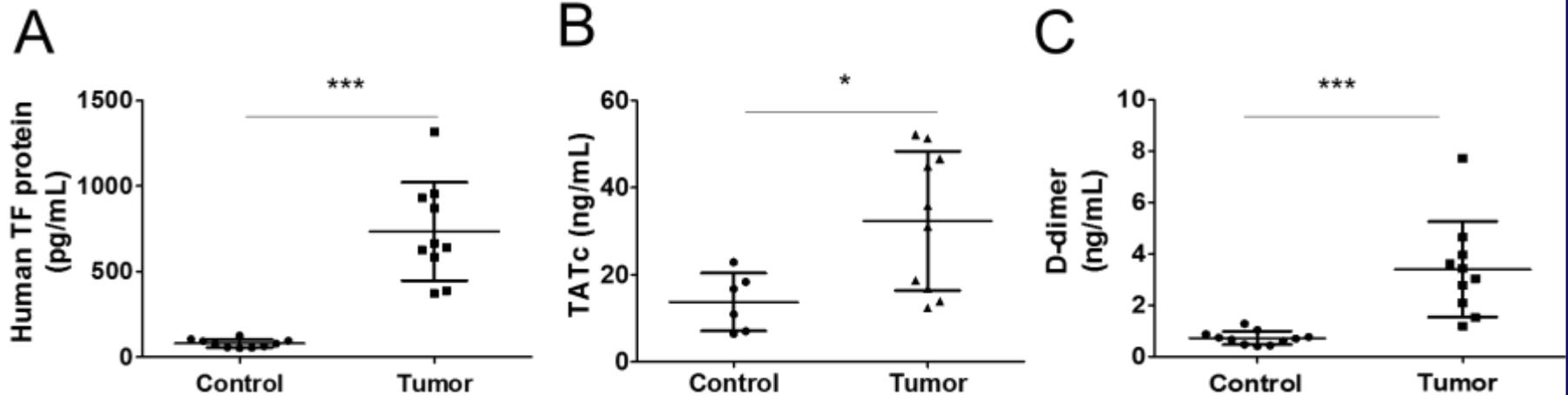
TraCS

# Tumor-derived Human TF is Present in Venous Clots in Mice

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# Levels of Human TF and D-dimer in Tumor-bearing Mice

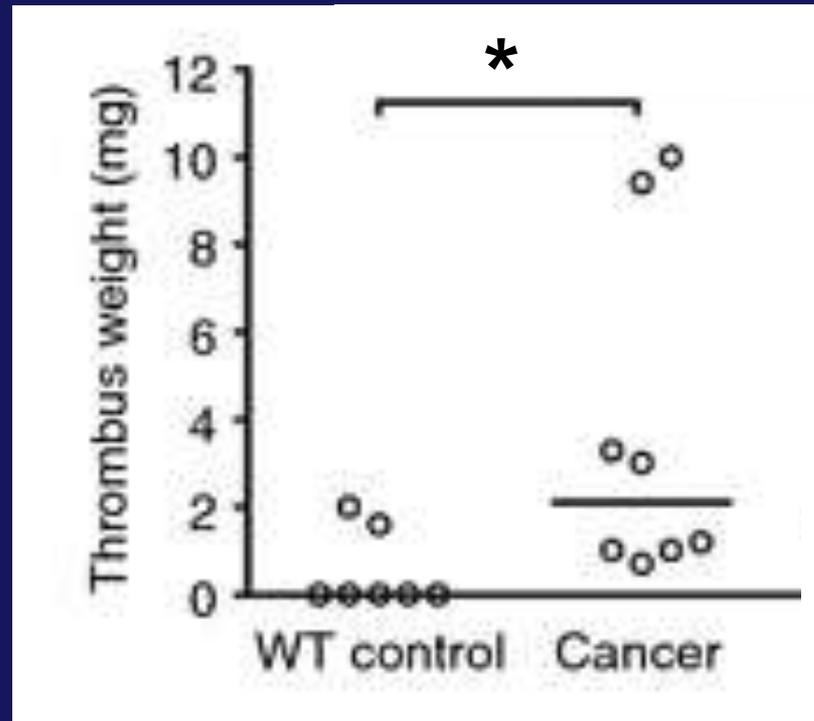


# Effect of Crossing Low TF Mice with FXI<sup>-/-</sup> Mice

FXI	mTF	hTF	N	Observed (%)	Expected (%)
+/+	+/+	-	5	7.813	1.5625
+/-	+/+	-	1	1.563	3.125
-/-	+/+	-	2	3.125	1.5625
+/+	+/+	+	6	9.375	4.6875
+/-	+/+	+	1	1.563	9.375
-/-	+/+	+	0	0.000	4.6875
+/+	+/-	-	1	1.563	3.125
+/-	+/-	-	6	9.375	6.25
-/-	+/-	-	1	1.563	3.125
+/+	+/-	+	8	12.500	9.375
+/-	+/-	+	14	21.875	18.75
-/-	+/-	+	6	9.375	9.375
+/+	-/-	-	0	0.000	1.5625
+/-	-/-	-	0	0.000	3.125
-/-	-/-	-	0	0.000	1.5625
+/+	-/-	+	5	7.813	4.6875
+/-	-/-	+	4	6.250	9.375
-/-	-/-	+	4	6.250	4.6875
Total			64	100	100

# Thrombosis is Increased by the Presence of a Mouse Pancreatic Tumor

Clot measured at 3 h

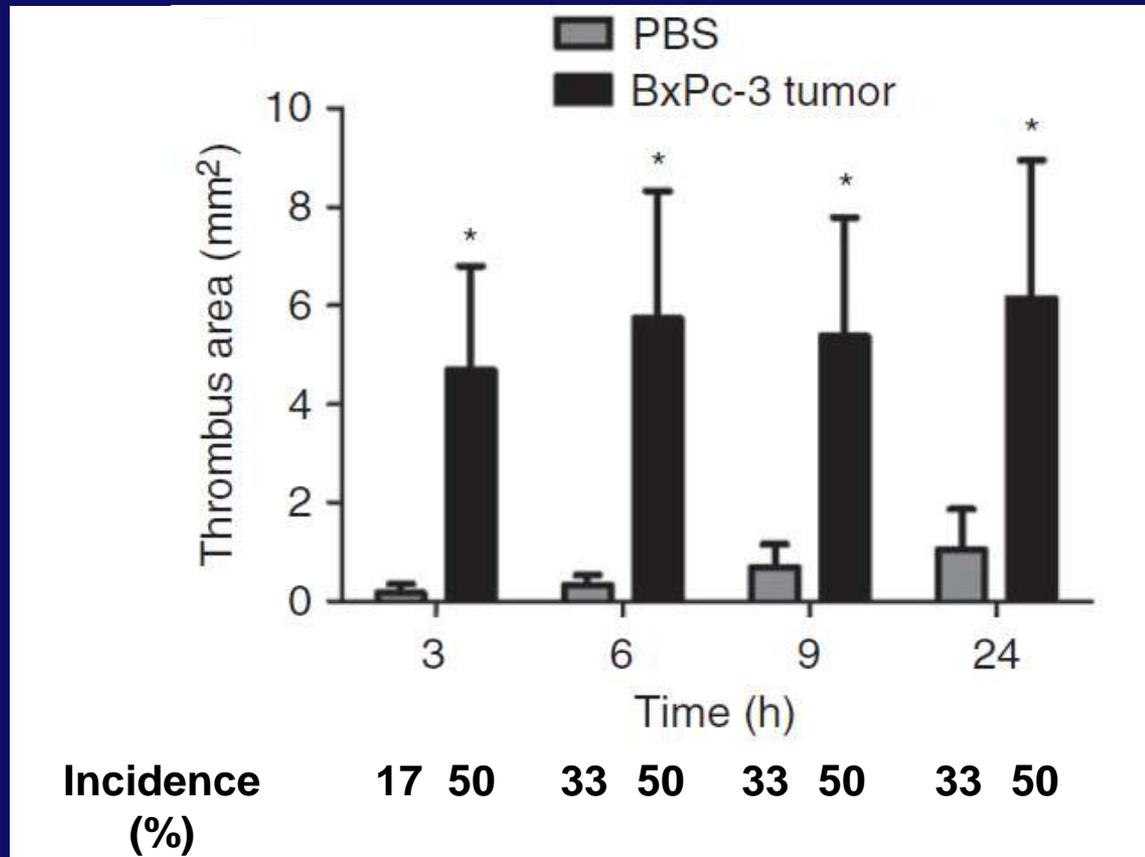


Panc02 is a mouse pancreatic cancer cell line

# Thrombosis is Increased by the Presence of Human Pancreatic Tumors Grown in Nude Mice



**Julia  
Geddings**



**BxPc-3 is a human pancreatic tumor**

**Geddings J et al JTH 2016**

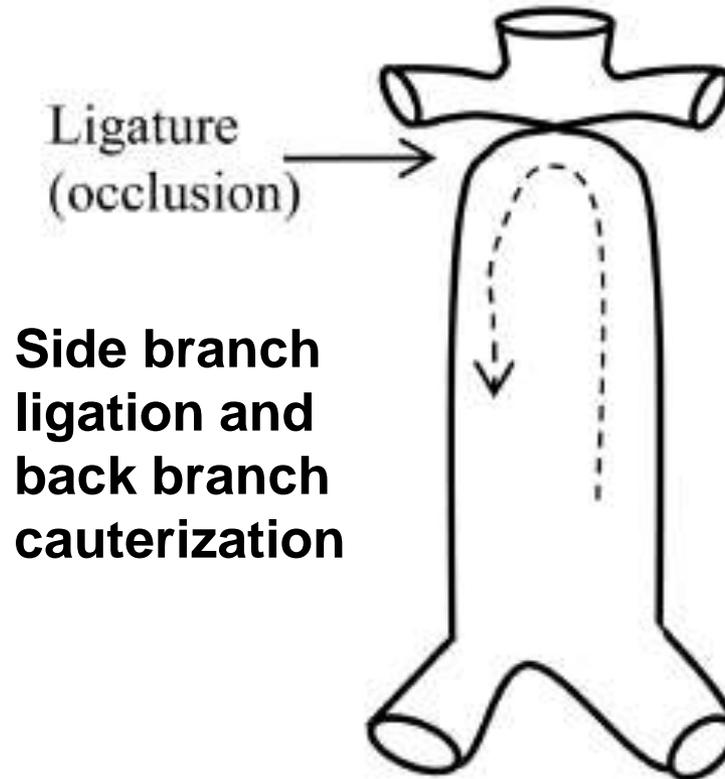
# Limitations of the Current Studies

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- **The incidence and clot size in the IVC stenosis model is quite variable.**
- **These studies do not specifically analyze the role of tumor-derived, TF<sup>+</sup> MVs in thrombosis.**

# IVC Stasis Model

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**The incidence is ~100% and the  
clot size is ~20 mg at 48 h**

# TF Expression in Pancreatic Cancer and VTE

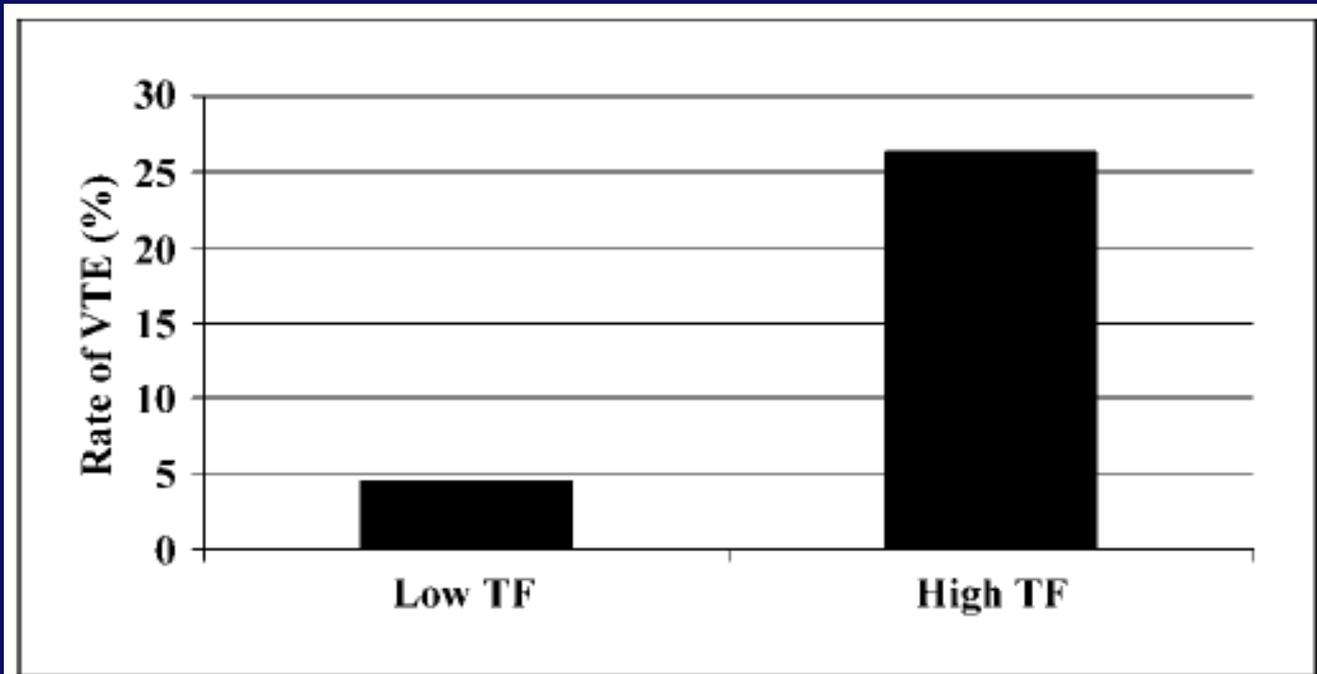
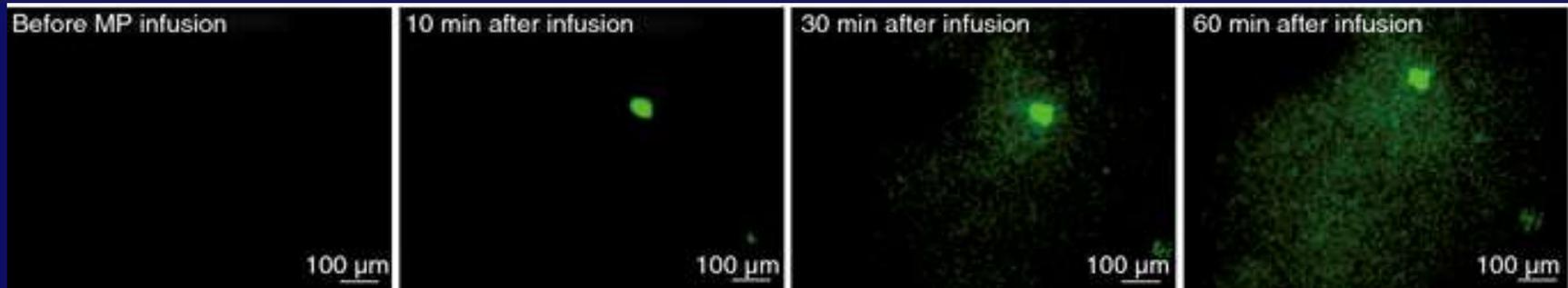


Fig. 2. TF expression and symptomatic venous thromboembolism (VTE) in pancreatic cancer. Pancreatic cancer patients with low TF expression had a venous thromboembolism rate of 4.5%, and this was elevated 4-fold to 26.3% in patients with high TF expression ( $P = 0.04$ ).

**Effect of injection of  
exogenous tumor-derived,  
TF<sup>+</sup> MVs on venous  
thrombosis in mice**

# Injected Tumor MVs Localize to the Site of Thrombosis in an IVC Stenosis Model

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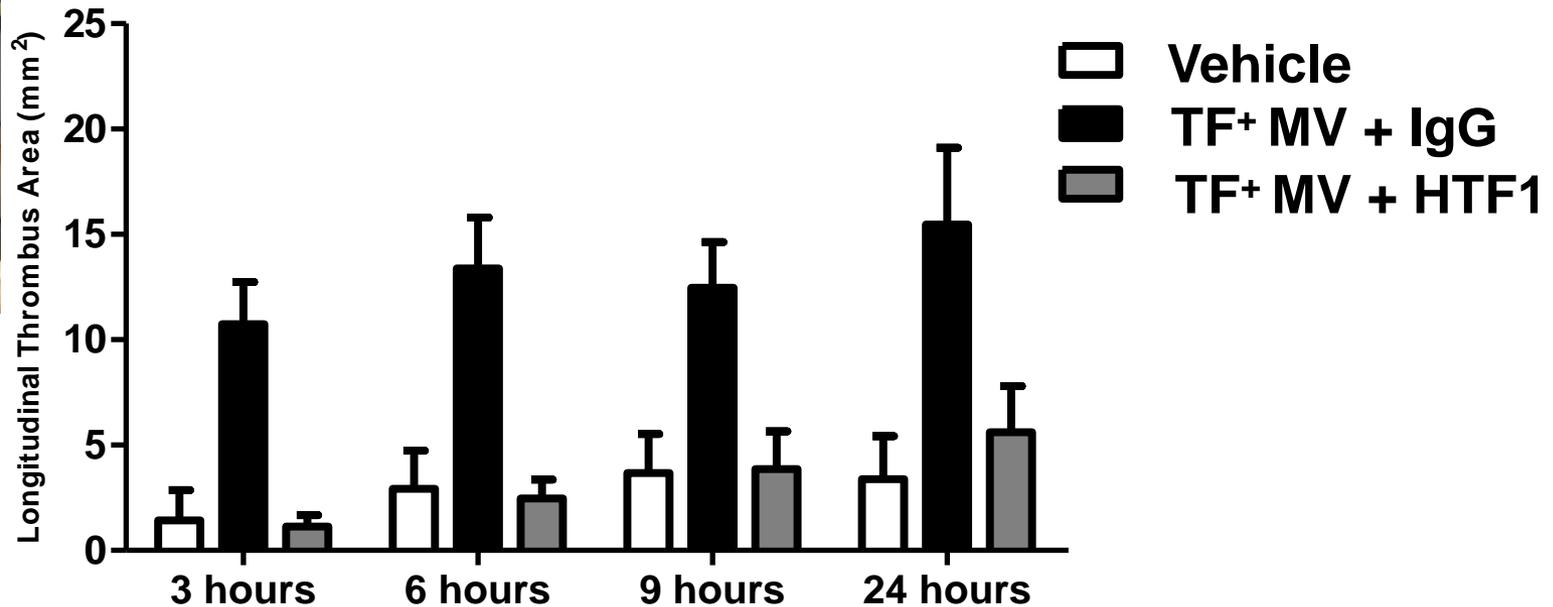


**Panc02 = mouse pancreatic cancer cell line**

# Exogenous TF<sup>+</sup> Tumor MVs Enhance Thrombosis in the IVC Stenosis Model

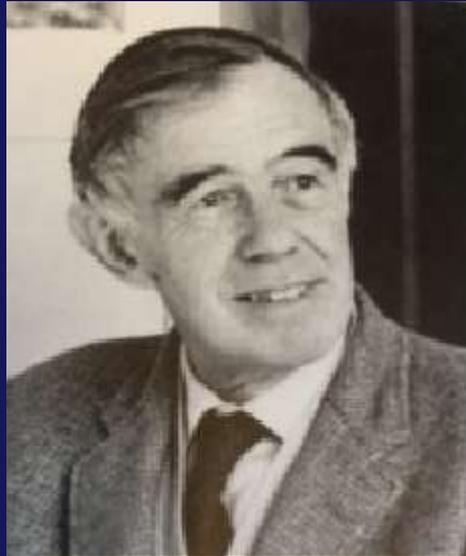


Julia  
Geddings



MVs isolated from a human pancreatic tumor cell line BxPc-3

**TF on tumor MVs is required for the enhancement of thrombosis**



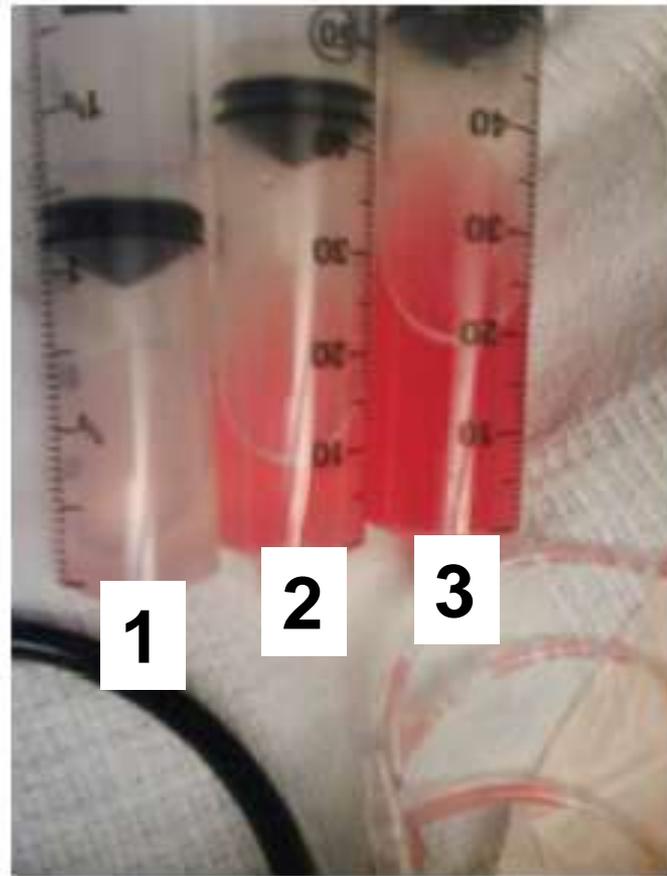
**Robert G MacFarlane**  
**(1907-1987)**



**Rosemary Biggs**  
**(1912-2001)**

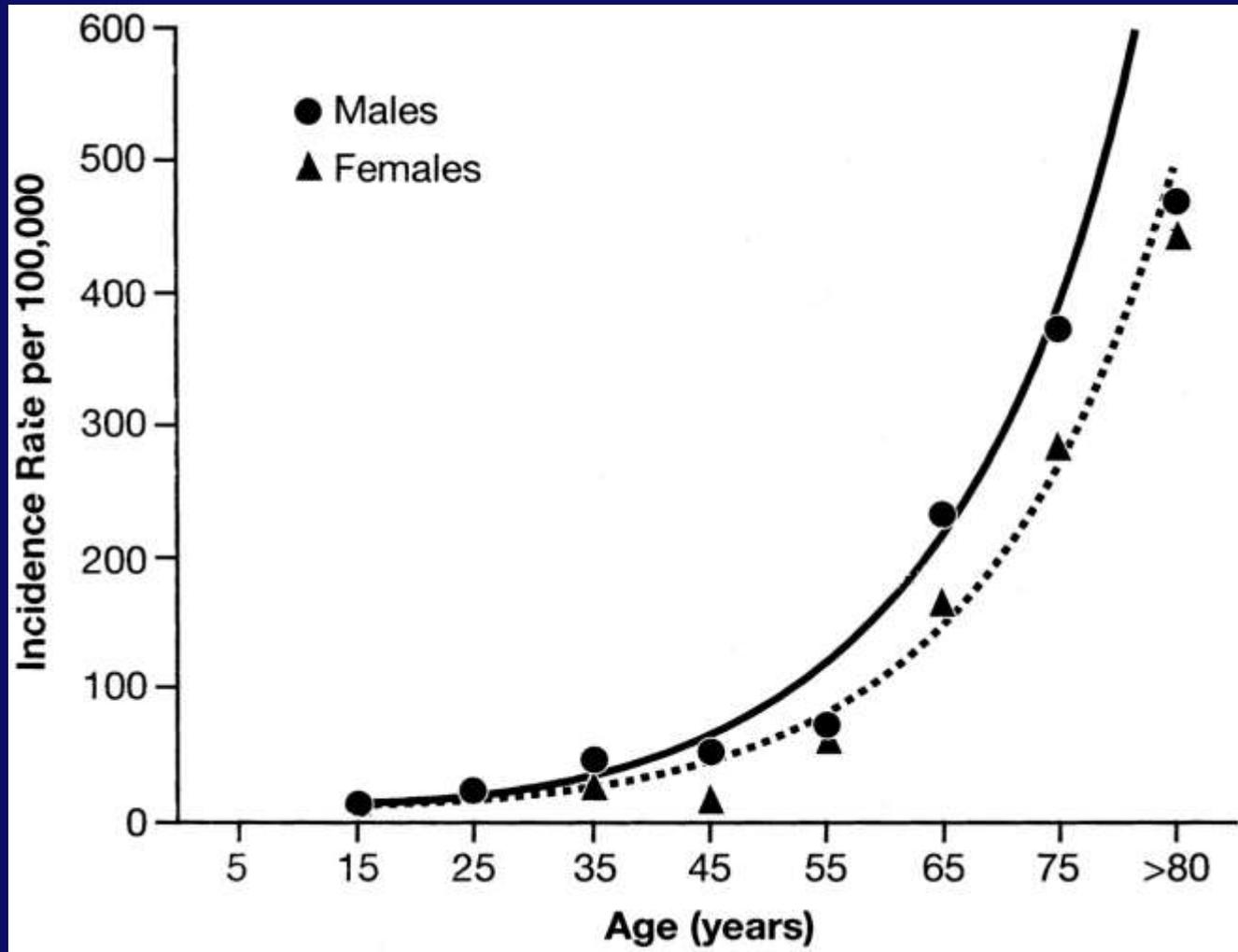
# Alveolar Hemorrhage in Patients with Severe Influenza Infection

Bronchiolar lavage fluid from a patient infected with influenza A





# Annual Incidence of VTE by Age and Sex



# Risk Factors for Venous Thrombosis

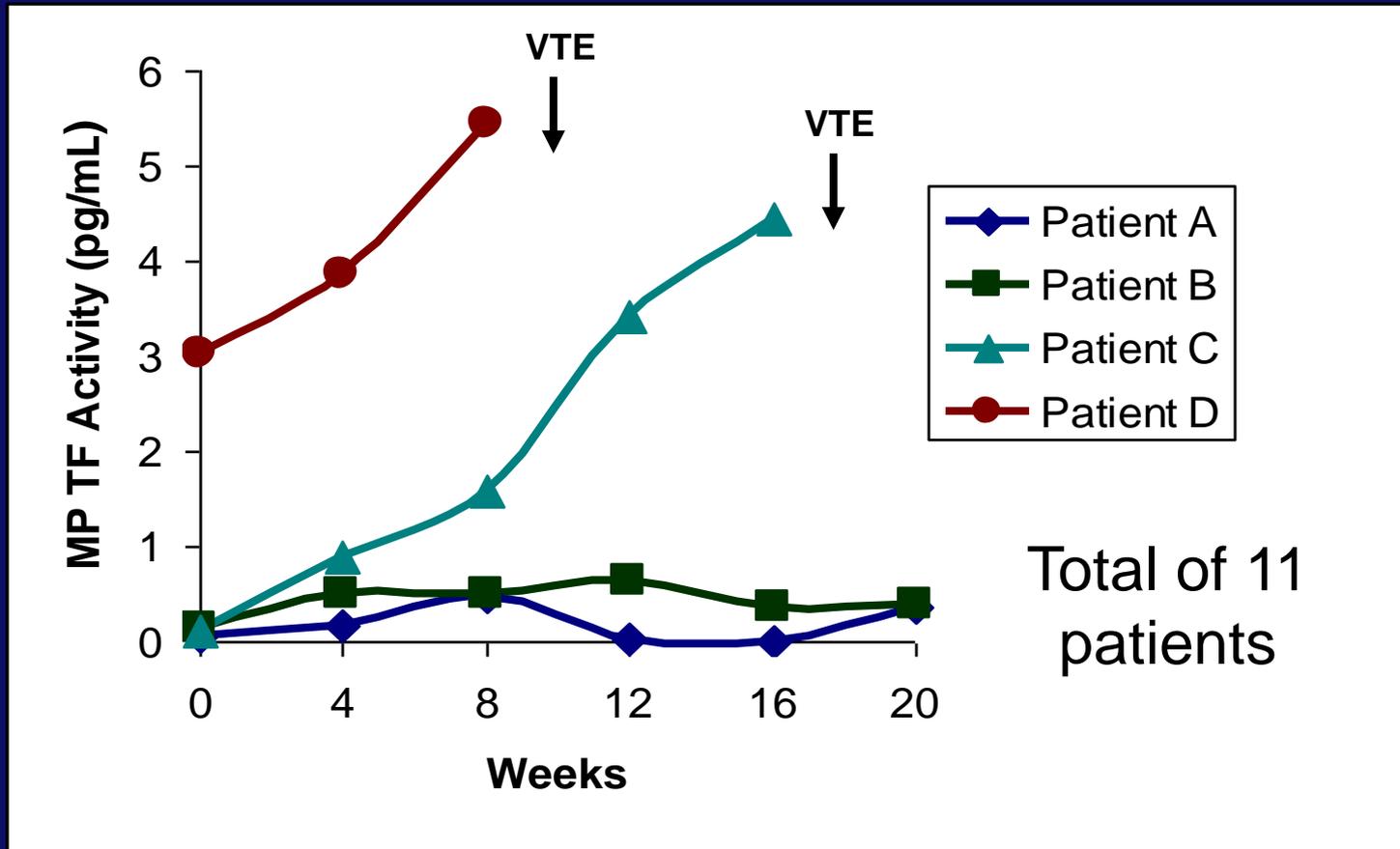
## ACCP Practice Guidelines

Padua Prediction Risk Score – high risk defined as a score  $\geq 4$

<b>Risk Score</b>	<b>Points</b>
Active cancer	3
Previous VTE	3
Reduced Mobility (bed rest $\geq 3$ days)	3
Known thrombophilic condition	3
Recent trauma and/or surgery	2
Elderly age ( $\geq 70$ years)	1
Heart and/or respiratory failure	1
Acute MI or ischemic stroke	1
Obesity (BMI $\geq 30$ )	1
Ongoing hormone treatment	1

Barbar S et al JTH 2010- Identification of hospitalized medical patients at risk for VTE

# Levels of MV TF Activity Increased in Pancreatic Cancer Patients before VTE

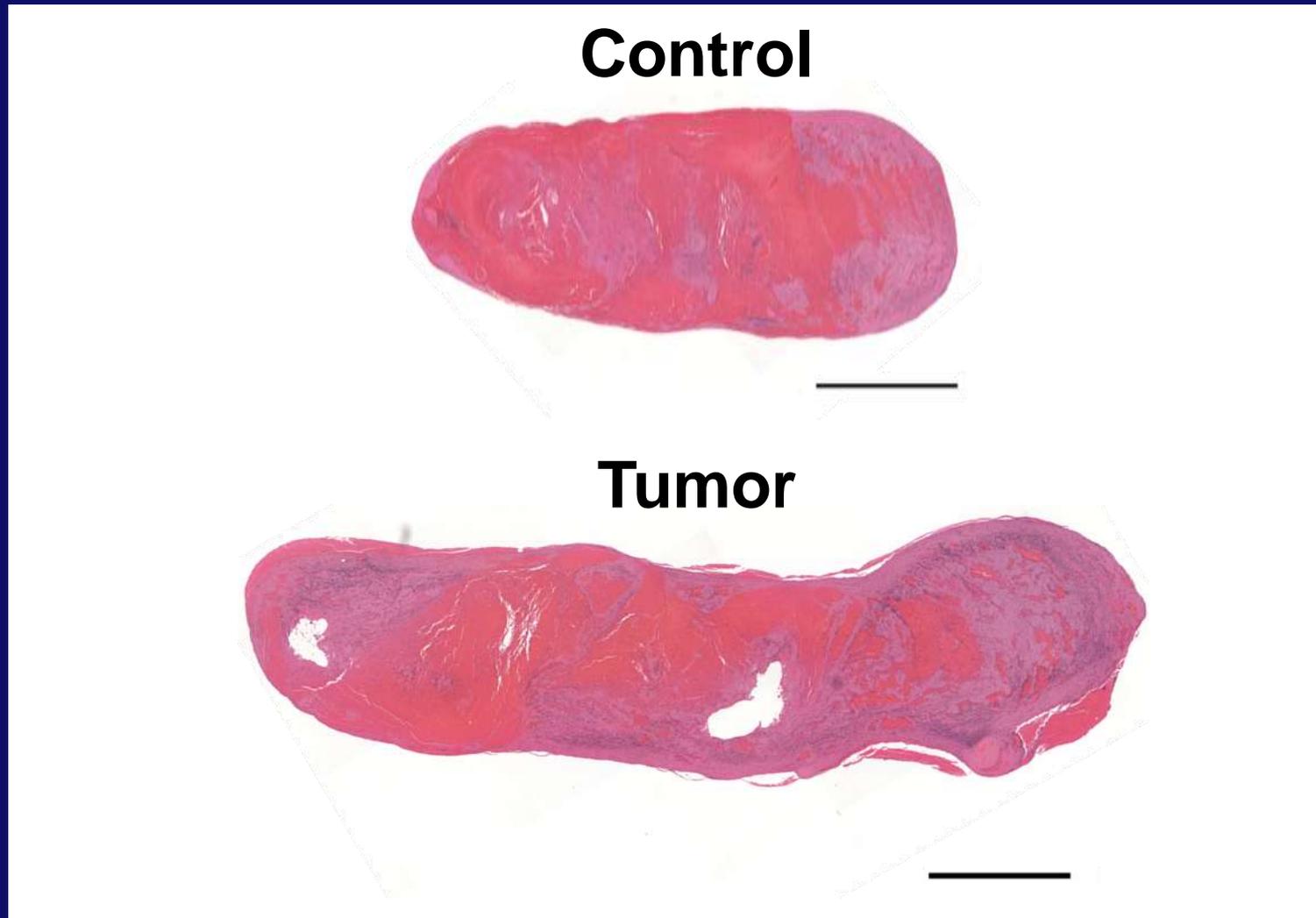


# TF+ MPs and VTE in Cancer Patients: Prospective Studies

Study	Method	Patients	Time of Follow-up	Association between MP TF and VTE?
<b>Khorana, 2008</b>	MP TF activity TF ELISA	2 pancreatic cancer patients who develop VTE 8 pancreatic cancer patients without VTE	Every 4 weeks for 20 weeks	Yes
<b>Zwicker, 2009</b>	impedance-based flow cytometry	4 cancer patients who develop VTE 56 cancer patients without VTE	1 year	Yes
<b>Bharthuar, 2010</b>	MP TF activity	52 Pancreaticobiliary cancer patients with VTE 65 Pancreaticobilliary cancer patients without VTE	6 months	Yes
<b>Van Doormaal, 2012</b> <a href="#">64</a>	MP TF activity MP-dependent fibrin generation flow cytometry TF ELISA	5 cancer patients who develop VTE (3 pancreatic) 38 cancer patients without VTE (10 pancreatic)	6 months	Yes by functional assay No by antigen assay
<b>Thaler, 2012</b> <a href="#">60</a>	MP TF activity	49 cancer patients who develop VTE (12 pancreatic) 299 cancer patients without VTE (48 pancreatic)	2 years	No

# Clots from Tumor-bearing Mice are Larger but have a Similar Composition

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# **Models to Study the Role of Tumor-derived MVs in Thrombosis in Mice**

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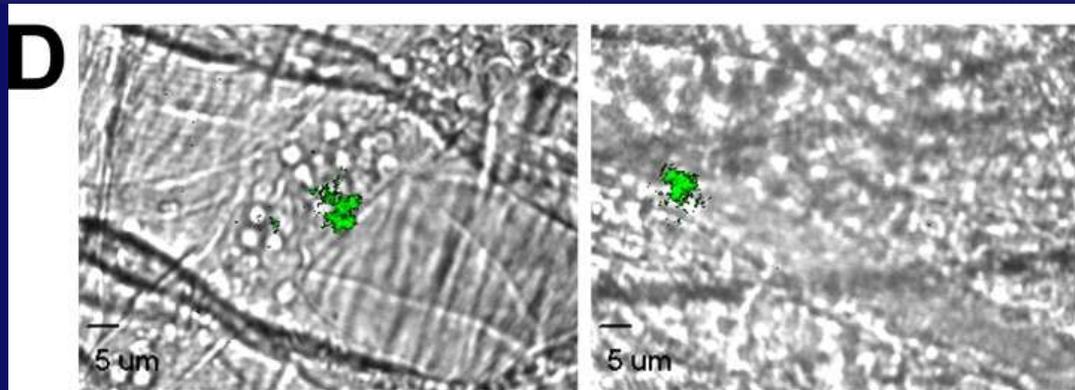
**Injection of exogenous tumor MVs injected into healthy mice**  
**Endogenous MVs in tumor-bearing mice**

**Different mouse models of thrombosis- small (mesenteric) versus large (inferior vena cava) vessels**

# Tumor-derived MPs Accumulate at the Site of Thrombus Formation In Vivo

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Mouse Panc02 pancreatic cells labeled with pEGFP



Venule

Arteriole

Thomas et al JEM 2009

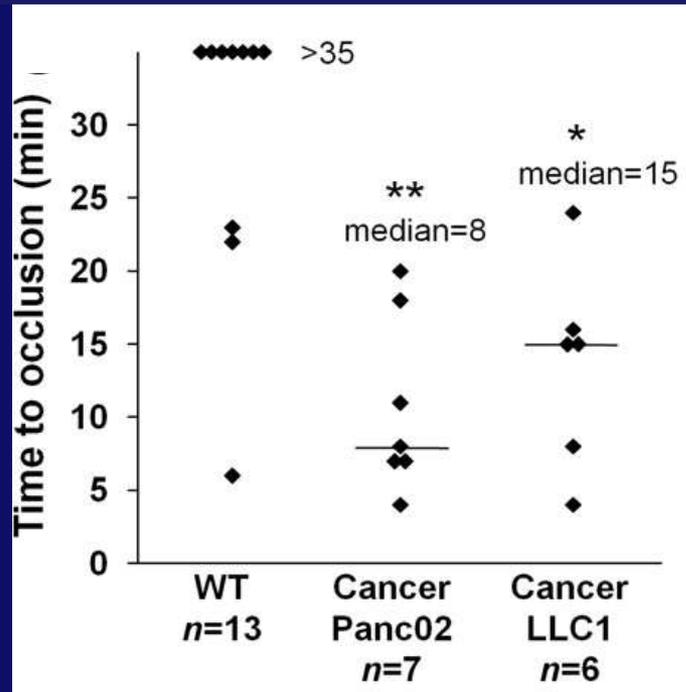
JEM

# Tumor-bearing Mice have Shorter Occlusion Times Larger in a Mesenteric Vessel Model

Panc02 = mouse  
pancreatic carcinoma

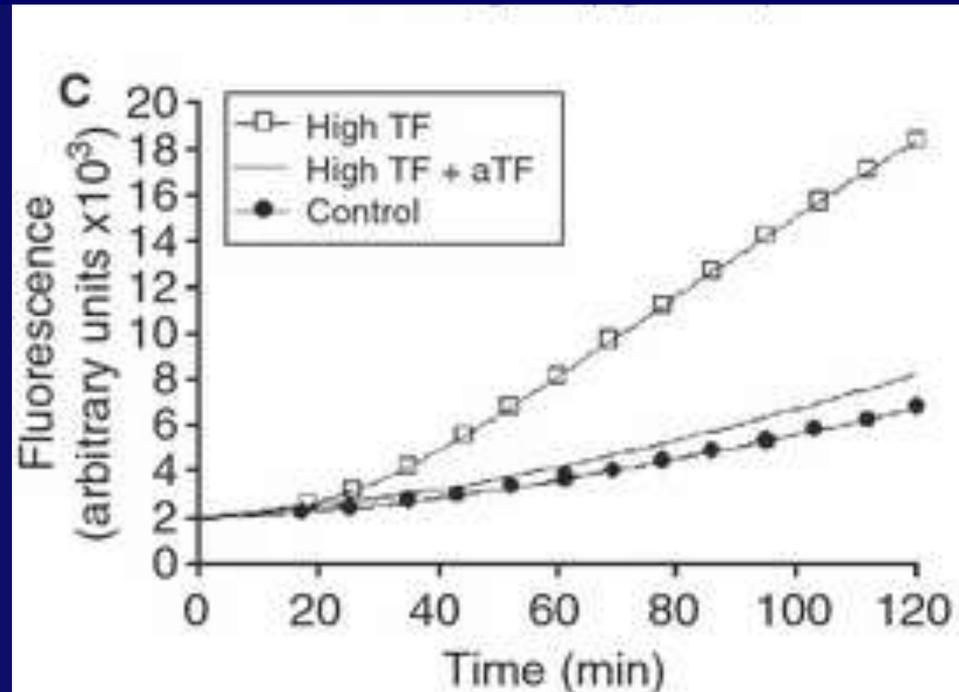
LLC = mouse lewis  
lung carcinoma

S.C.



Thomas et al JEM 2009

# Tumor-derived Human TF is Functional in an In Vitro Assay



Human pancreatic L3.6pl cells grown orthotopically in nude mice

Davila et al JTH 2008

# Conclusions

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- Human pancreatic tumors grown orthotopically released TF<sup>+</sup> MVs into the blood
- Tumor-bearing mice have bigger clots than controls
- Human TF is present in clots from tumor-bearing mice
- Inhibition of human TF reduces thrombosis

# Conclusions

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**Human pancreatic tumors grown orthotopically released TF<sup>+</sup> MVs into the blood**

**Human TF is present in clots from tumor-bearing mice**

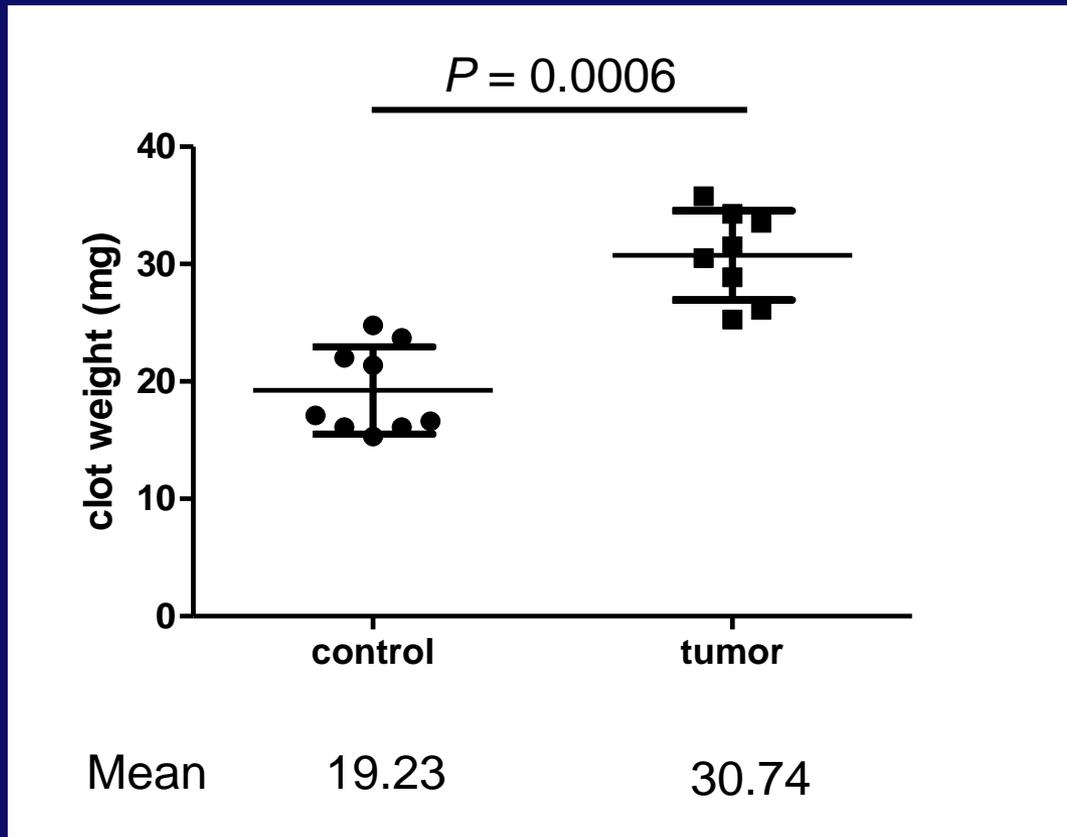
**Inhibition of human TF reduces thrombosis suggesting that tumor-derived, human TF enhances thrombosis**

# Myocardial Infarction

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# Human Pancreatic Tumors Increase Thrombosis in Nude Mice



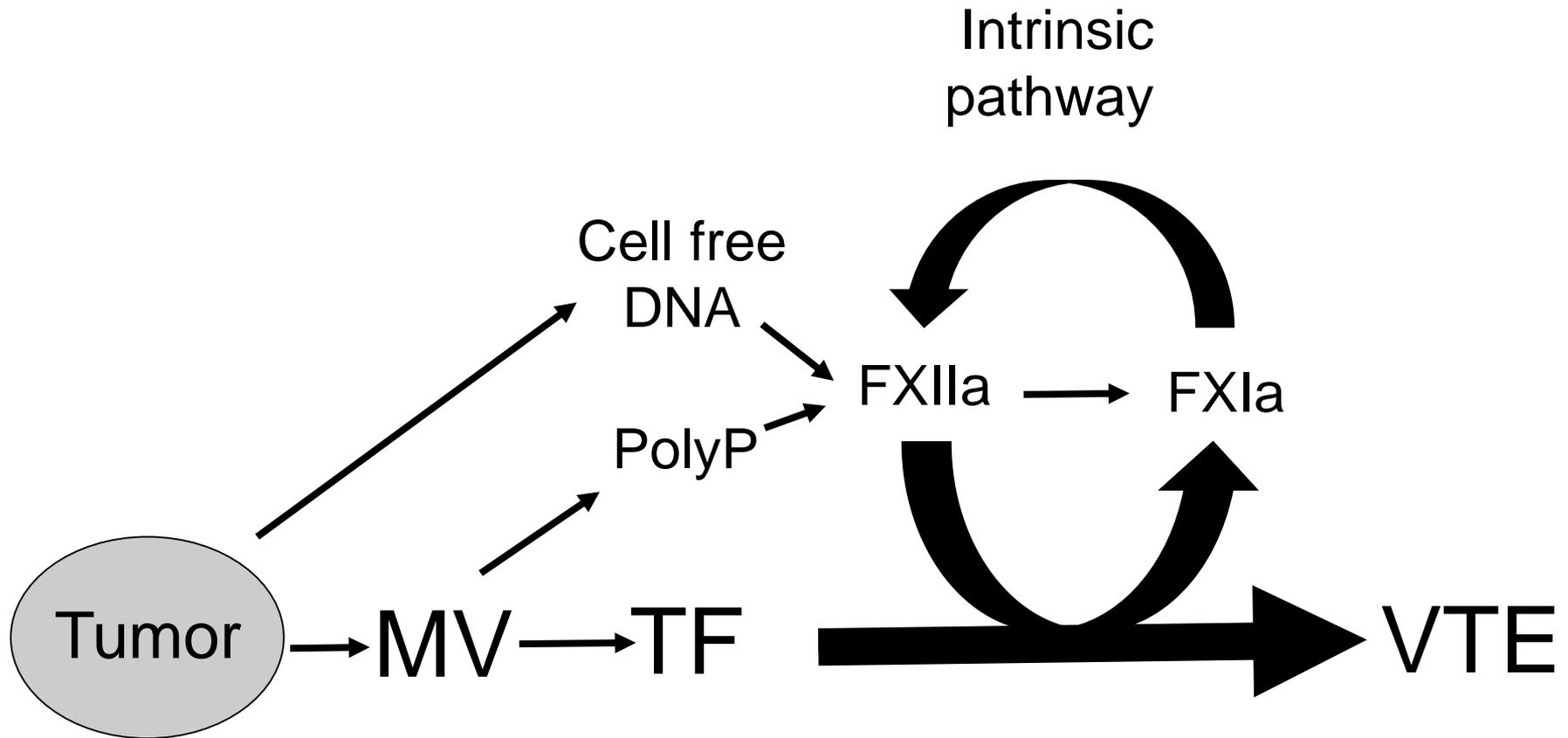
The ant-human TF antibody HTF-1 reduces thrombosis

IVC ligation model- 48 h

(Brian Cooley- Advanced rodent surgery Core)

Hisada Y et al unpublished data

# Role of the Intrinsic Coagulation Pathway in Cancer Associated Thrombosis





# Conclusion

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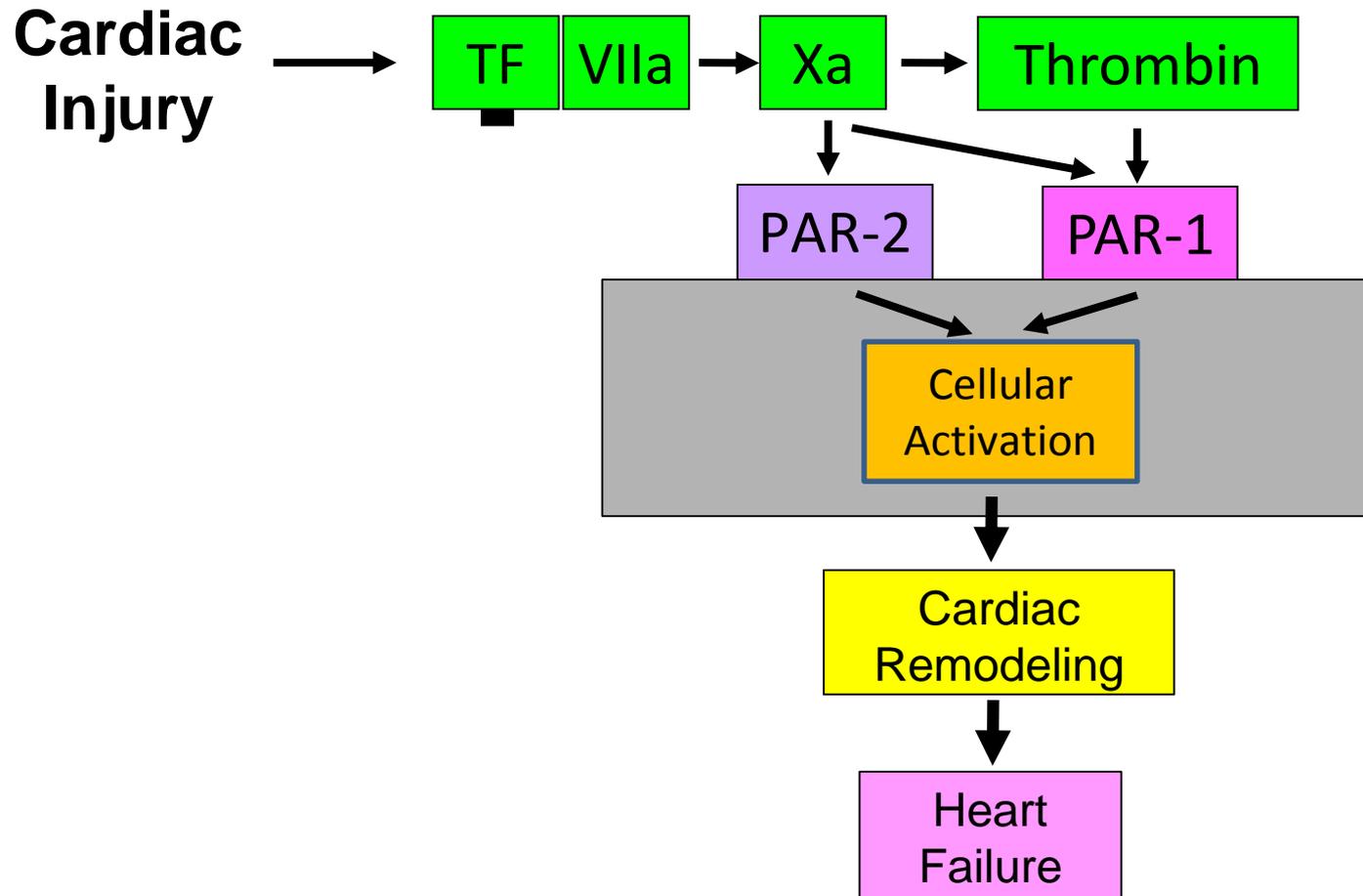
**Anticoagulants may increase  
alveolar hemorrhage and  
morbidity in patients infected with  
influenza A**

# Mouse Models of Cardiac Injury

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- Ischemia-reperfusion (I/R) injury
- LAD ligation
- Doxorubicin (anthracycline)
- Transaortic constriction – pressure overload
- Viral myocarditis (CVB3)

# Role of the Clotting Cascade and PARs in Cardiac Remodeling and Heart Failure

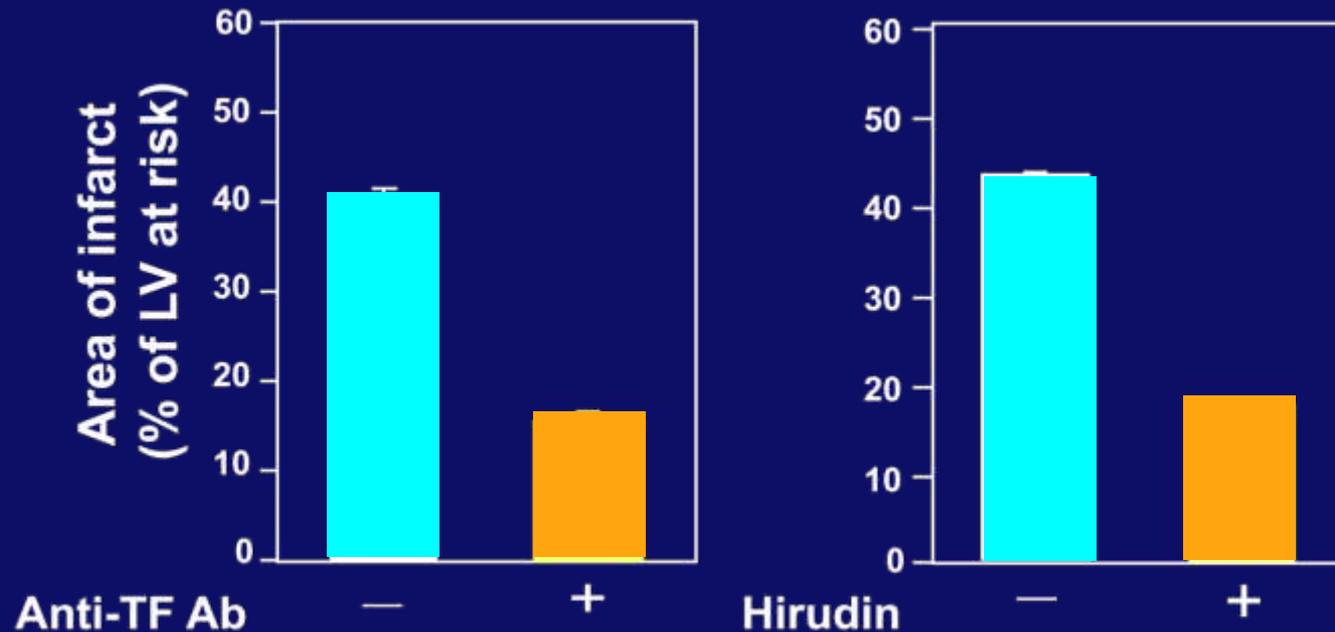


# Hypothesis

**Inhibition of TF and coagulation proteases or a deficiency in PAR-1 or PAR-2 will reduce infarct size and cardiac remodeling after cardiac injury in animal models**

# Effect of Inhibition of TF or Thrombin on Infarct Size in a Rabbit Cardiac I/R Model

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Inhibition of TF or thrombin also reduced inflammation

Erlich et al., *Am J Path* 2000

# Clinical Studies with Rivaroxaban

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**ATLAS ACS-2-TIMI-51 trial- the factor Xa inhibitor rivaroxaban reduced death due to cardiovascular causes (MI and stroke) in patients with a recent ACS event** Mega J et al NEJM 2012.

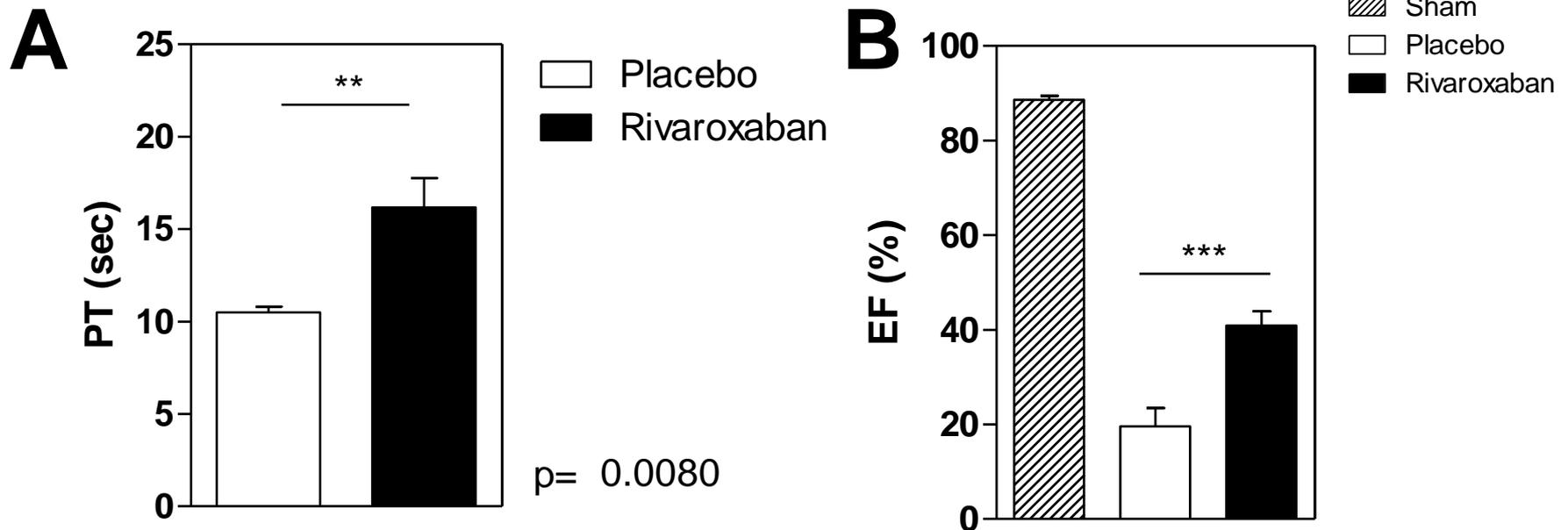
**In a subgroup analysis of HF patients, treatment with rivaroxaban 2.5 mg twice daily plus antiplatelet therapy demonstrated decreased rates of cardiovascular events and death compared with antiplatelet therapy alone.**

**COMMANDER-HF is a phase III clinical trial to evaluate the effect of the new oral anticoagulant rivaroxaban in patients with chronic heart failure.**

# Hypothesis

**Inhibition of FXa with rivaroxaban will reduce cardiac remodeling and heart failure in mice**

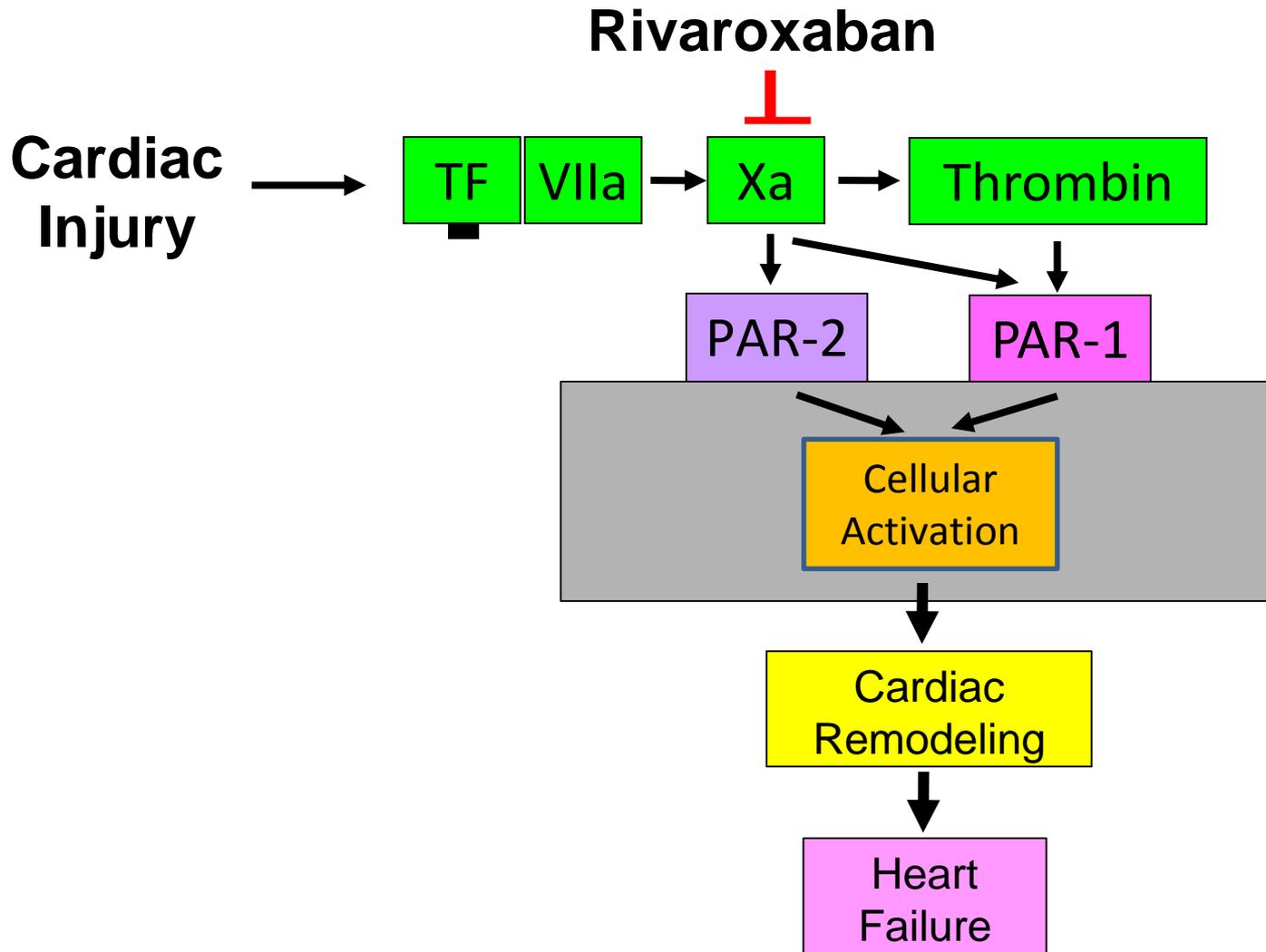
# Effect of Rivaroxaban on Cardiac Remodeling and Heart Failure in a Mouse Model of LAD Ligation



28 days after LAD ligation

Bode M unpublished results

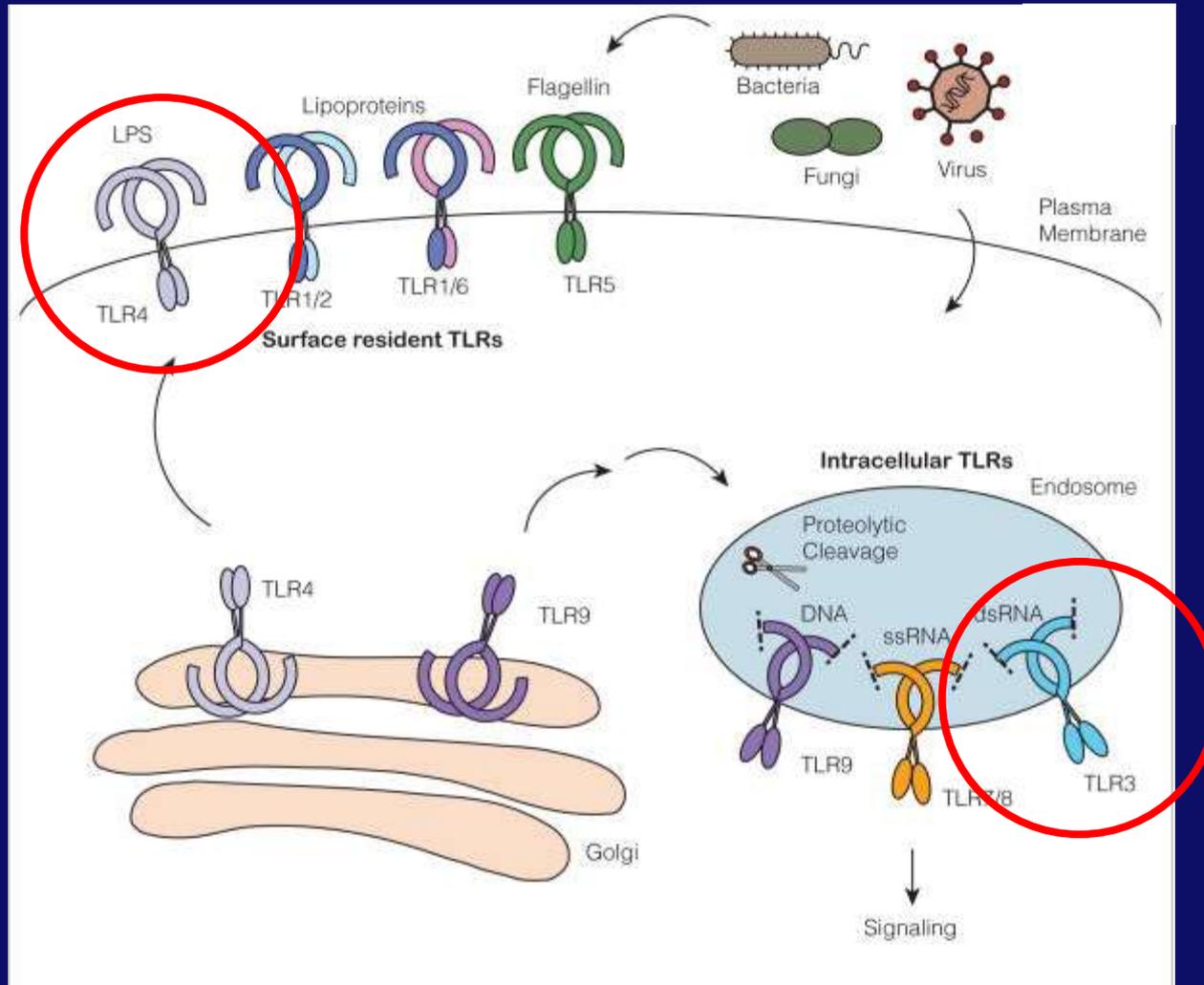
# Effect of the FXa Inhibitor Rivaroxaban on Cardiac Remodeling and Heart Failure





# **TLRs and PARs in host defense**

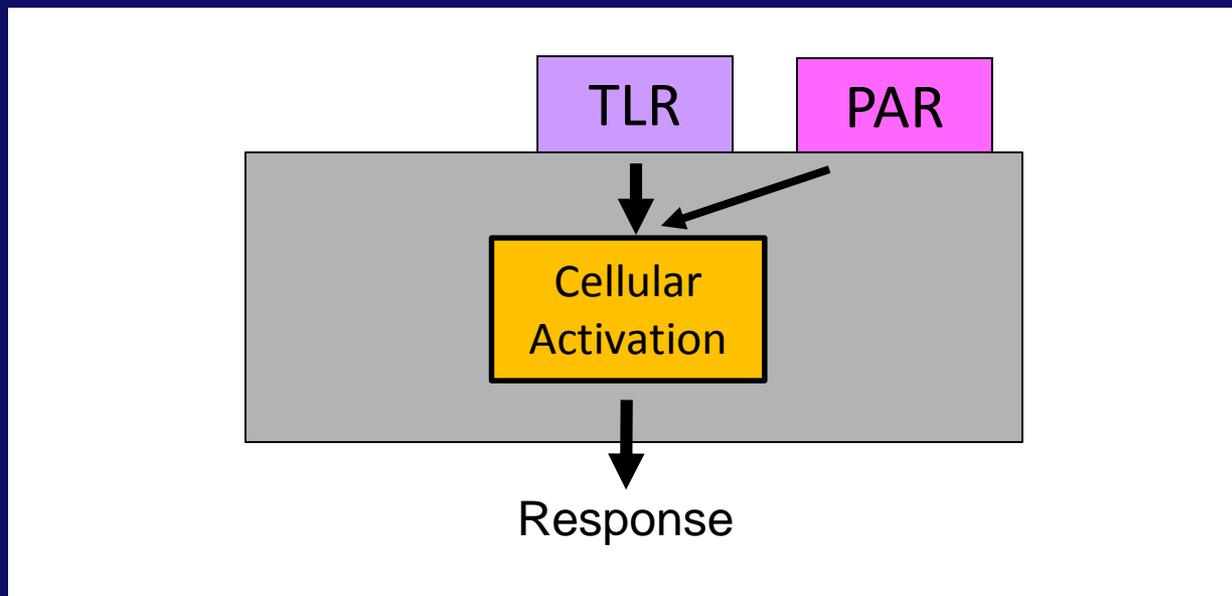
# Detection of Microbial Pathogens by the TLR Receptor Family



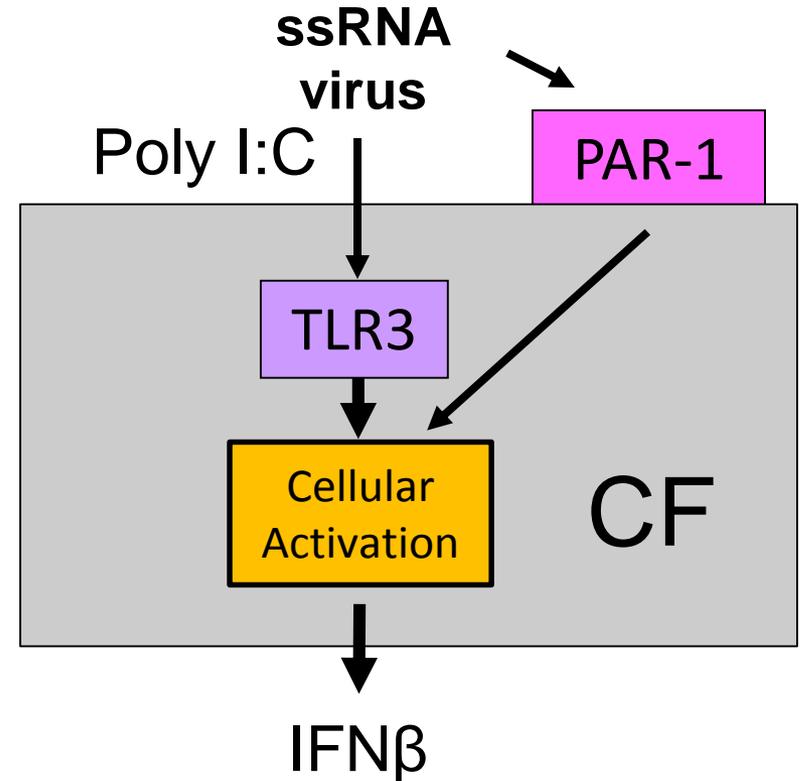
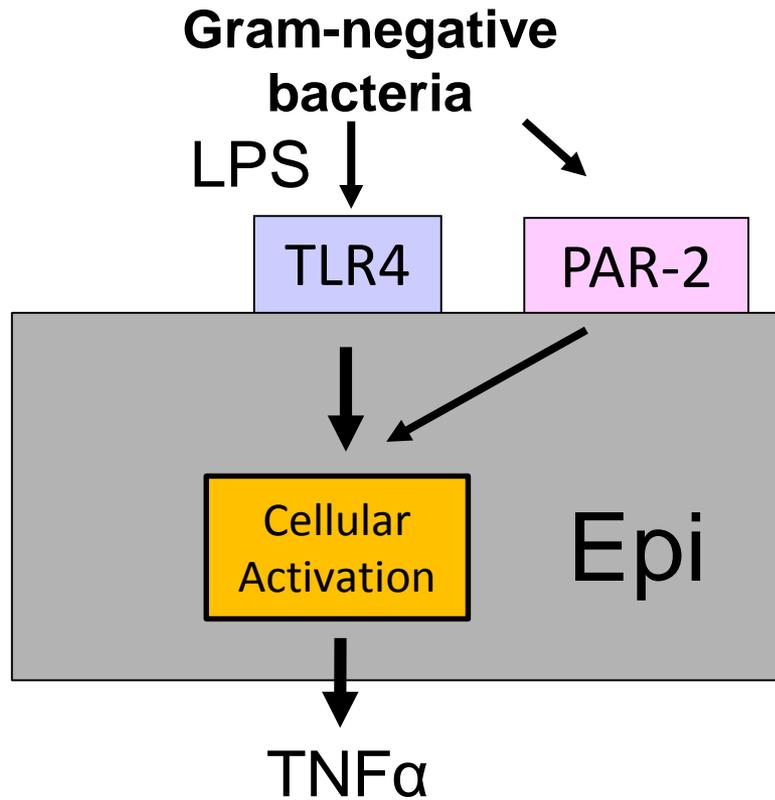
# TLRs and PARs as Co-receptors in the Detection of Pathogens

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- It has been proposed that the TLR family and the PAR family of receptors act as a dual-sensor system to detect pathogens. (Moretti S et al Mucosal Immunol 2008).



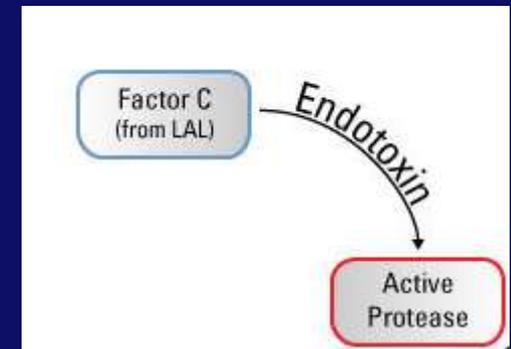
# TLRs and PARs in the Detection of Bacterial and Viral Pathogens



**Why is the clotting  
system activated during  
microbial infection?**

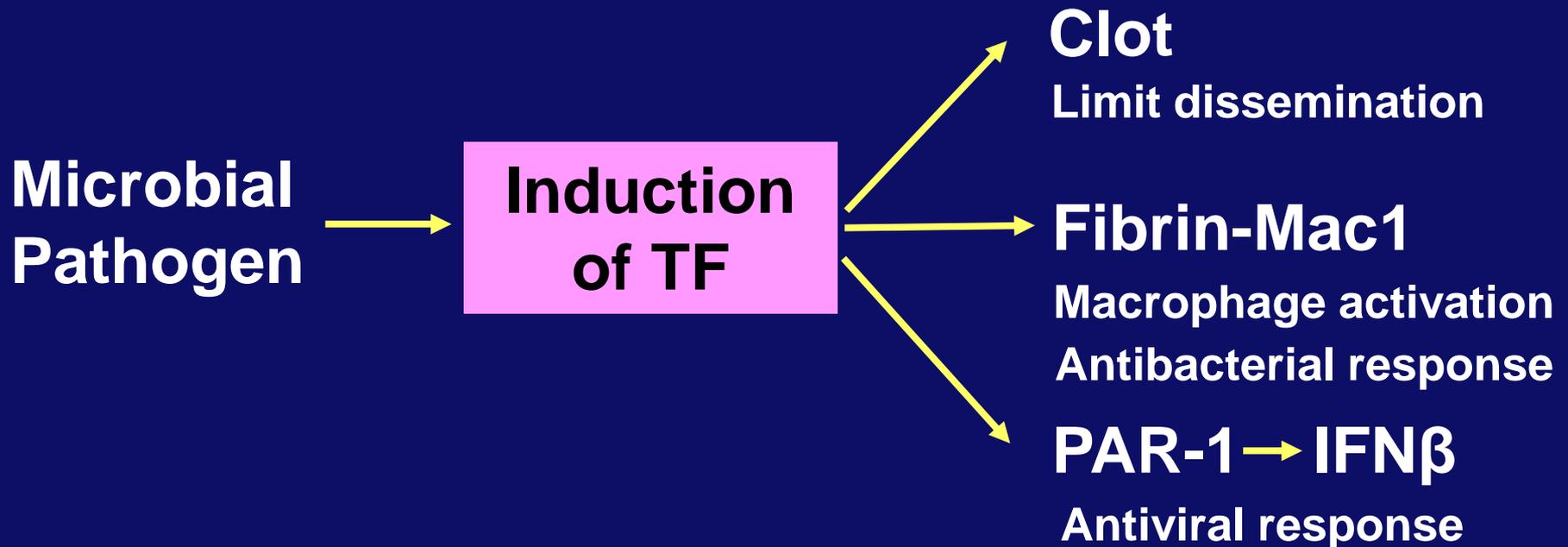
# Hemolymph Coagulation

Amebocytes in the horseshoe crab (*Limulus polyphemus*) detect bacterial infection and trigger clotting of the hemolymph. Basis of the Limulus Amebocyte Assay (LAL) for detecting bacterial LPS.



# The Clotting Cascade and Host Defense

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# Parallels between Gram-Negative Bacterial Sepsis and Ebola Hemorrhagic Fever

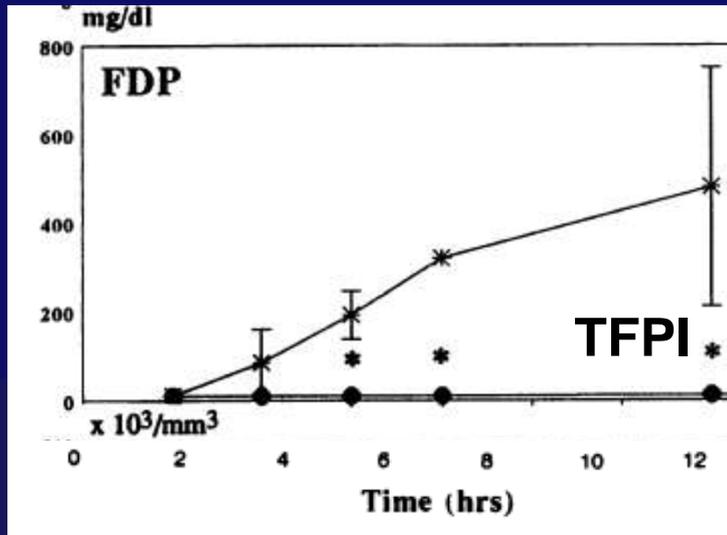
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- Localized coagulation is part of the host response to a microbial infection and is protective.
- However, systemic activation of coagulation during sepsis leads to disseminated intravascular coagulation (DIC) that consumes clotting factors (consumptive coagulopathy) and can lead to secondary bleeding.



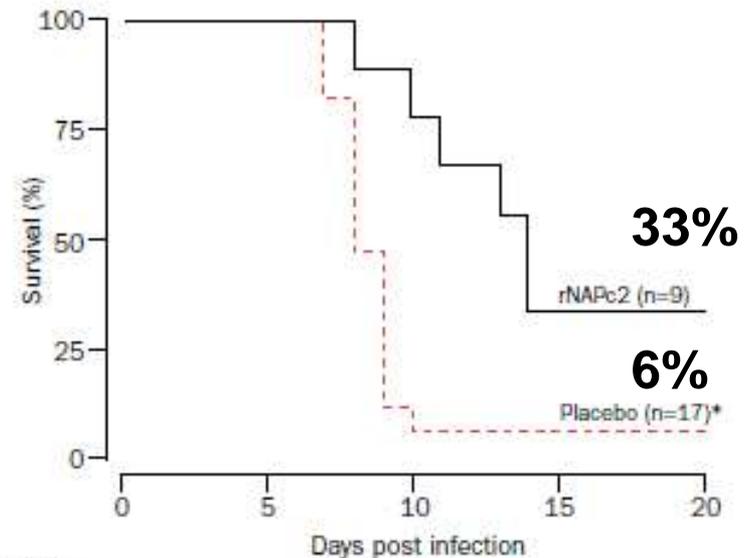
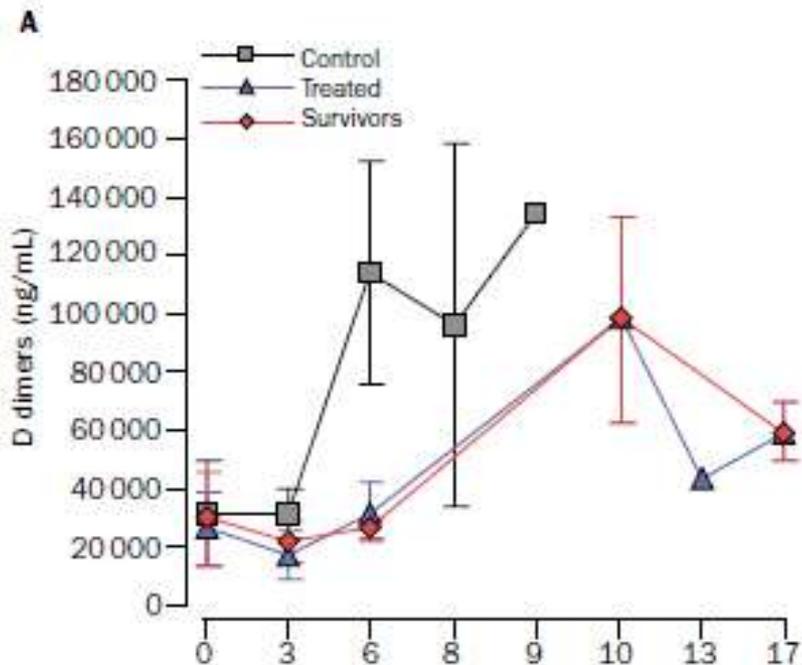
Petechial rash in a monkey infection with Ebola virus (Giesbert et al J Infect Dis 2007)

# Role of Tissue Factor in a Lethal Baboon Model of *E.coli* Sepsis



- Controls (n=5) mean survival was 39.9 h.
- Animals (n=5) treated with TFPI inhibitor all survived.

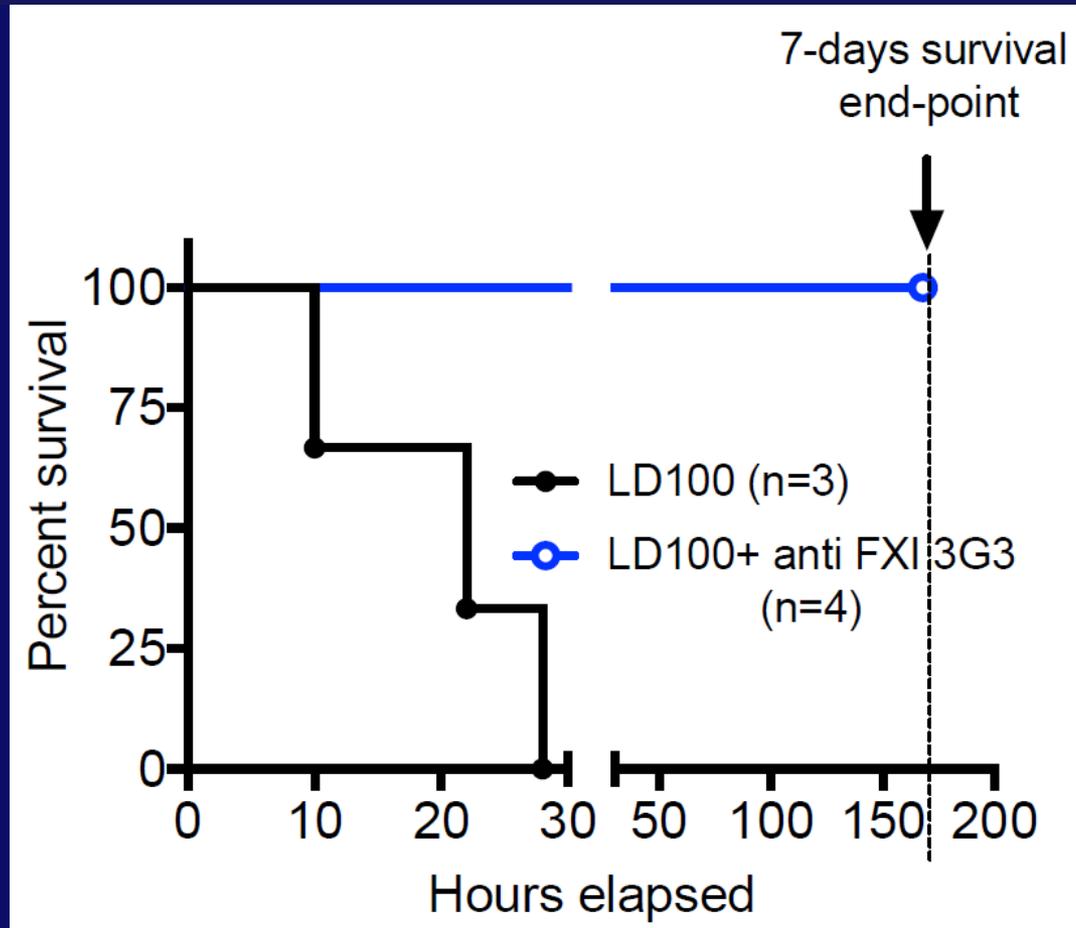
# Role of Tissue Factor in a Monkey Model of Ebola Virus Infection



Monkeys treated with NAPc2

Geisbert T et al Lancet 2003

# Effect of Inhibition of the Formation of FXIa on Survival in a Baboon Model of *S.aureus* Sepsis



McCarty O et al unpublished data

# **Cellular Sources of Tissues Factor that Activate Coagulation in Bacterial and Viral Infections**

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**TF induced on monocyte**

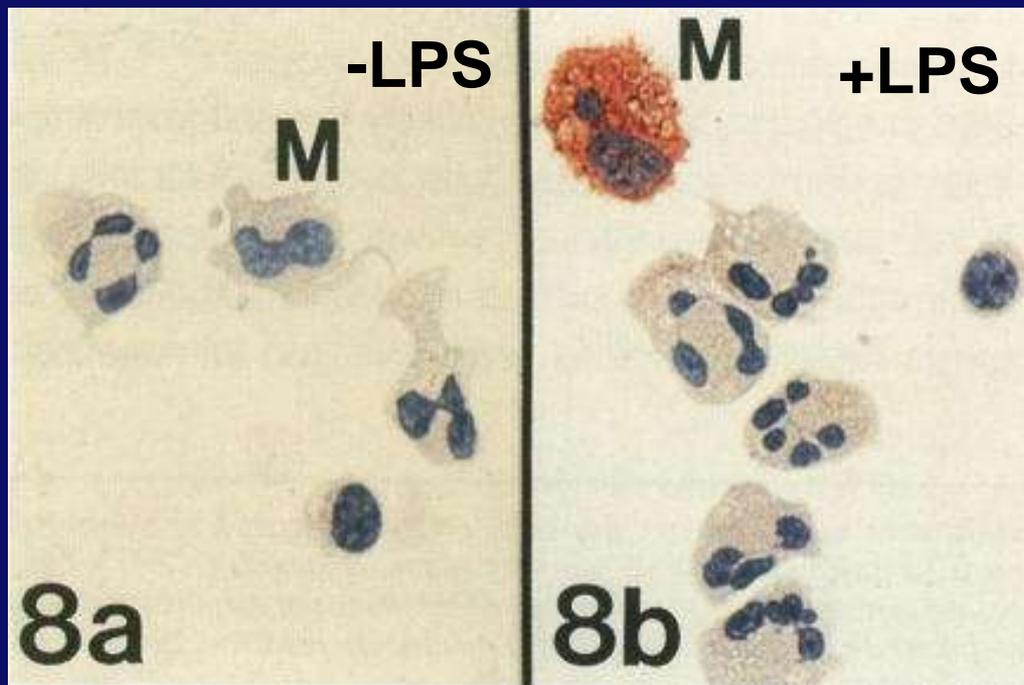
**TF induced on endothelial cells**

**LPS - used to model Gram-  
negative sepsis (TLR4)**

**Poly I:C - used to model infection  
with a ssRNA virus (TLR3)**

# LPS Induction of TF Expression in Monocytes

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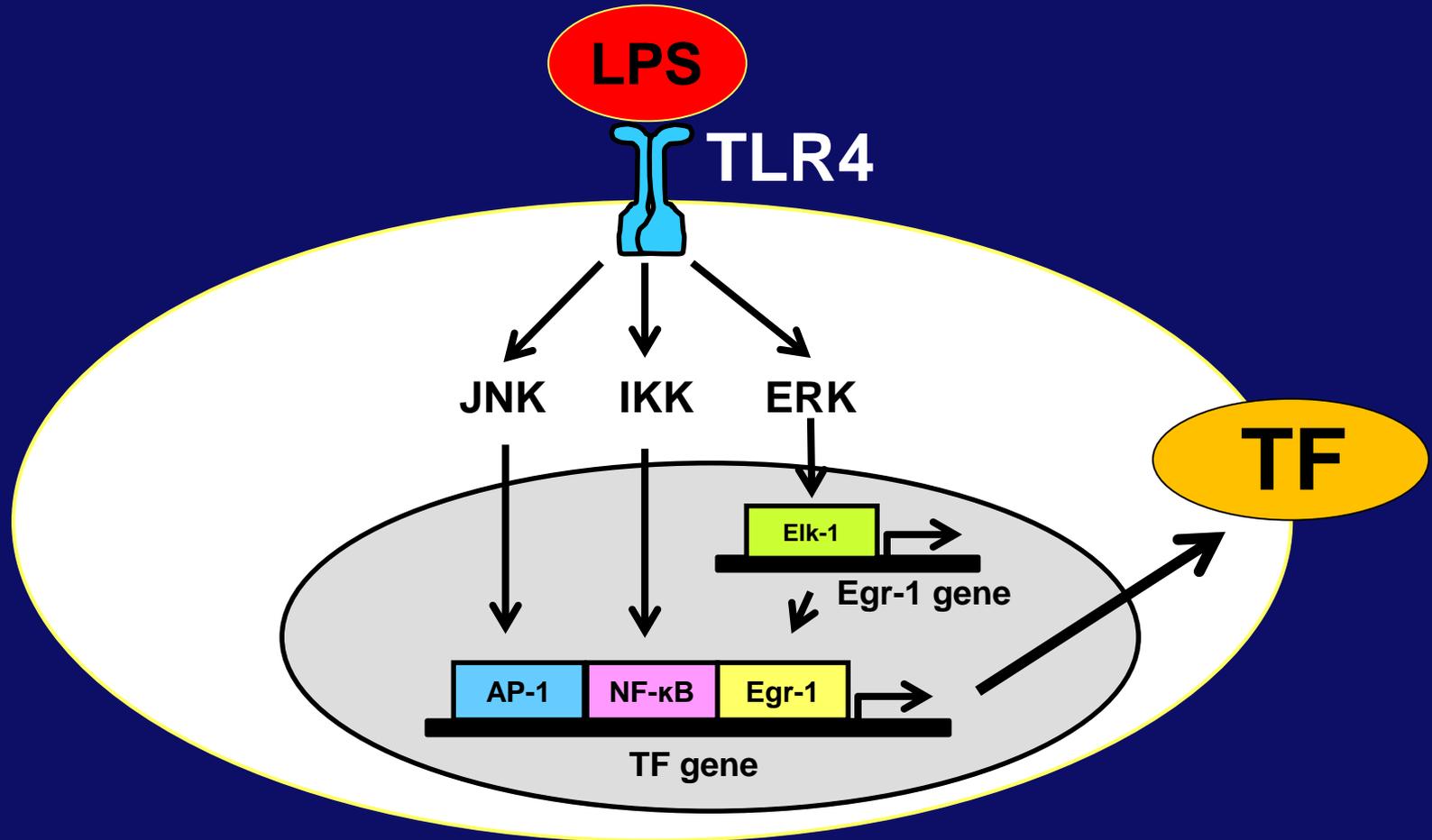


Host Defense

6 hrs

Drake et al AJP 1989

# LPS Induction of the TF Gene in Monocytic Cells



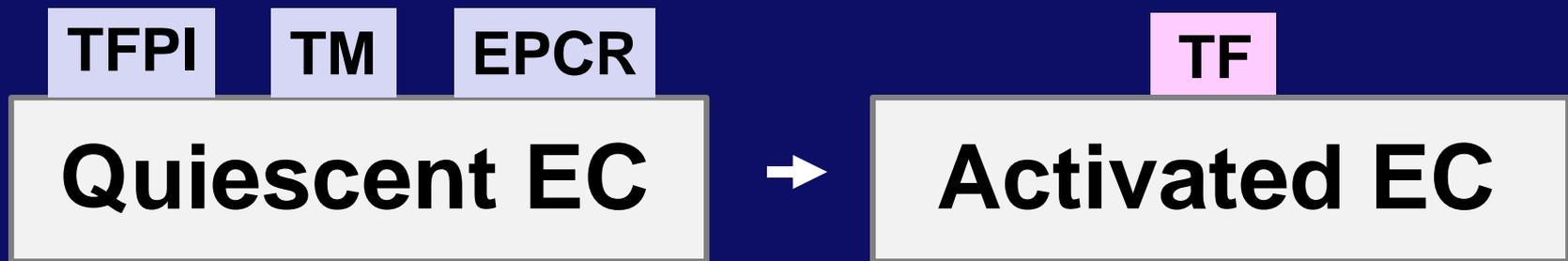
Mackman et al, JEM 1991; Mackman JBC 1994; Parry and Mackman JBC 1994; Oeth et al, MCB 1994; Guha et al, Blood 2002

# Expression of Anticoagulant and Procoagulant Proteins by ECs

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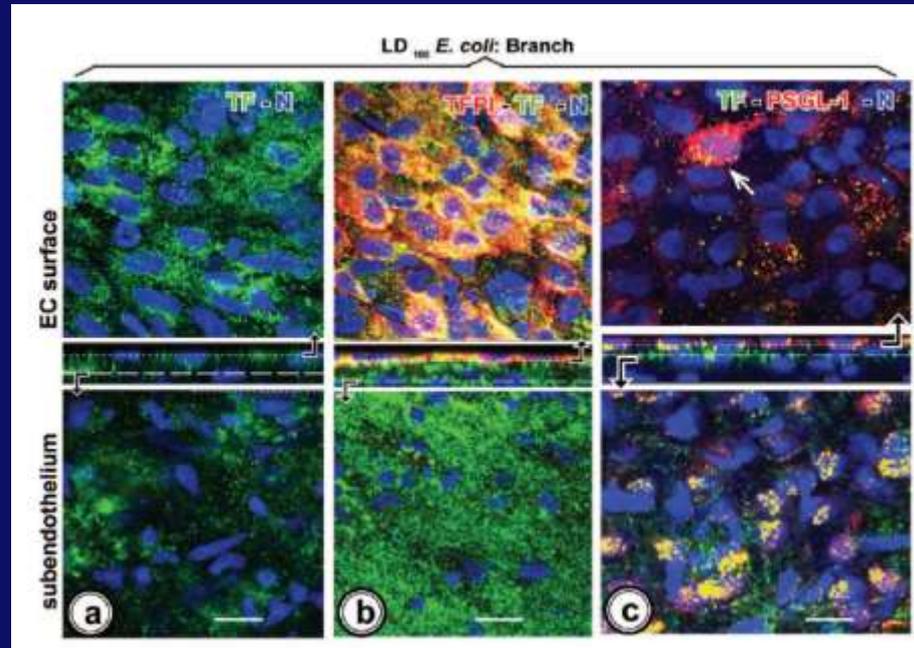
**Anticoagulant**

**Procoagulant**



LPS, inflammatory mediators and various viruses induce TF expression in cultured Ecs. However, there is limited evidence that TF is expression by the endothelium in vivo.

# TF Expression in the Endothelium of Arterial Branching Areas in Baboons with *E.coli* Sepsis



TF expression in the endothelium is difficult to detect in septic baboons, and may be due to binding of monocyte-derived TF<sup>+</sup> MVs

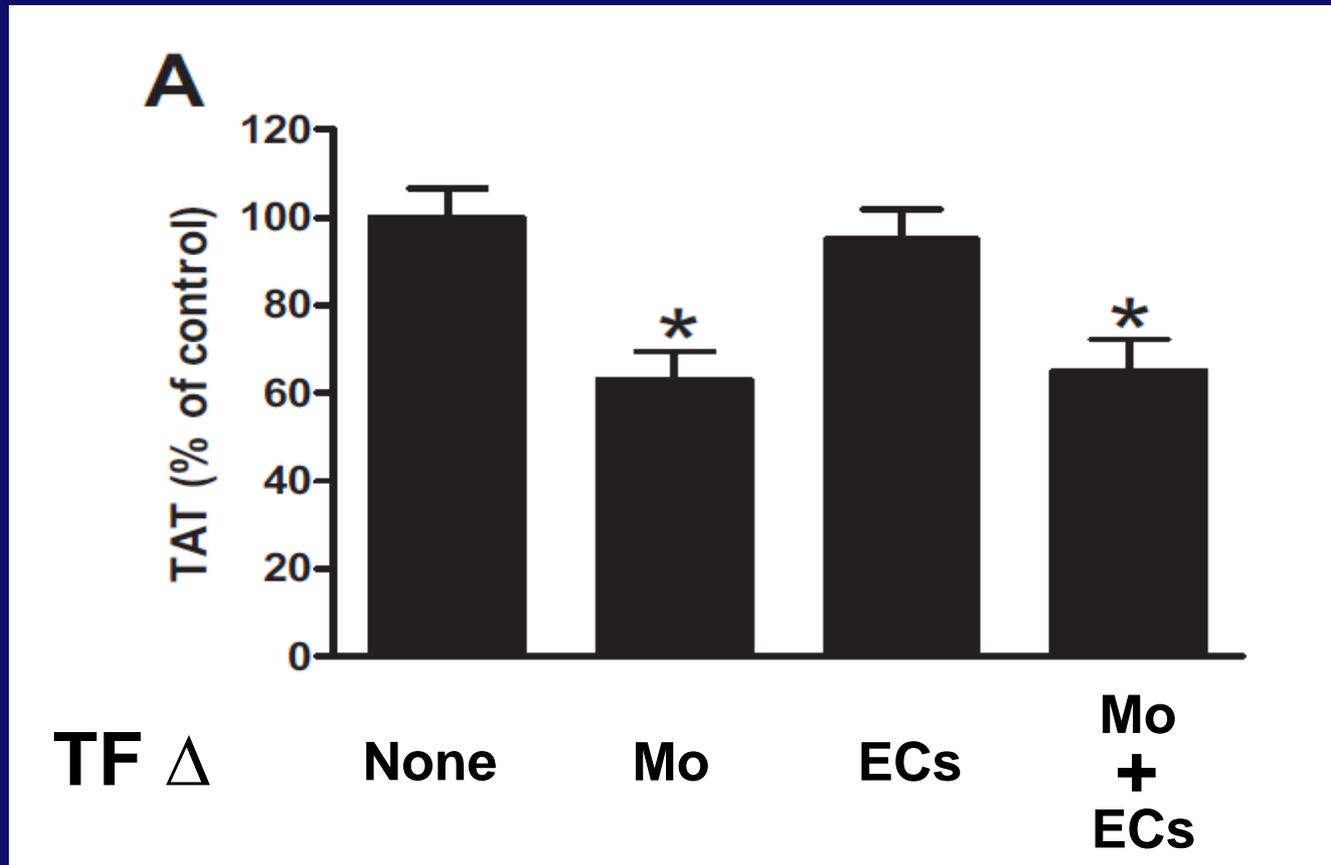
Lupu C et al Am J Path 2005

# Role of Monocyte and Endothelial Cell TF in the Activation of Coagulation in a Mouse Model of Endotoxemia

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- $TF^{fl/fl}$ , LysMcre mice (deletion of TF in myeloid cell), decrease in TAT.
- $TF^{fl/fl}$ , Tie2Cre mice (deletion of TF in ECs and hematopoietic cells), decrease in TAT.
- Transplant  $TF^{fl/fl}$ , Tie2Cre mice with WT BM to look at contribution of EC TF.

# Role of TF Expression by Monocytes and EC in the Activation of Coagulation in a Mouse Endotoxemia Model



Model – LPS (5 mg/kg) IP for 8 h

Pawlinski R et al Blood 2010

# Conclusion

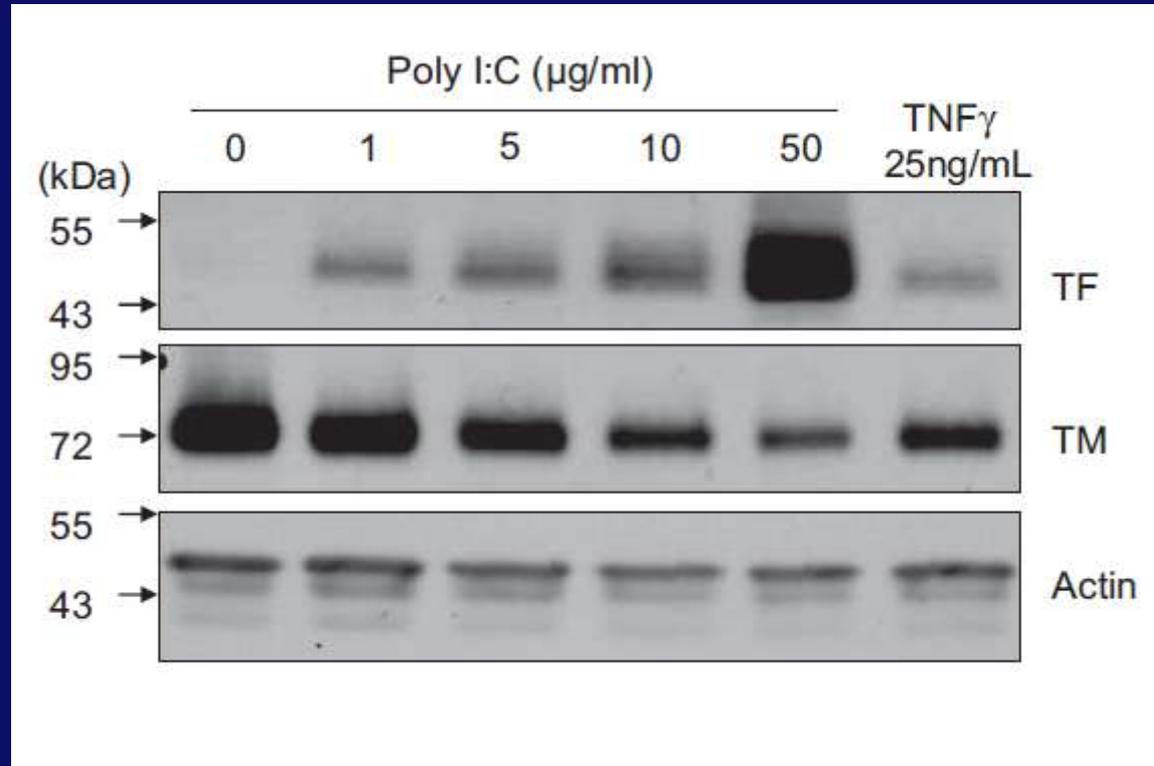
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**Monocytes are a major source of inducible TF expression that activates the coagulation cascade in endotoxemia**

# Poly I:C Induction of TF Expression in HUVECs



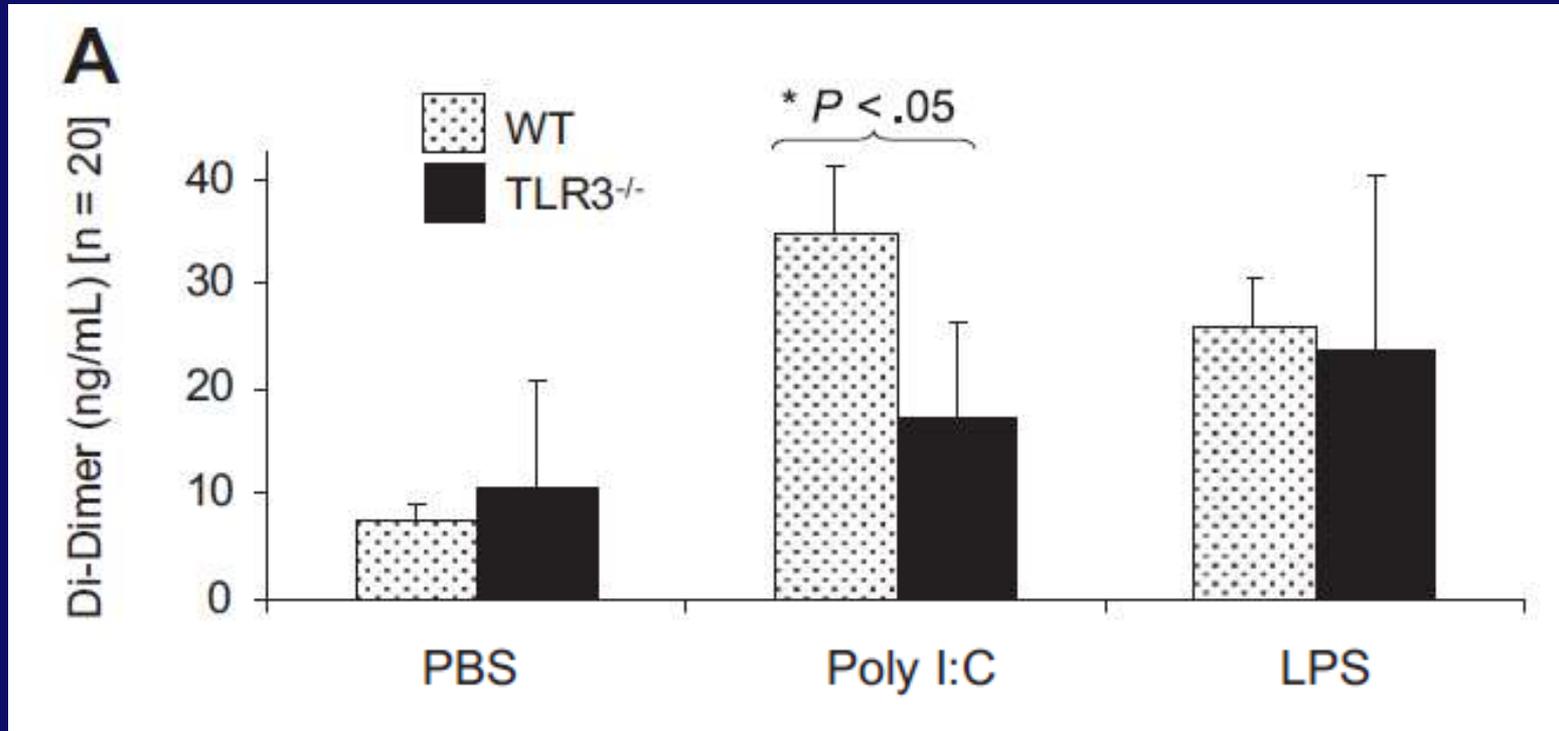
Saravanan  
Subramanian



**Poly I:C does not induce TF expression  
in monocytes**

Kanse S et al Blood 2009

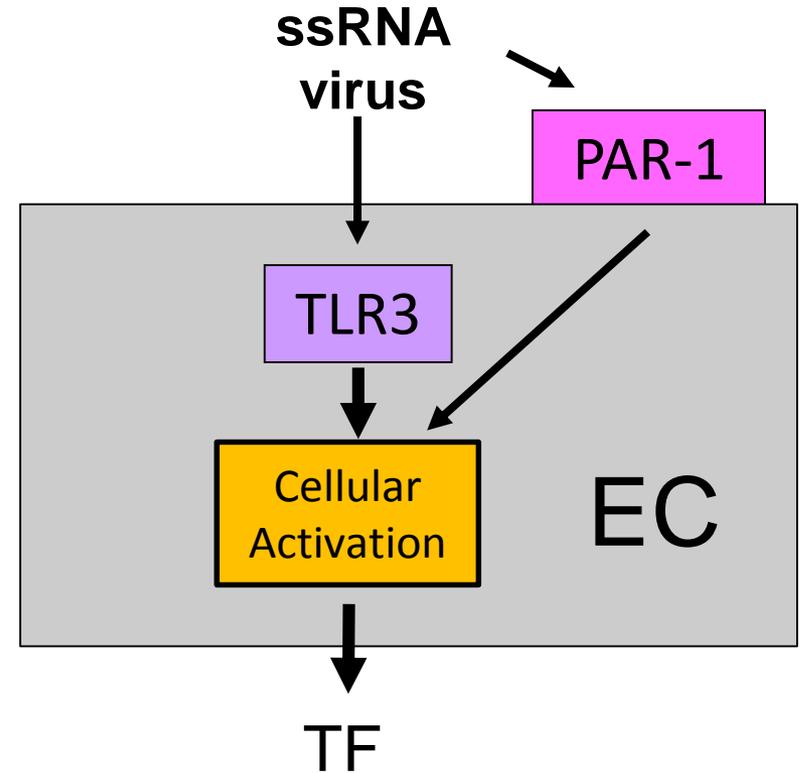
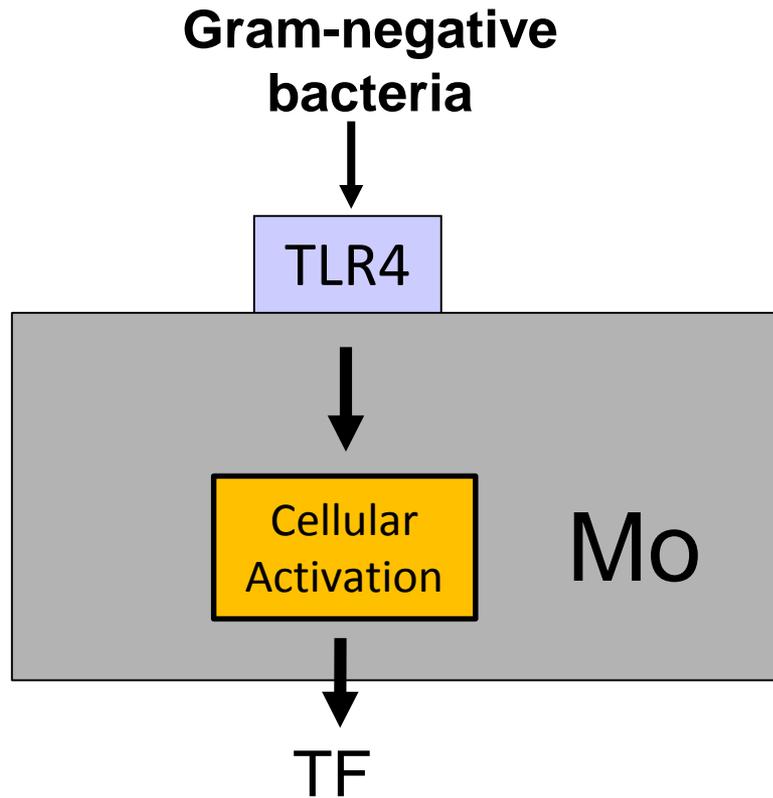
# Role of TLR3 in Poly I:C Activation of Coagulation in Mice



We are currently determining the role of TF expression by monocytes and ECs in poly I:C activation of coagulation in mice

Kanse S et al Blood 2009

# TF Expression by Monocytes and ECs during Bacterial and Viral Infections



# **Role of the TF-thrombin-PAR-1 pathway in the antiviral response**

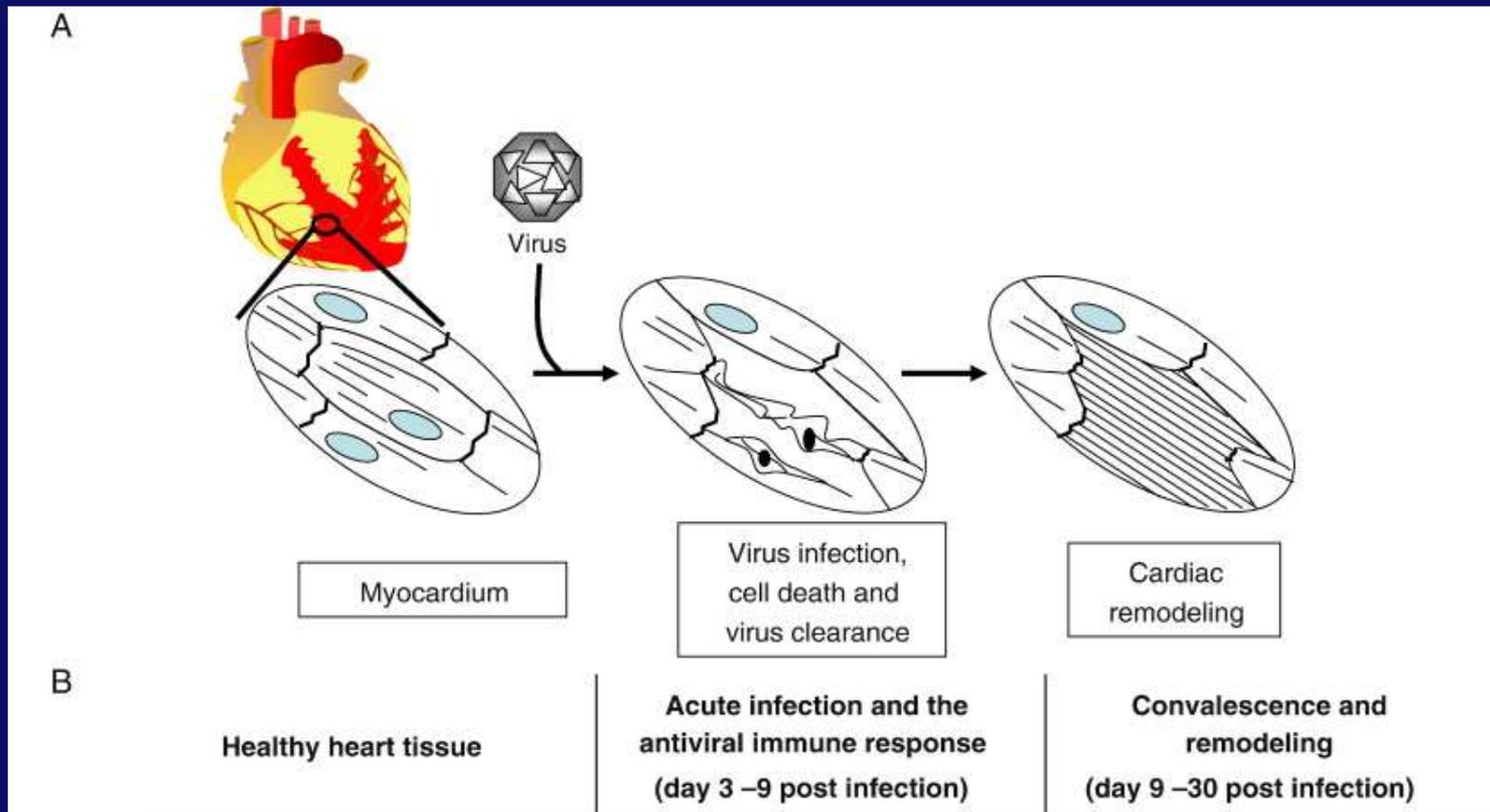
# Outline

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- **Coxsackievirus B3 virus model**
- **Influenza A virus model**

# Viral Myocarditis

Accounts for up to 20% of sudden cardiac death among young people (<40 years of age)

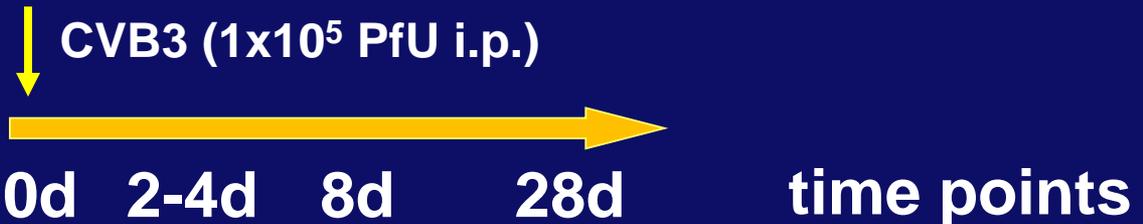


# Mouse Model of CVB3-induced Myocarditis

- Coxsackievirus B3 (CVB3) = cardiotropic ssRNA(+) virus



**Silvio Antoniak**



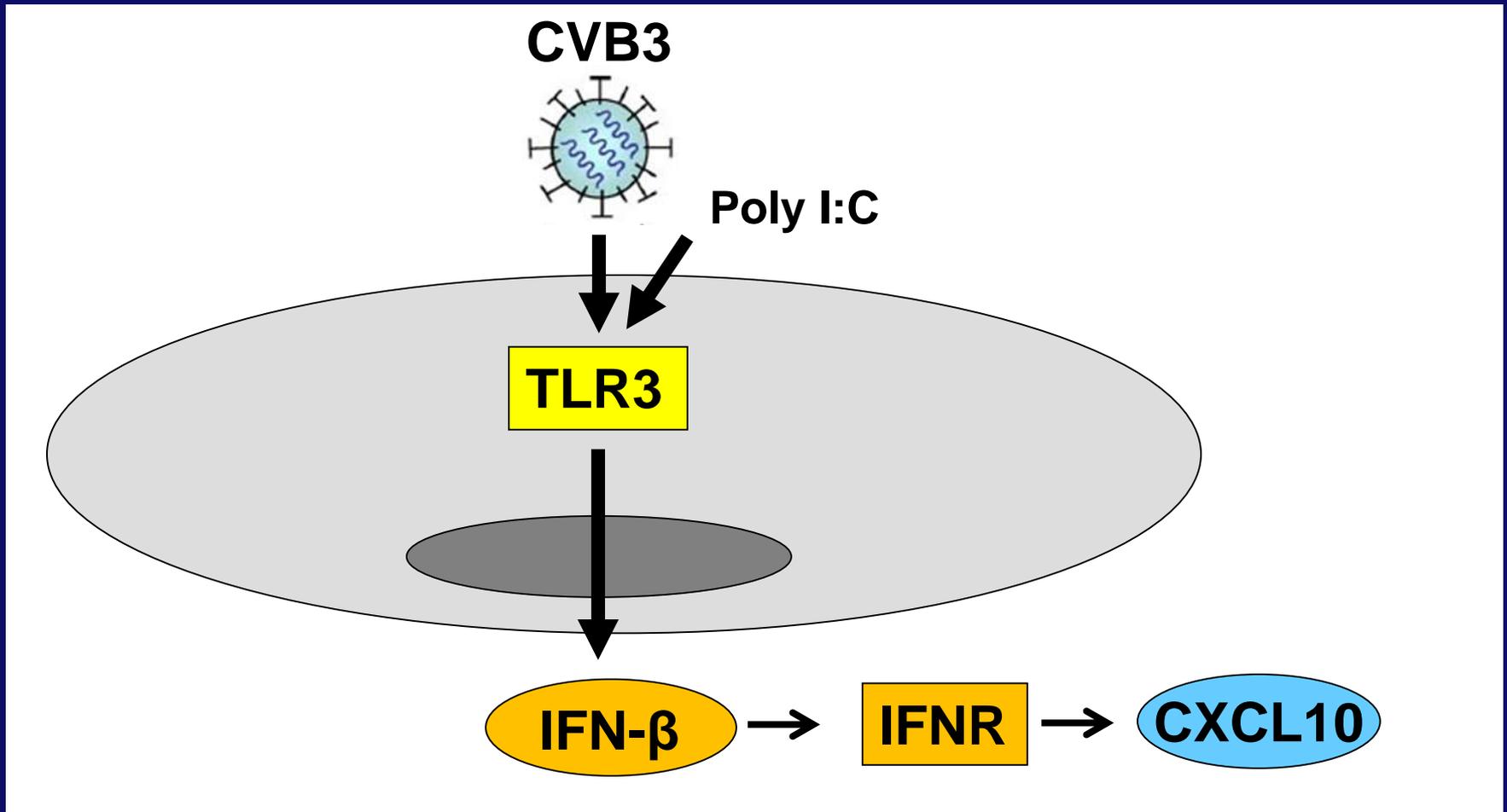
Immune response  
IFN $\beta$  and CXCL10

Cardiac injury and  
viral replication

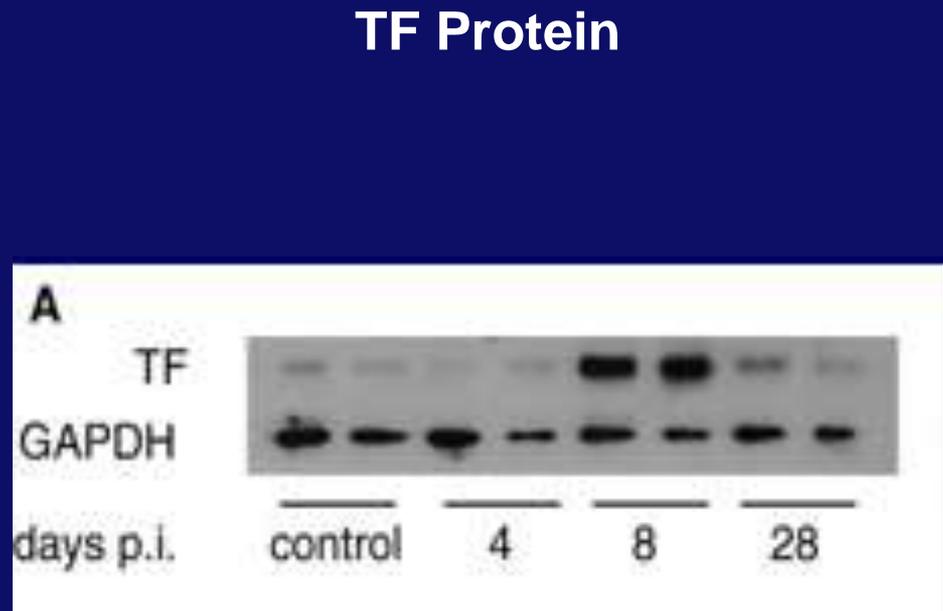
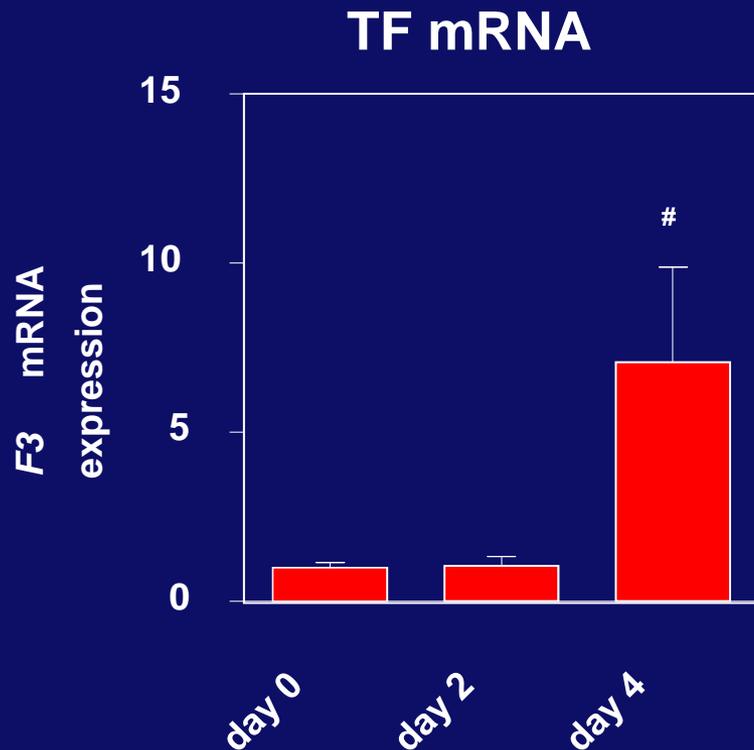
Heart failure

Silvio Antoniak

# Innate Immune Response to CVB3 Infection



# CVB3 Infection of Mice Increases TF Expression in the Heart



Antoniak S et al JMCC 2008; Antoniak S et al JCI 2013

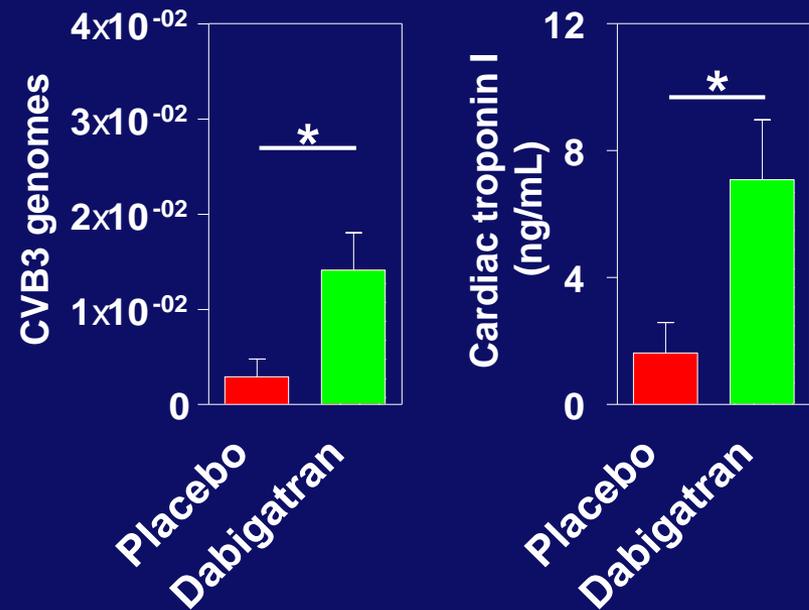
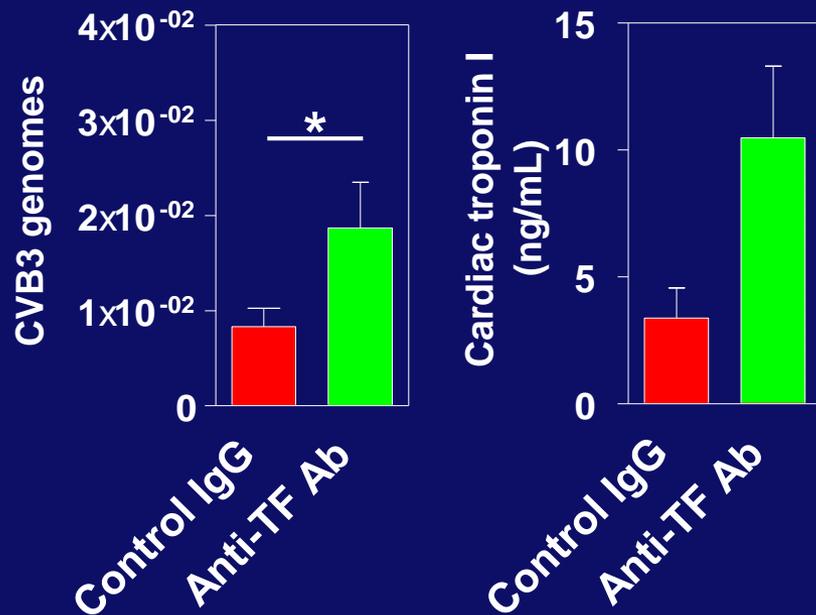
# Effect of Inhibition of TF or Thrombin on CVB3-induced Myocarditis

## Anti-TF Ab

## Dabigatran

day 8

day 8

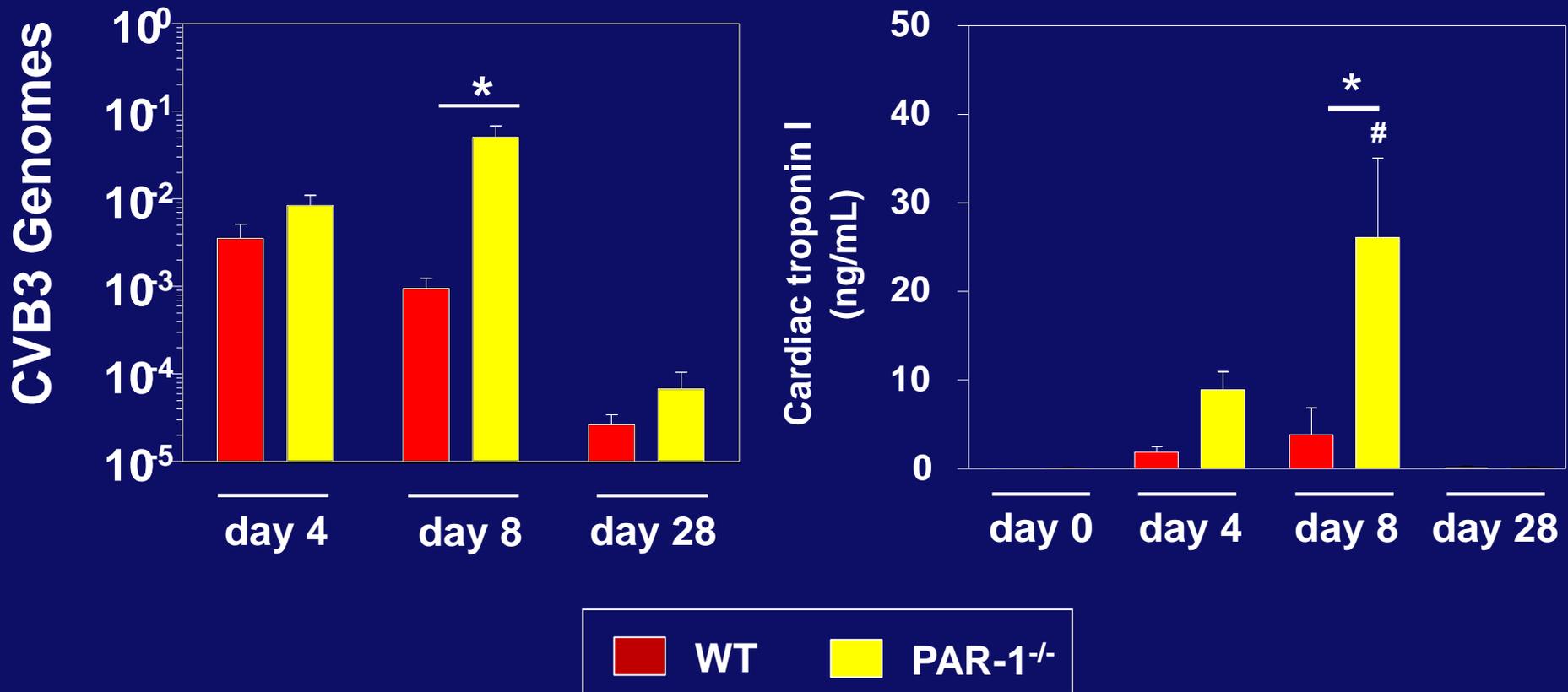


\* $P < 0.05$

Student's *t*-test

Antoniak et al, JCI 2013

# PAR-1 Deficiency is Associated with Increased Viral Load and Cardiac Injury after CVB3 infection

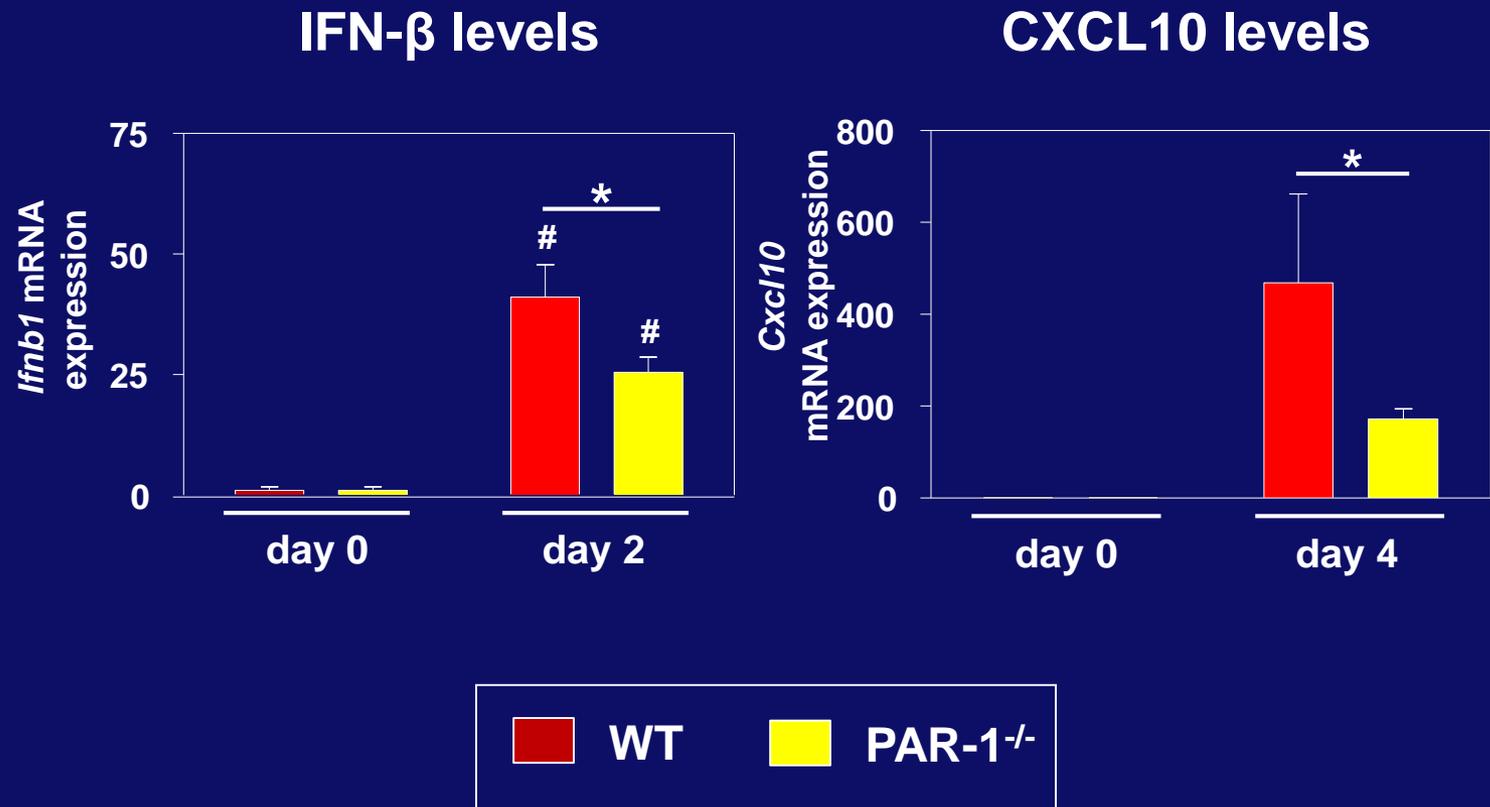


\*P<0.05

Two-Way ANOVA

Antoniak et al, JCI 2013

# PAR-1 Deficiency is Associated with Reduced IFN- $\beta$ and CXCL10 expression After CVB3 Infection



\*P<0.05; #P<0.05 vs. day 0  
Two-Way ANOVA

Antoniak et al, JCI 2013

**BMT studies indicated that PAR-1  
on both hematopoietic and non-  
hematopoietic cells contributed  
to the protection from CVB3  
infection.**

# Establishing a Cell Model to Study the Role of PAR-1 in CVB3 Infection

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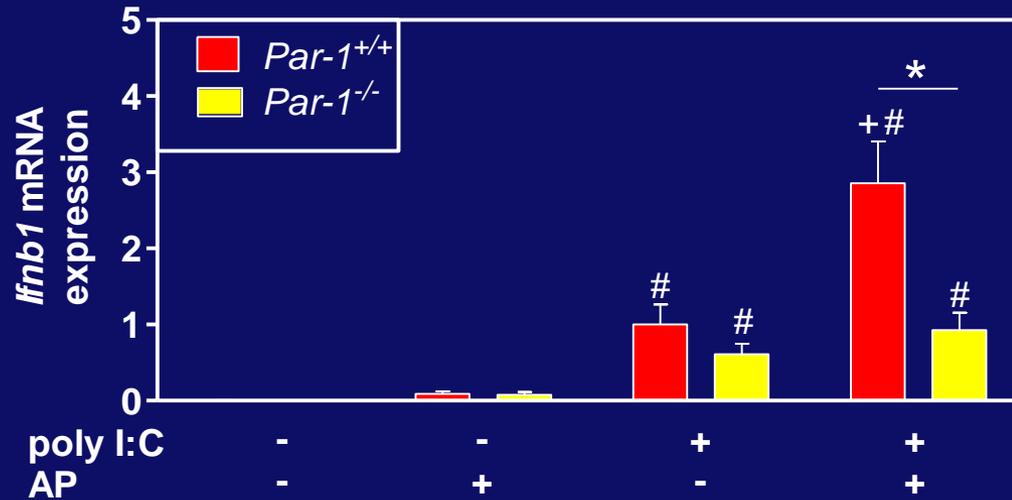
Poly I:C (dsRNA mimetic) was used to stimulate TLR3.

PAR-1 agonist peptide (AP) was used to stimulate PAR-1.

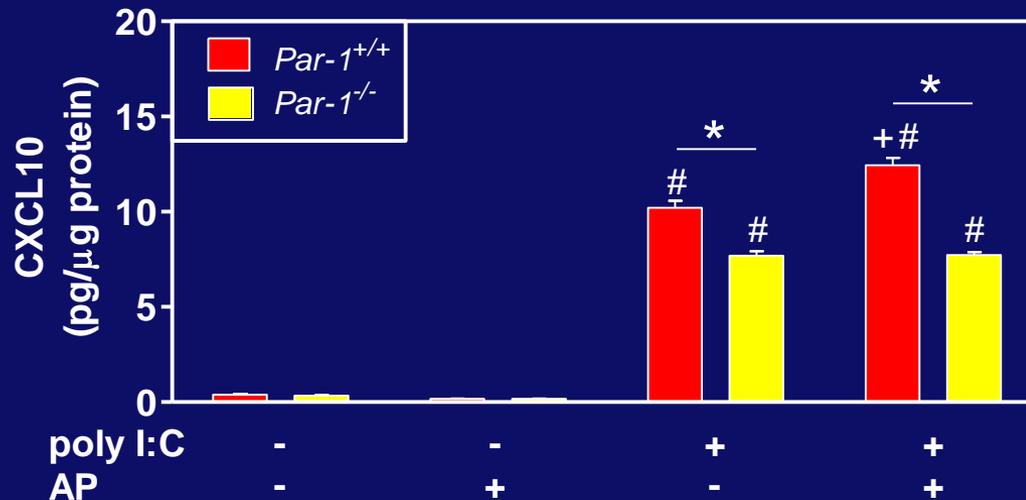
Use mouse cardiac fibroblasts.

# PAR-1 Activation Enhances Poly I:C Induction of IFN- $\beta$ and CXCL10 Expression in Cardiac Fibroblasts

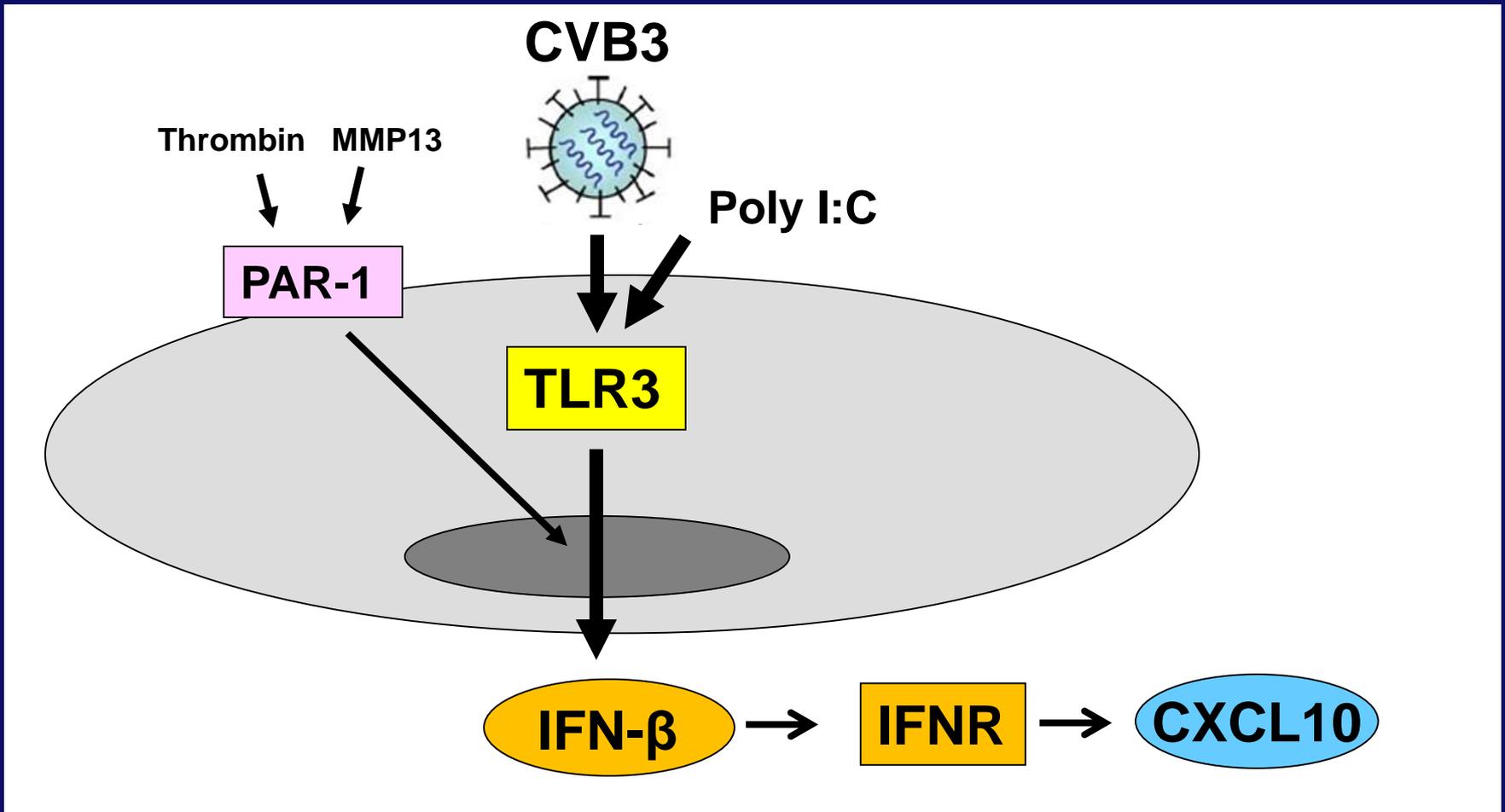
IFN $\beta$   
mRNA



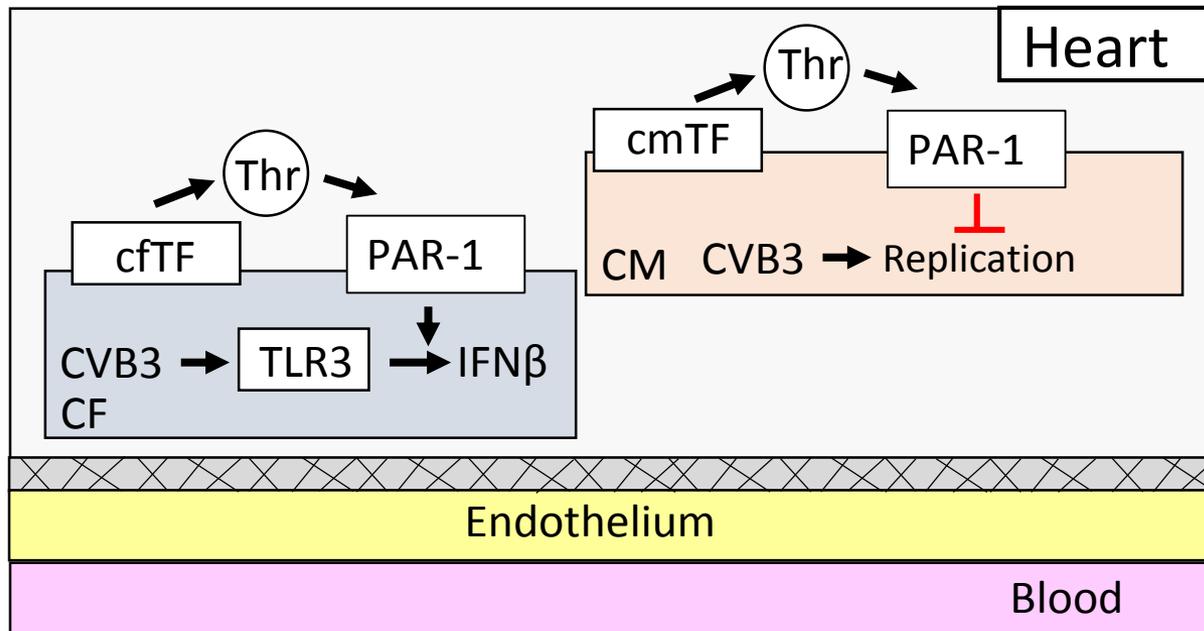
CXCL10  
Protein



# Innate Immune Response to CVB3 Infection



# Roles of the TF-Thrombin-PAR-1 Pathways in CVB3-induced Myocarditis



# Influenza A Infection

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- Influenza A (IAV) is a respiratory pathogen
- IAV causes acute infection of the upper respiratory tract.
- Severe IAV infection can cause infection of the lower respiratory tract, resulting in viral pneumonia.
- Damage to the barrier formed by epithelial and endothelial cells in the pulmonary alveolus leads to respiratory dysfunction.

# Mouse Model of Influenza A Infection (IAV)

---



**Kohei Tatsumi**

- Influenza A is a ssRNA virus.
- Influenza A/Puerto Rico/8/1934 H1N1.
- Mouse adapted and highly pathogenic.
- Infected intranasally.

**Silvio Antoniak and Kohei Tatsumi**

# Mouse Model of Influenza A Infection

---

H1N1 (0.02  
HAU, 50 $\mu$ L i.n.)



0d

3-7d

14d time points

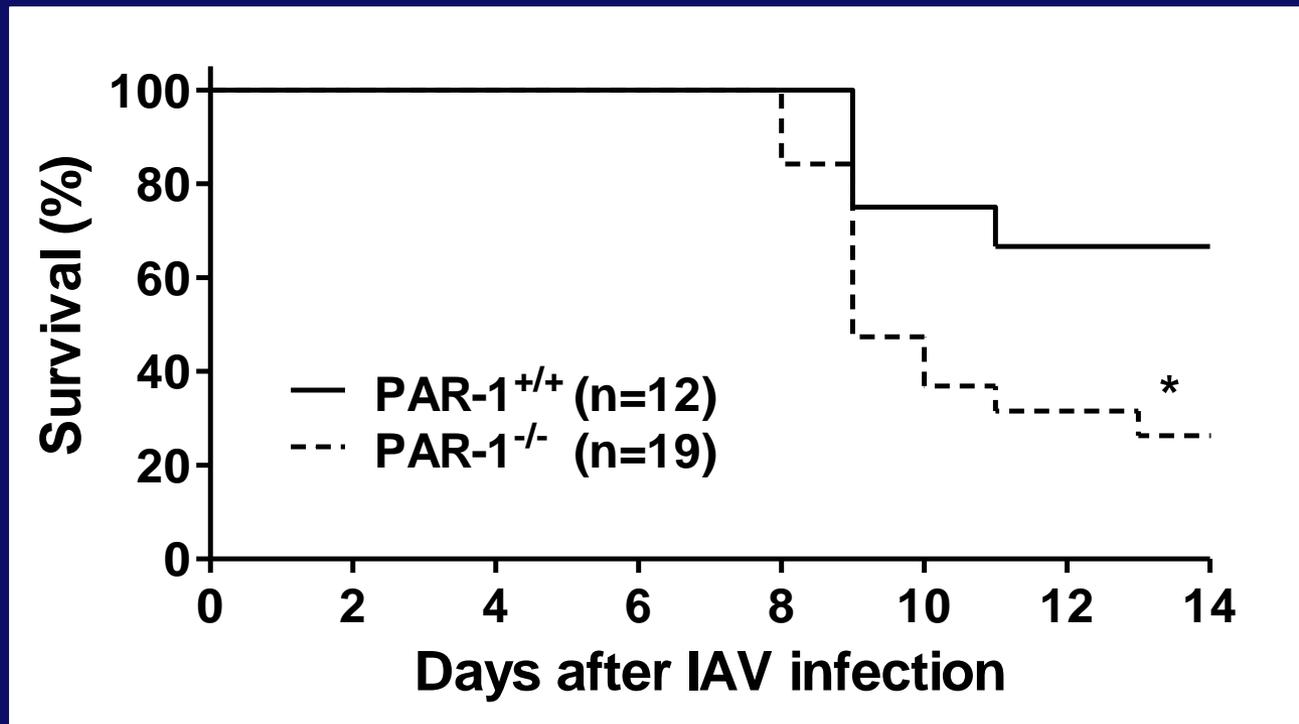
Inflammation,  
viral load and  
vascular  
permeability

Survival and  
weight loss

Model leads to ~20% mortality

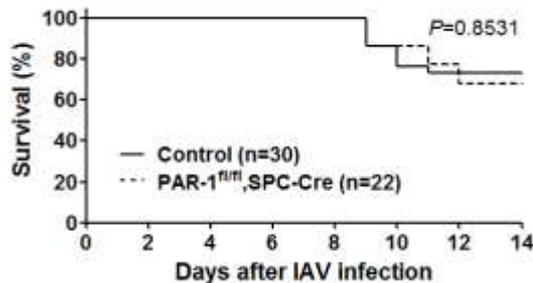
# PAR-1 Deficiency is Associated with Increased Mortality after Influenza A Infection

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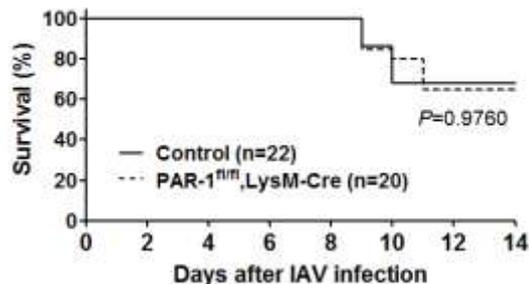


Tatsumi K et al. unpublished data

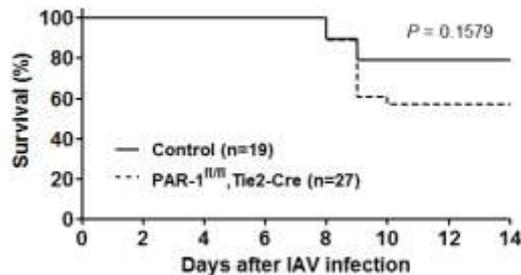
# Effect of Deletion of PAR-1 in Different Cell Types on IAV Infection



PAR-1 deletion in lung epithelial cells

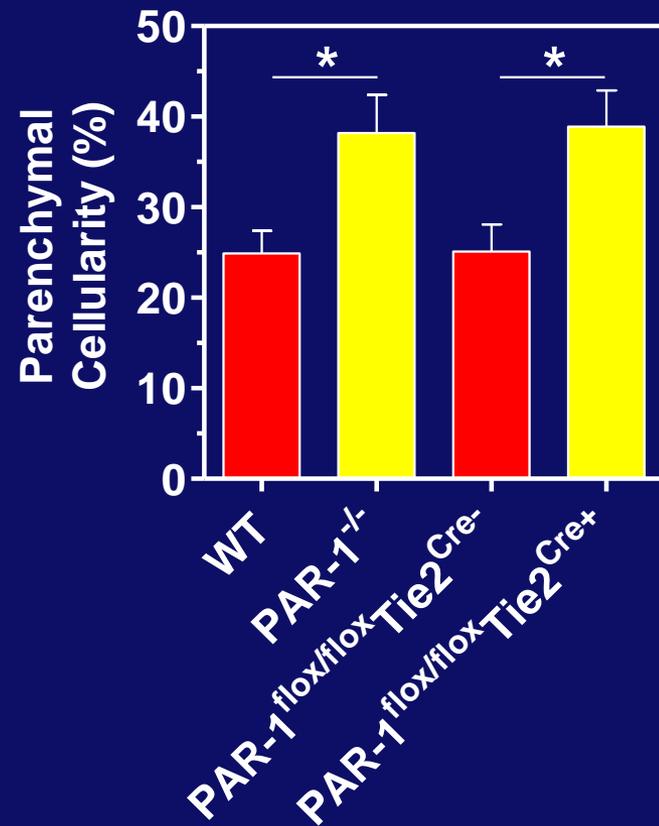
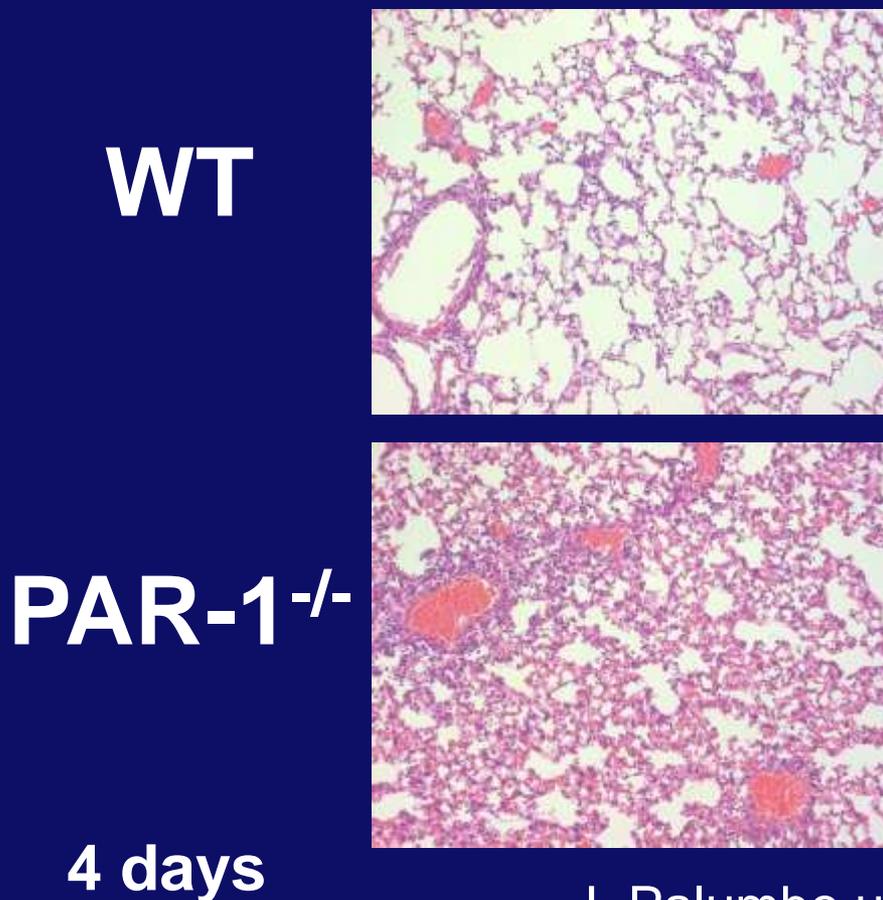


PAR-1 deletion in myeloid cells



PAR-1 deletion in ECs and hematopoietic cells

# PAR-1 Deficiency is Associated with Increased Levels of Parenchymal Cellularity after Influenza A Infection

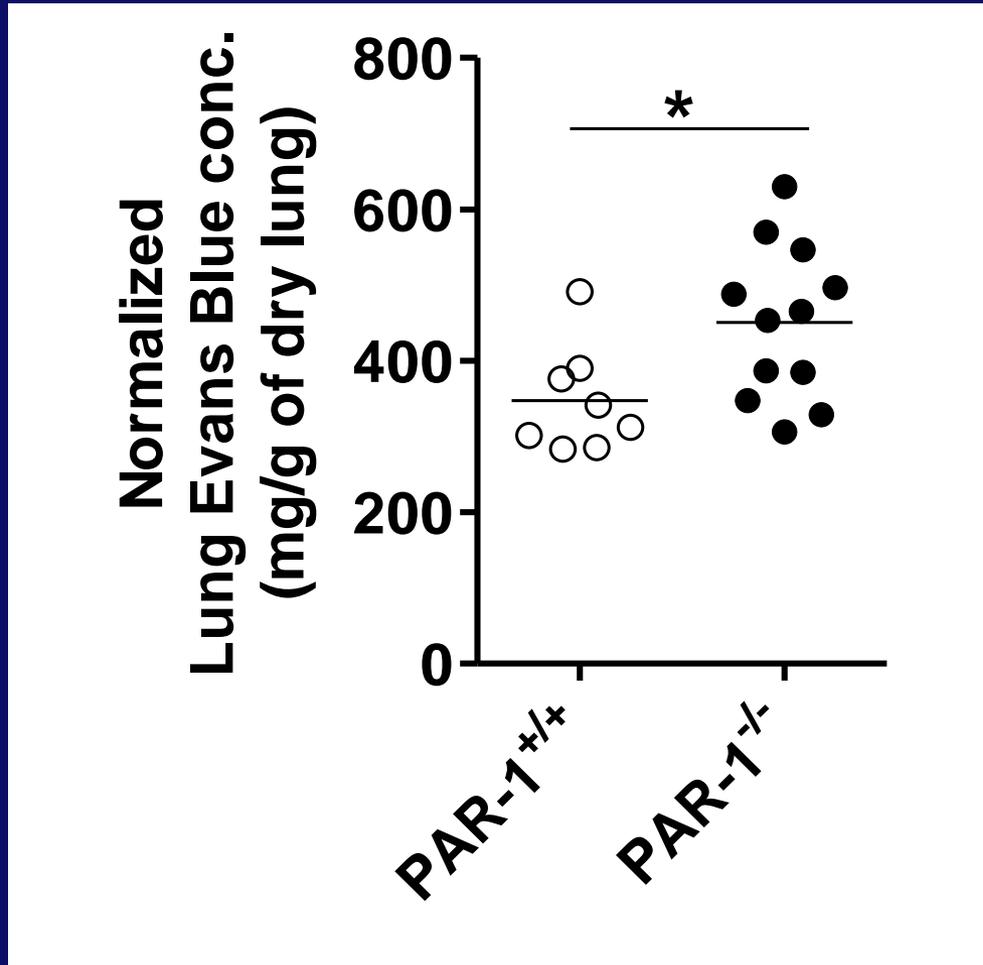


J. Palumbo unpublished data

# Hypothesis

**PAR-1 expression on the endothelium is barrier protective during influenza A infection.**

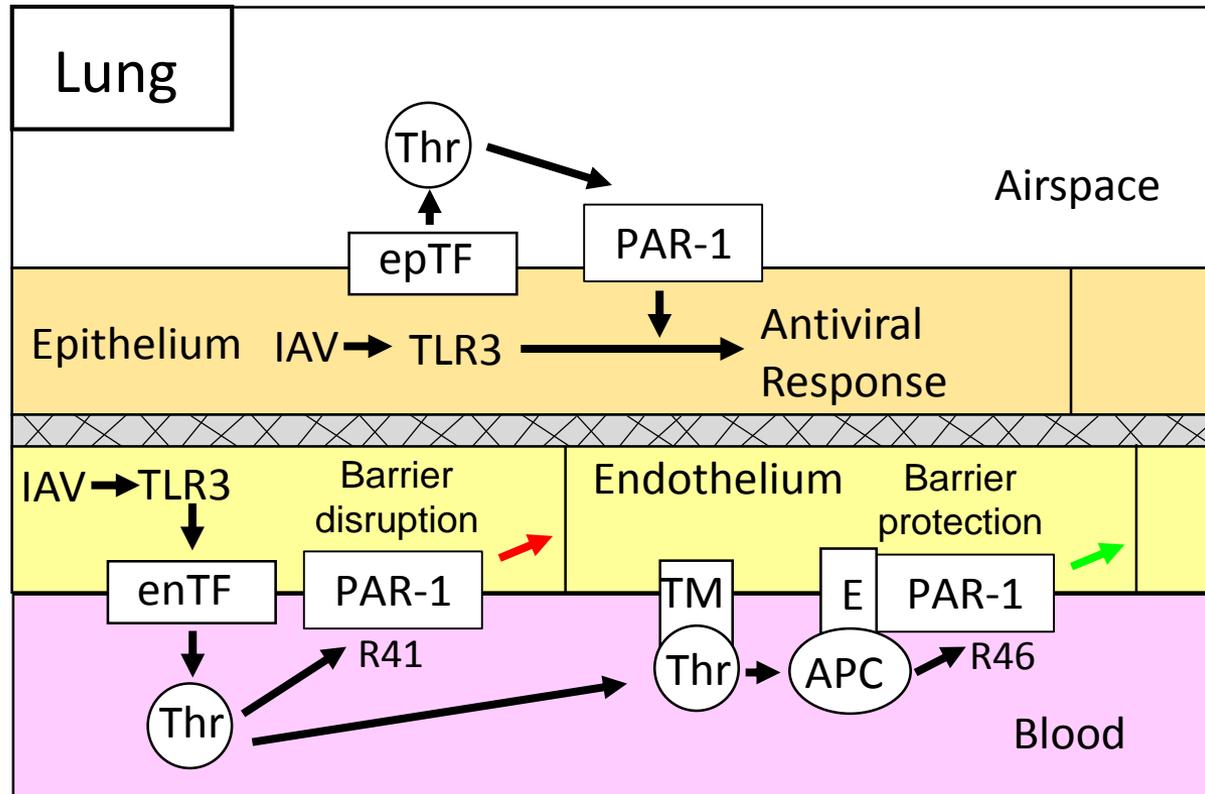
# Role of PAR-1 in Vascular Permeability in the Lung after Influenza A Infection



7 days

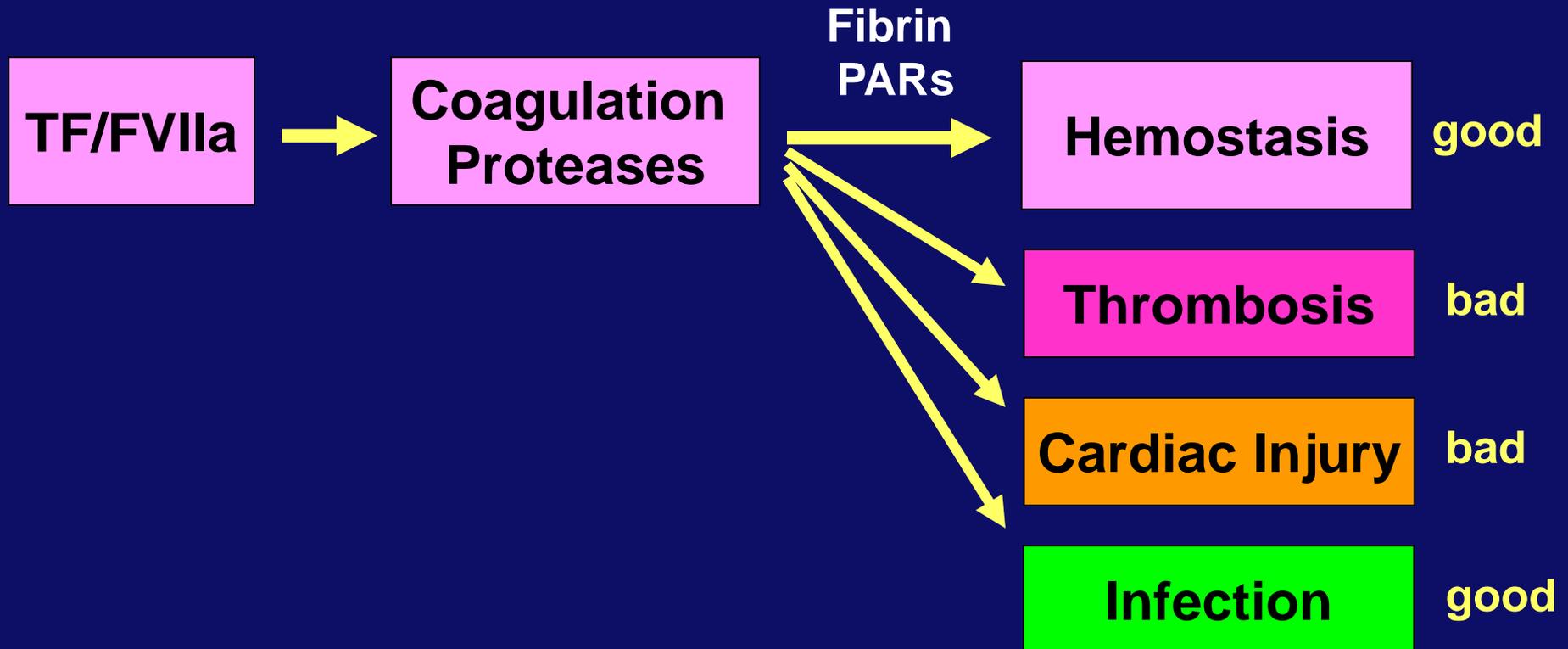
Tatsumi K et al. unpublished data

# Roles of the TF-Thrombin-PAR-1 Pathways in Influenza A Infection

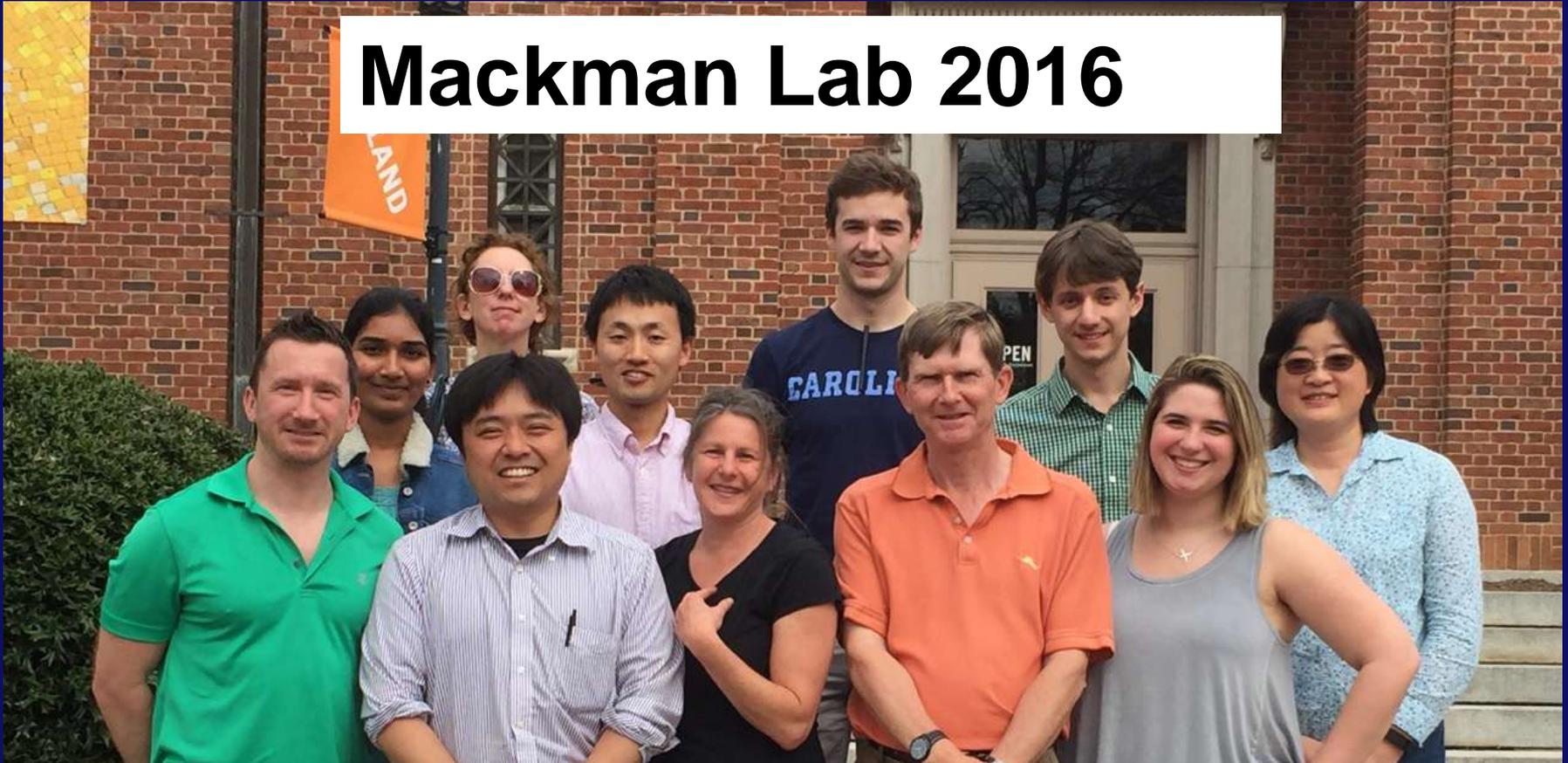


# Tissue Factor, Coagulation Proteases, Fibrin and PARs in Health and Disease

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# Mackman Lab 2016



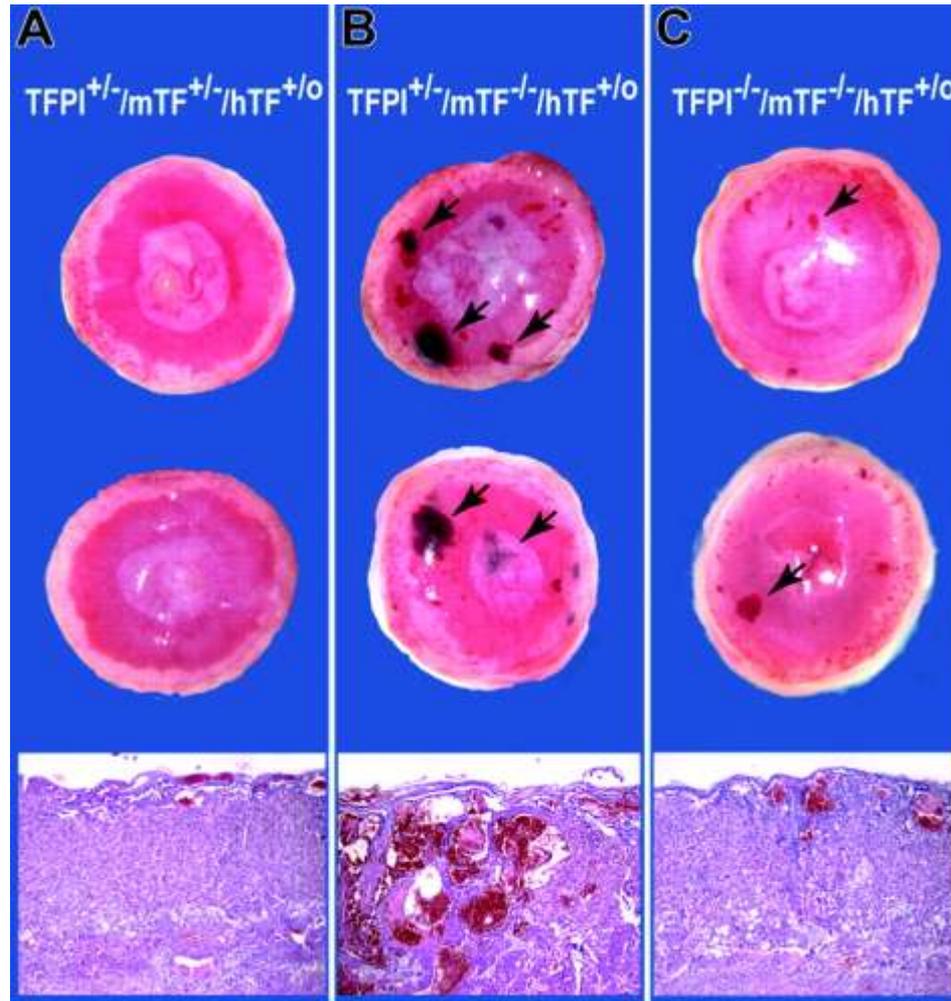
TraCS





**Biggs R and Nossel H Tissue  
extract and the contact reaction in  
blood coagulation Thromb Diath  
Haeofrrh 61: 1, 1961**

# Reduced Blood Pools in “Rescued” Placentas



Pedersen B et al Blood 2005

Undergraduates

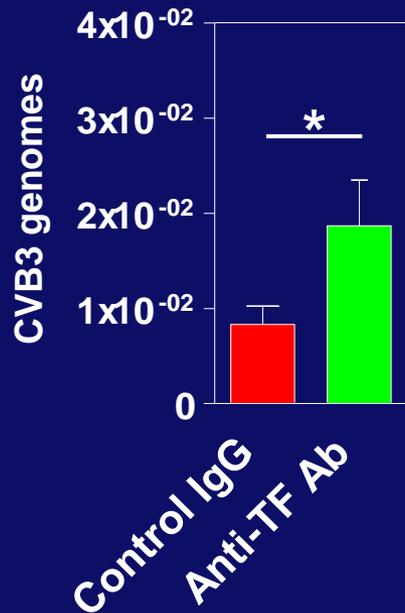


Janet Yan

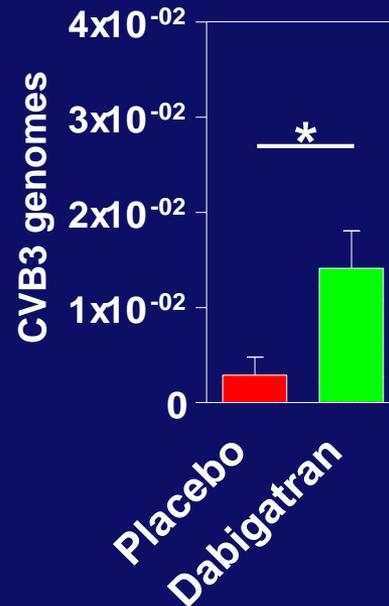


# Role of the TF-thrombin-PAR-1 Pathway in CVB3-induced Myocarditis

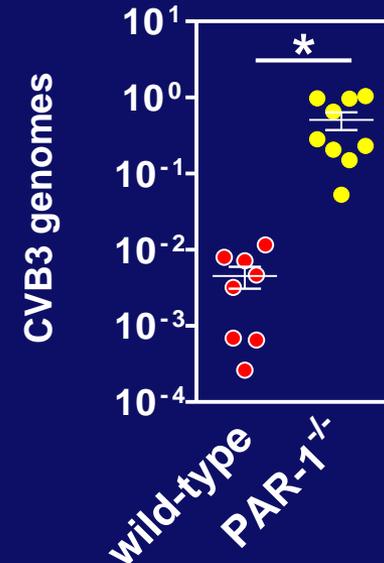
anti-TF Ab  
(1H1)



Dabigatran



PAR-1 Deficiency



# Microvesicles

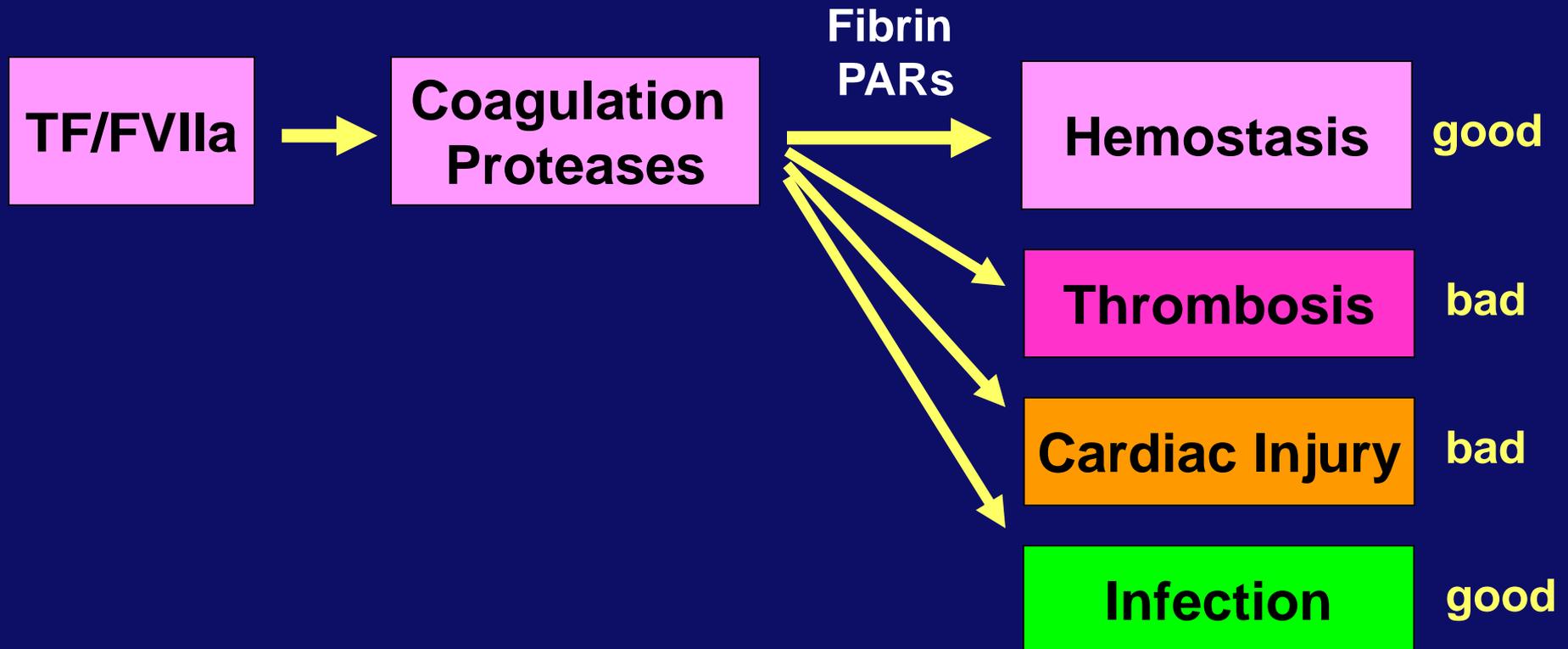
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**Microvesicles (MVs) were originally defined as small (0.1-1  $\mu\text{m}$ ) membrane vesicles that are released from activated or apoptotic cells**

**MVs are also referred to as microparticles (MPs) or extracellular vesicles (EVs) (= any type of vesicle released from a cell)**

# Tissue Factor, Coagulation Proteases, Fibrin and PARs in Health and Disease

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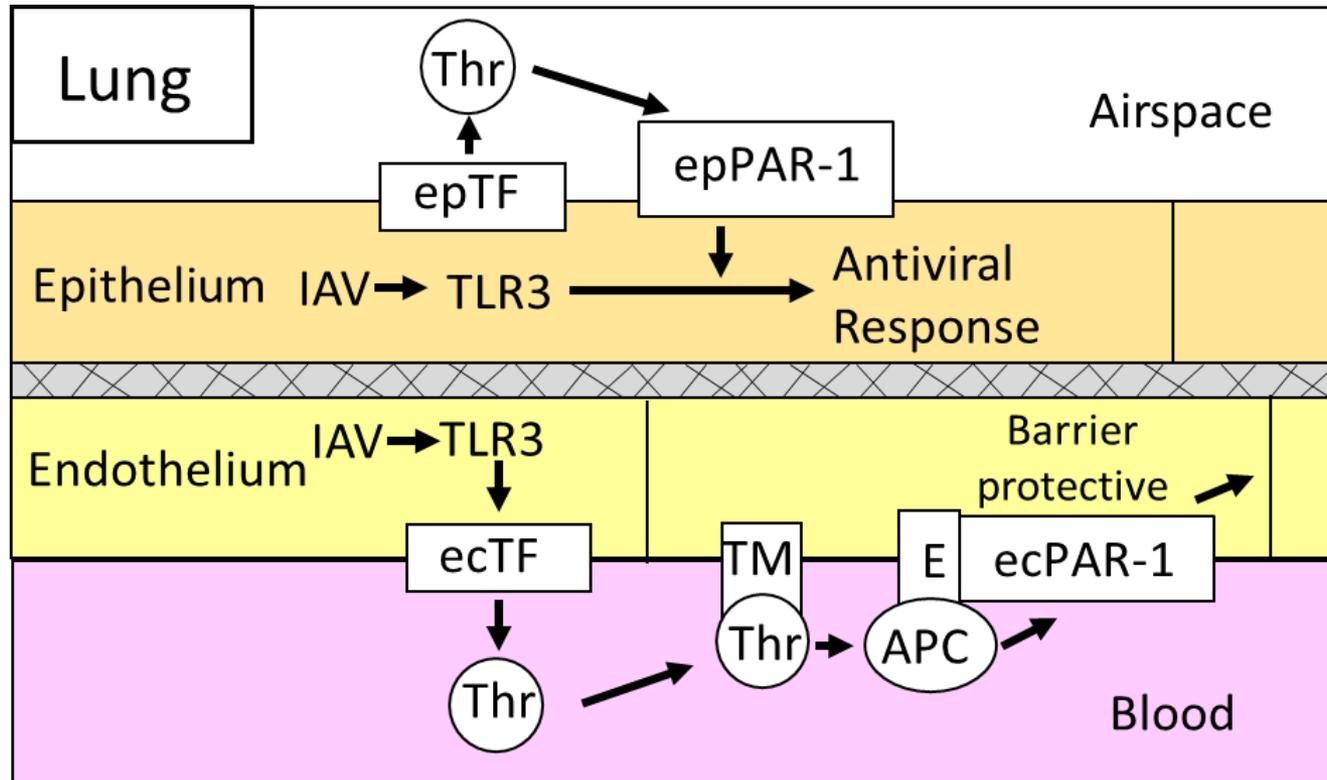


# Conclusions

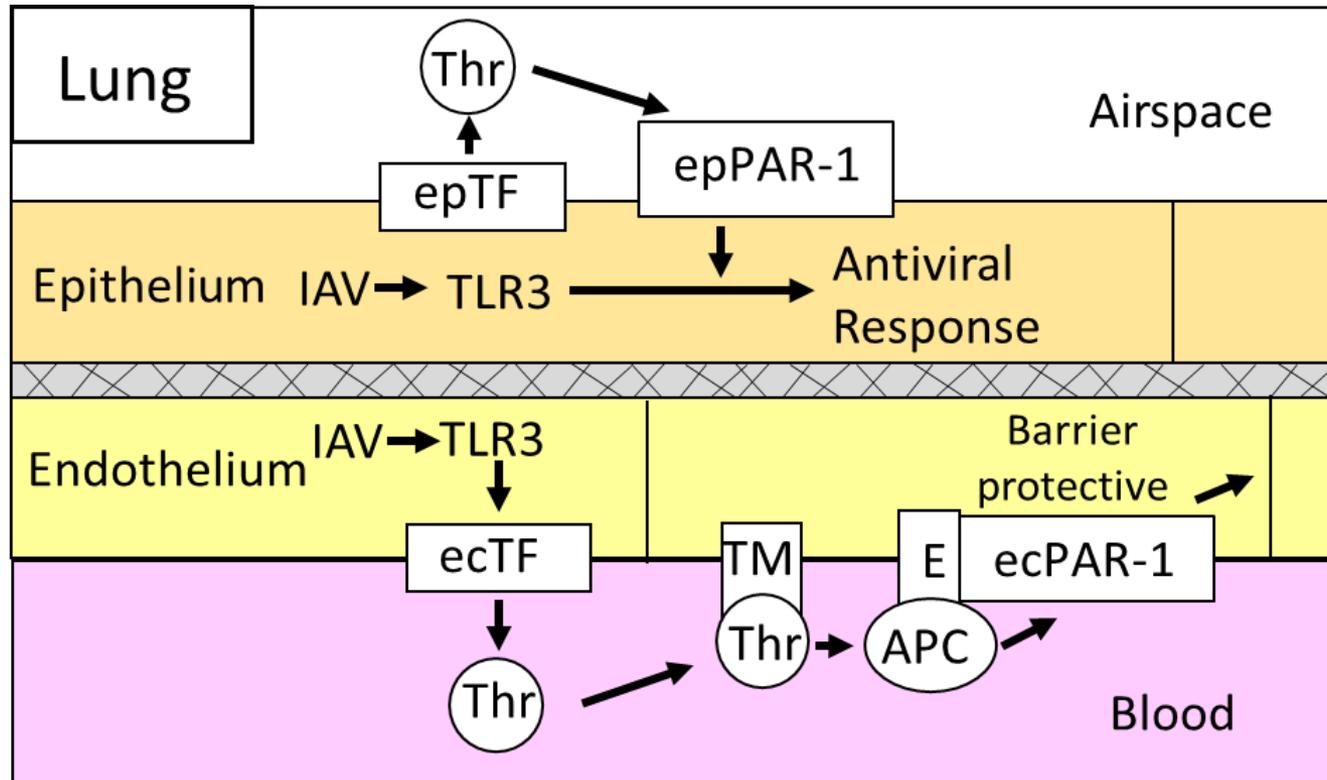
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- PAR-1 deficiency increases IAV-induced mortality.
- PAR-1 deficiency increases vascular permeability in the lung after IAV infection.
- PAR-1 also contributes to the immune response to IAV infection.

# Roles of the TF-Thrombin-PAR-1 Pathways in Influenza A Infection



# Roles of the TF-Thrombin-PAR-1 Pathways in Influenza A Infection



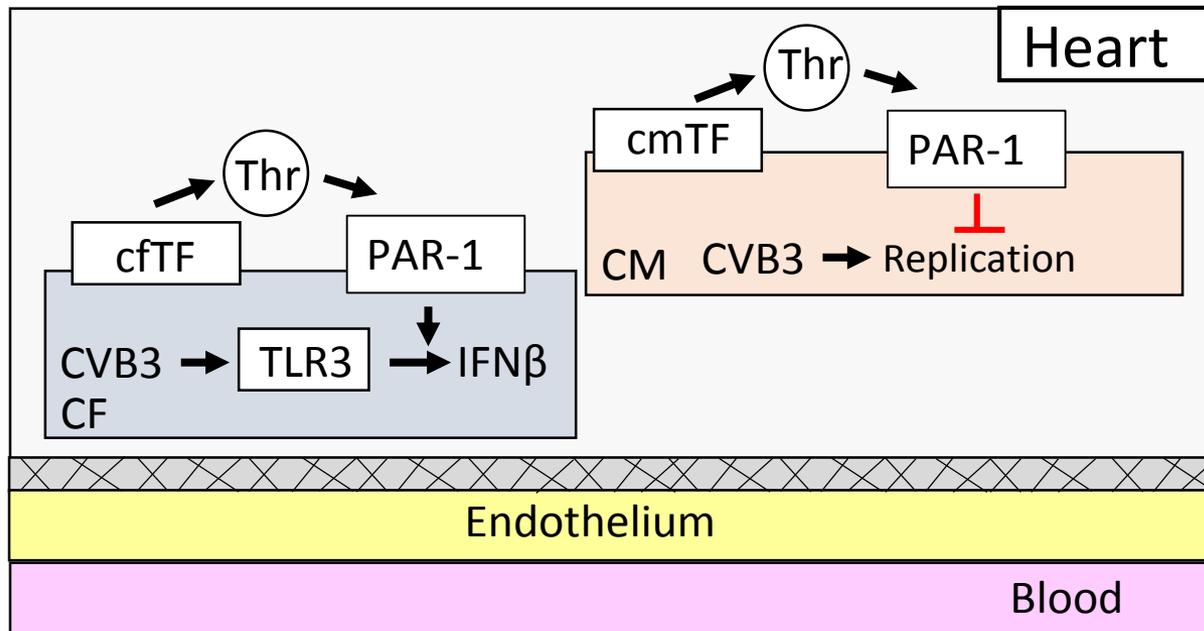
# Statins, TF and Thrombosis

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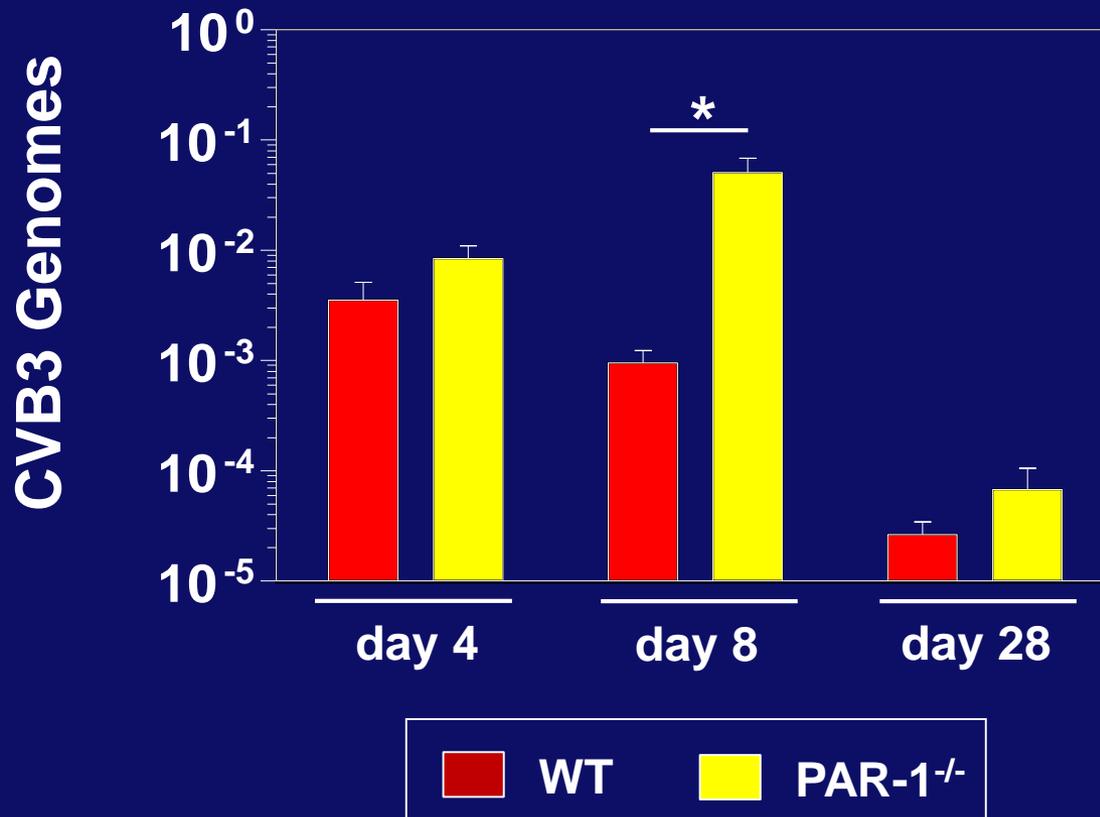
**Statins inhibit TF expression in activated monocytes in vitro and in vivo (Colli et al. 1997 ATVB, Ferro et al. Lancet 1997, Ferro et al. 2000 Atherosclerosis, Nagata et al. 2002 Atherosclerosis, Steiner et al. 2005 Circulation)**

**Can simvastatin reduce monocyte TF expression sepsis and other diseases?**

# Roles of the TF-Thrombin-PAR-1 Pathways in CVB3-induced Myocarditis



# PAR-1 Deficiency is Associated with Increased Viral Load in the Heart after CVB3 infection



\*P<0.05

Two-Way ANOVA

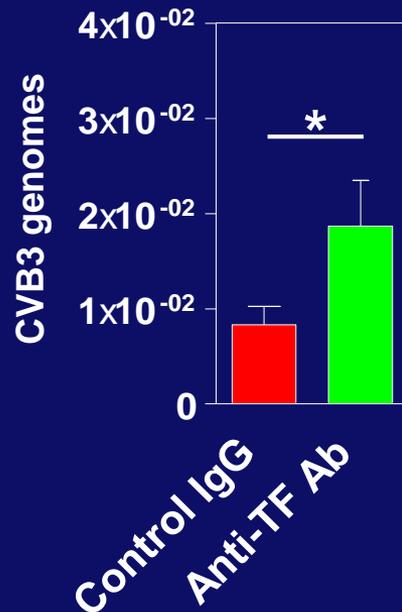
Antoniak S et al, JCI 2013

# Effect of Inhibition of Tissue Factor or Thrombin on CVB3-induced Myocarditis

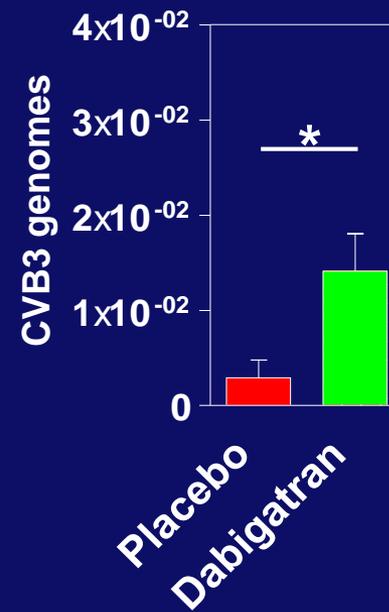
## Anti-TF Ab

## Dabigatran

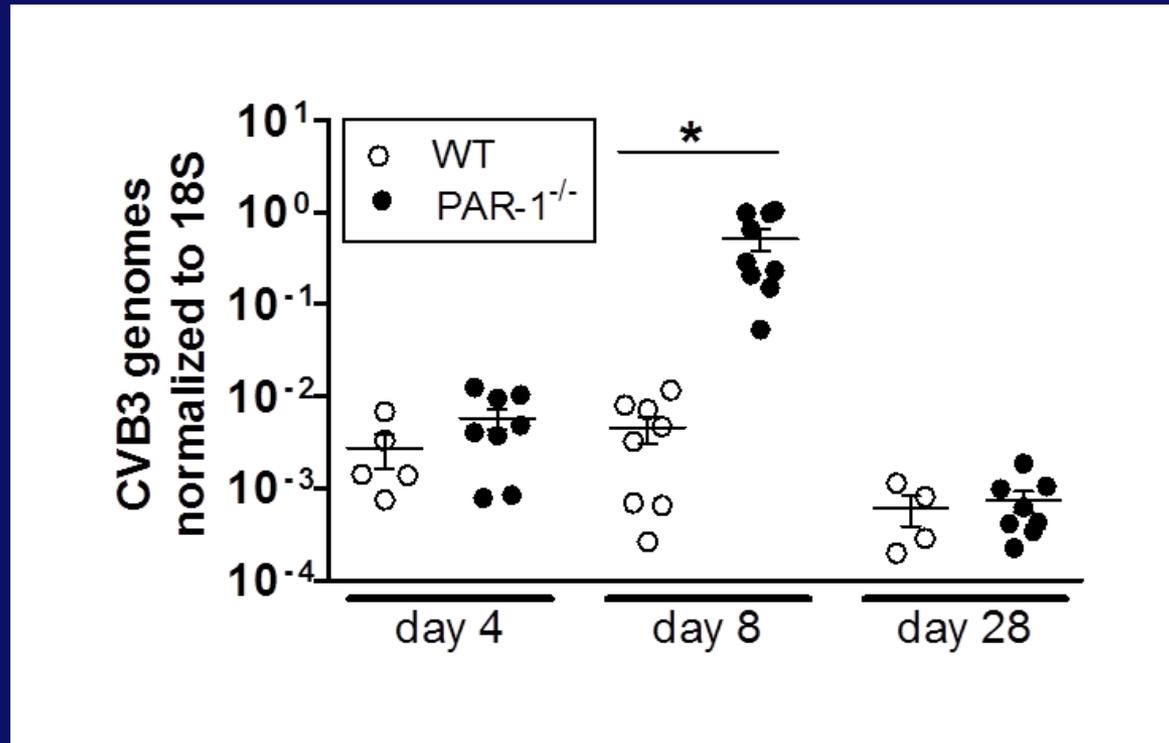
day 8



day 8



# PAR-1 Deficiency is Associated with Increased Viral Load in the Heart after CVB3 infection



\*P<0.05

Two-Way ANOVA

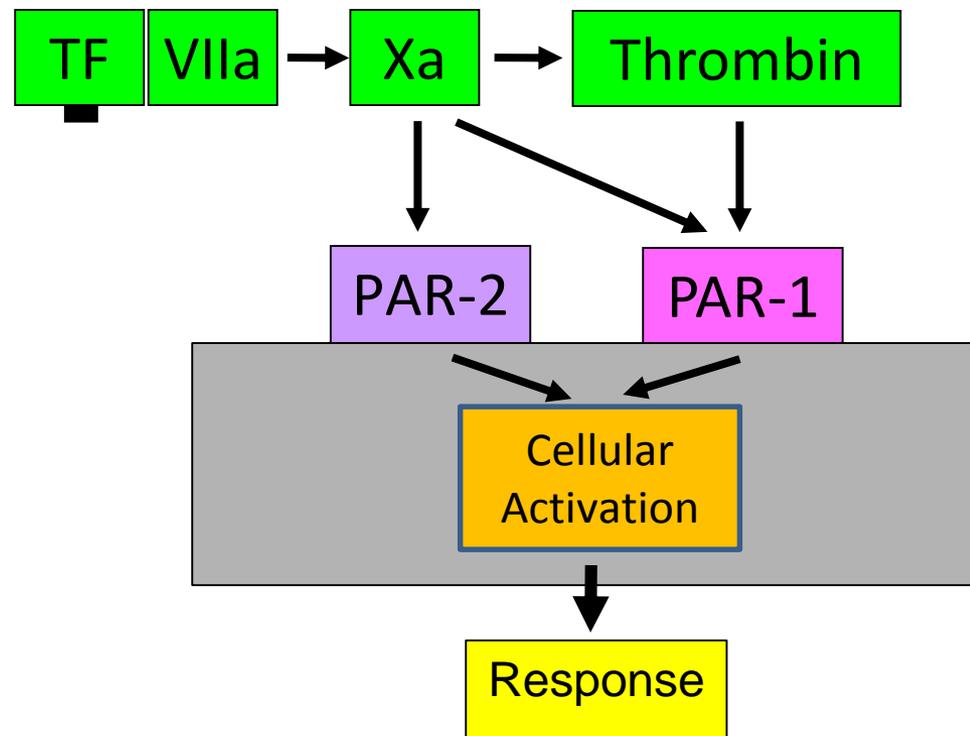
Antoniak et al, JCI 2013

# Outline

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- **Coxsackievirus B3 virus model**
- **Influenza A virus model**

# Role of Protease-activated Receptors as Sensors of the Extracellular Environment

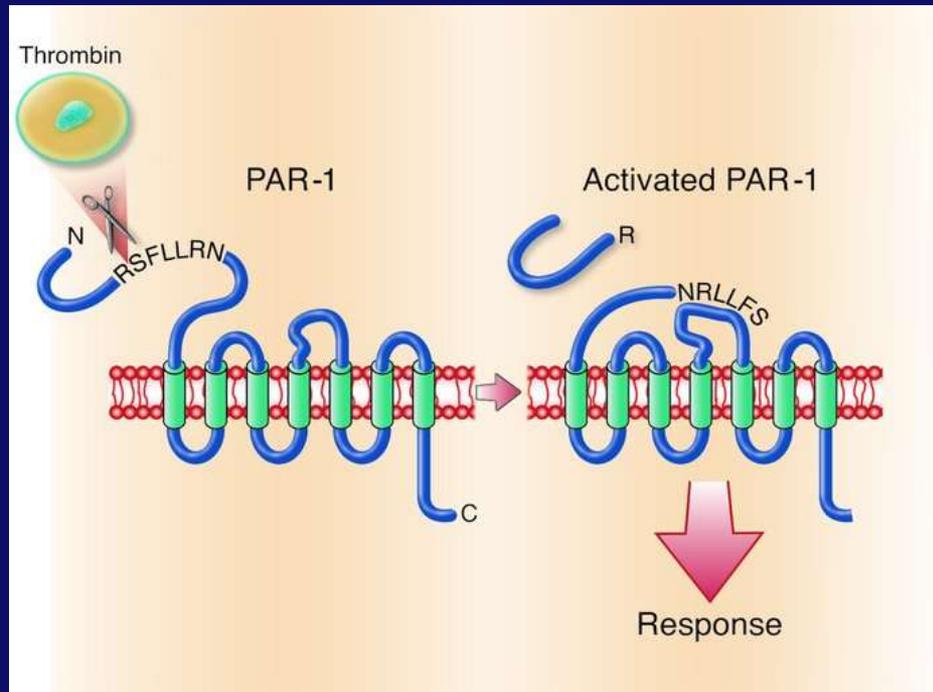


# The Protease-Activated Receptor Family

	PAR-1	PAR-2	PAR-3	PAR-4
Primary activating proteases (EC <sub>50</sub> )	Thrombin (50 pM)	Trypsin (1 nM) Tryptase (1 nM)	Thrombin (0.2 nM)	Thrombin (5 nM) Trypsin (5 nM)
Secondary activating proteases	Granzyme A FXa Trypsin Plasmin MMP-1	FVIIa FXa MT-SP1		Cathepsin G
PAR-APs	SFLLRN TFLLRN	SLIGKV SLIGIRL SFLLRN		GYPGKF AYPGKF

Adapted from Major et al., 2003 *Arterioscler. Thromb. Vasc. Biol.*

# Thrombin Activation of PAR-1



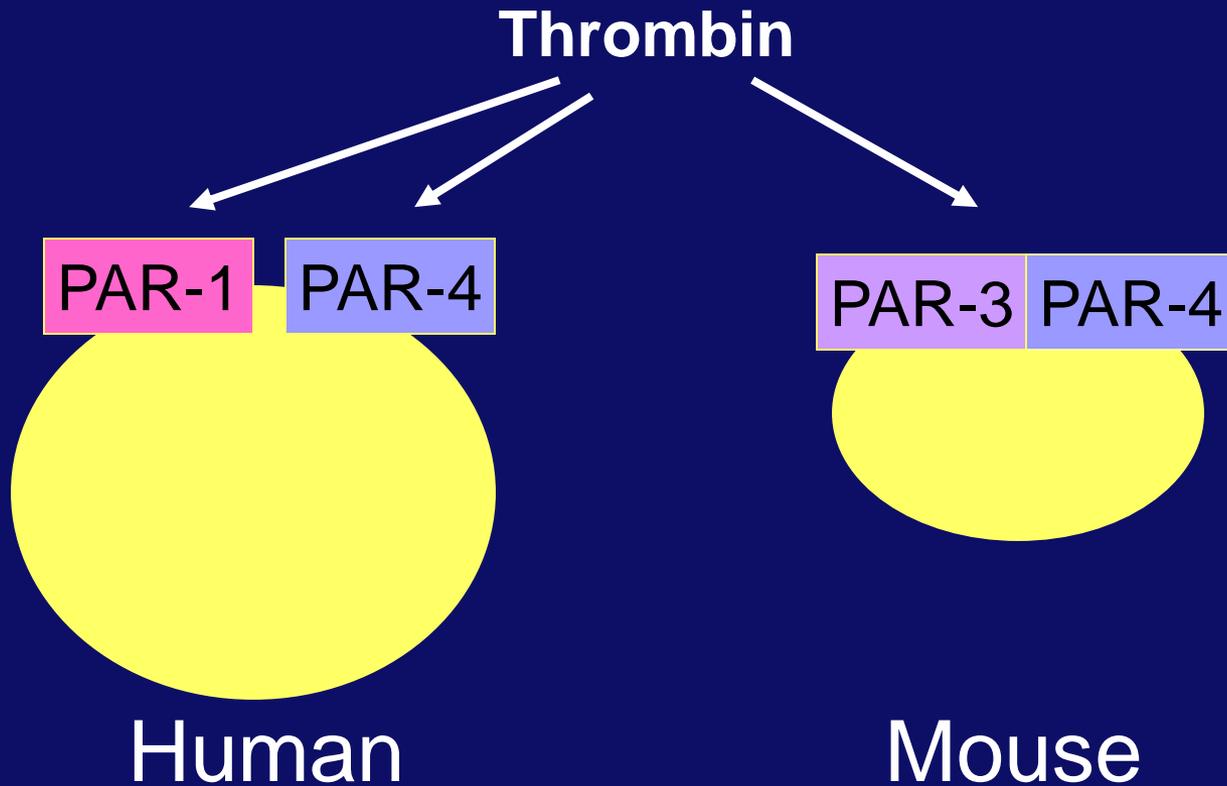
**PAR-1 is a unique GPCR because it is activated by proteolytic cleavage. It is the major thrombin receptor on human platelets.**

**Vorapaxar (Zontivity) was approved by the FDA in 2014 for use with daily aspirin and/or clopidogrel.**

**Camerer and Coughlin JCI 2003**

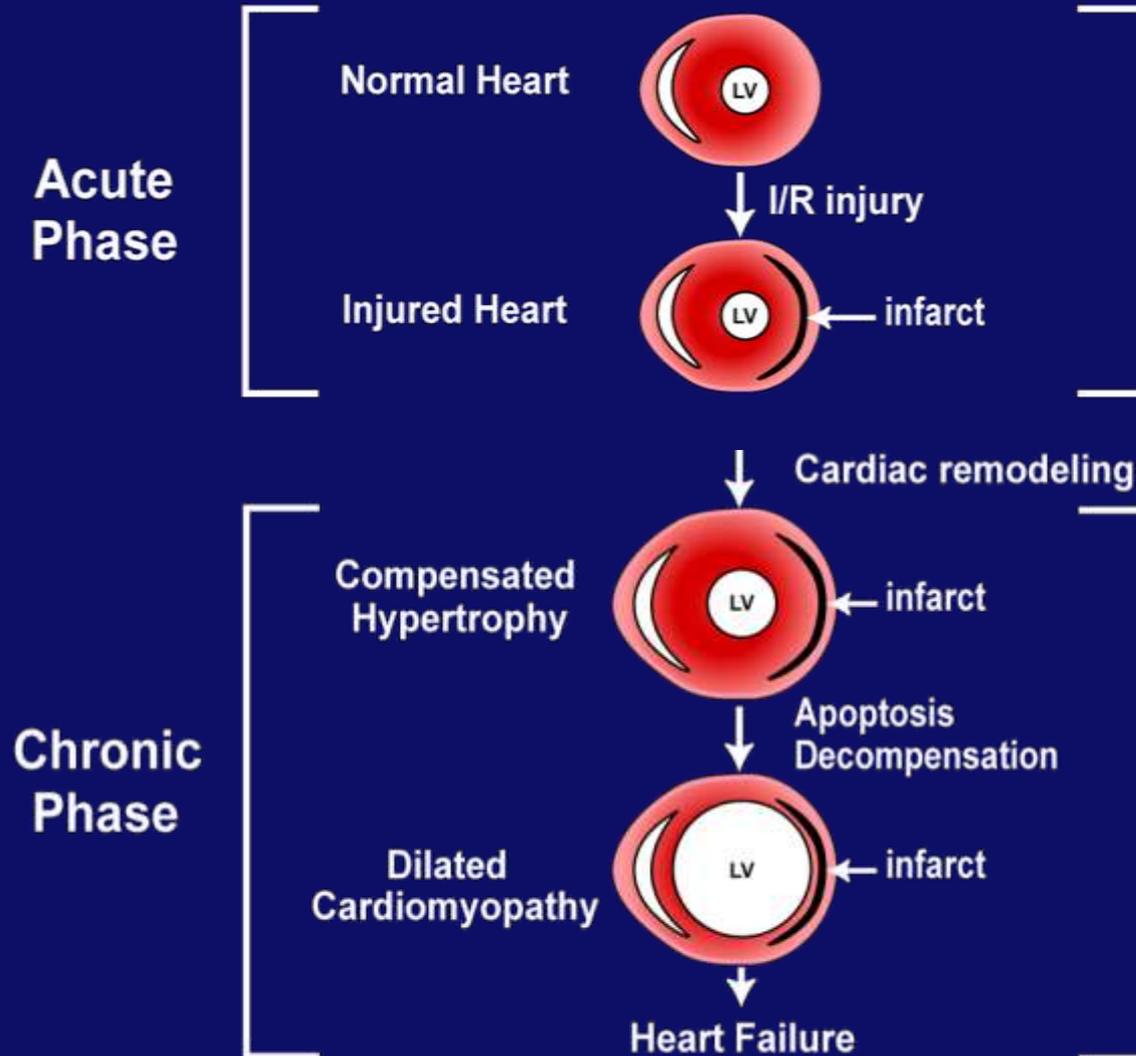
# PAR Expression on Human and Mouse Platelets

---



Any phenotype in PAR-1<sup>-/-</sup> mice cannot be due to a defect in thrombin activation of platelets

# Cardiac I/R Injury Induces Infarction and Remodeling



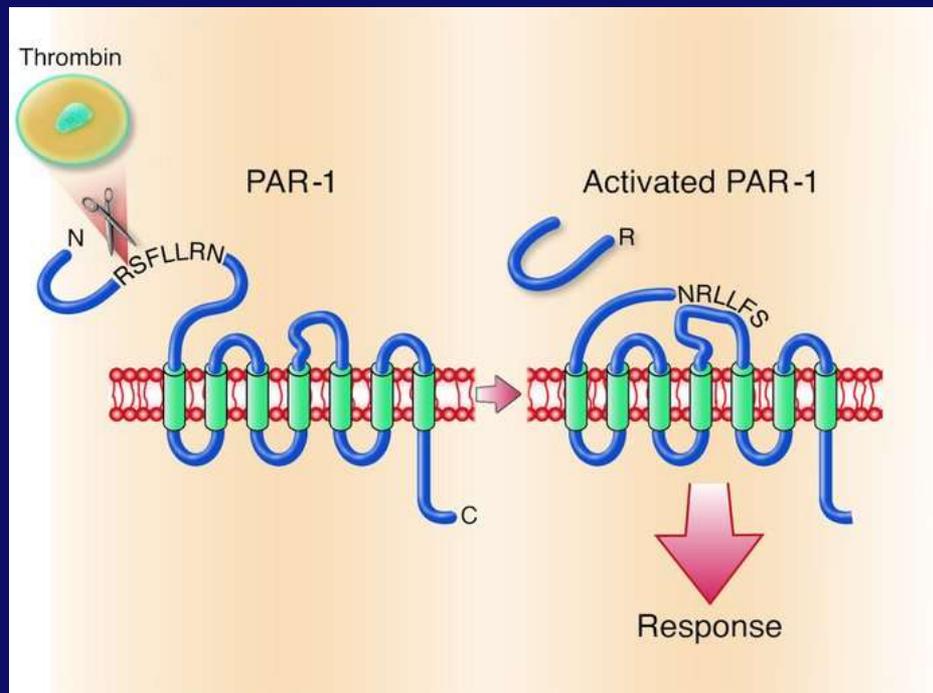


# Viral Infections and the Clotting Cascade

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- Acute viral infections lead to strong activation of the clotting cascade and secondary hemorrhage (i.e. Ebola hemorrhagic fever).
- Inhibition of TF reduced death in a monkey model of Ebola.
- Chronic viral infections lead to a low level activation of the clotting cascade that increases cardiovascular disease (i.e. HIV).
- Two ongoing clinical trials are targeting FXa and PAR-1 in HIV patients.

# Thrombin Activation of PAR-1



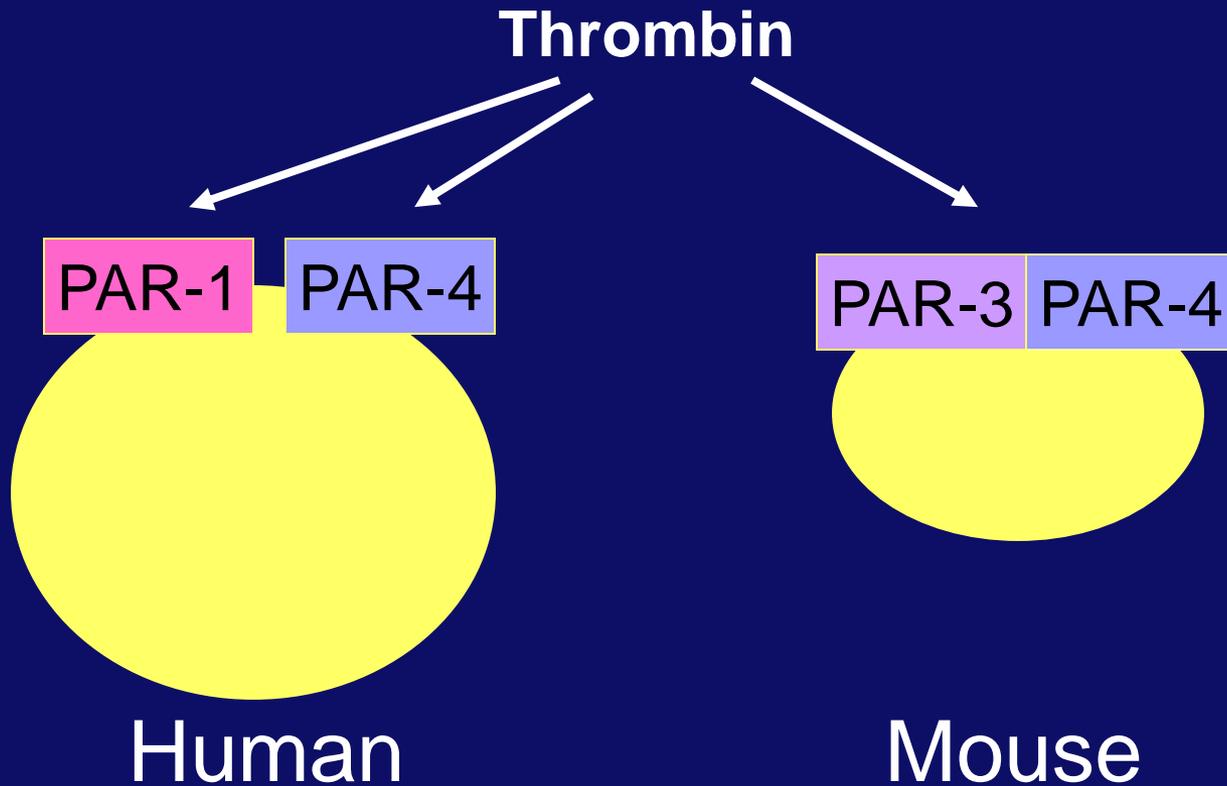
**PAR-1 is a unique GPCR because it is activated by proteolytic cleavage. It is the major thrombin receptor on human platelets.**

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**Camerer and Coughlin JCI 2003**

# PAR Expression on Human and Mouse Platelets

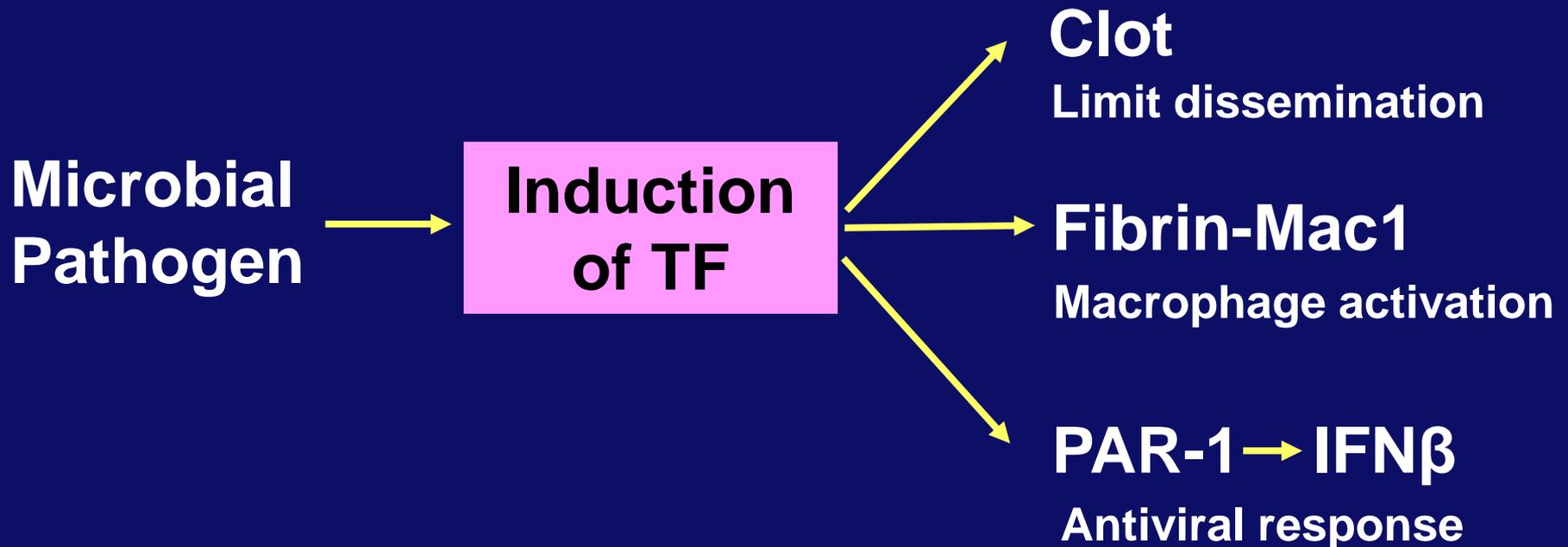
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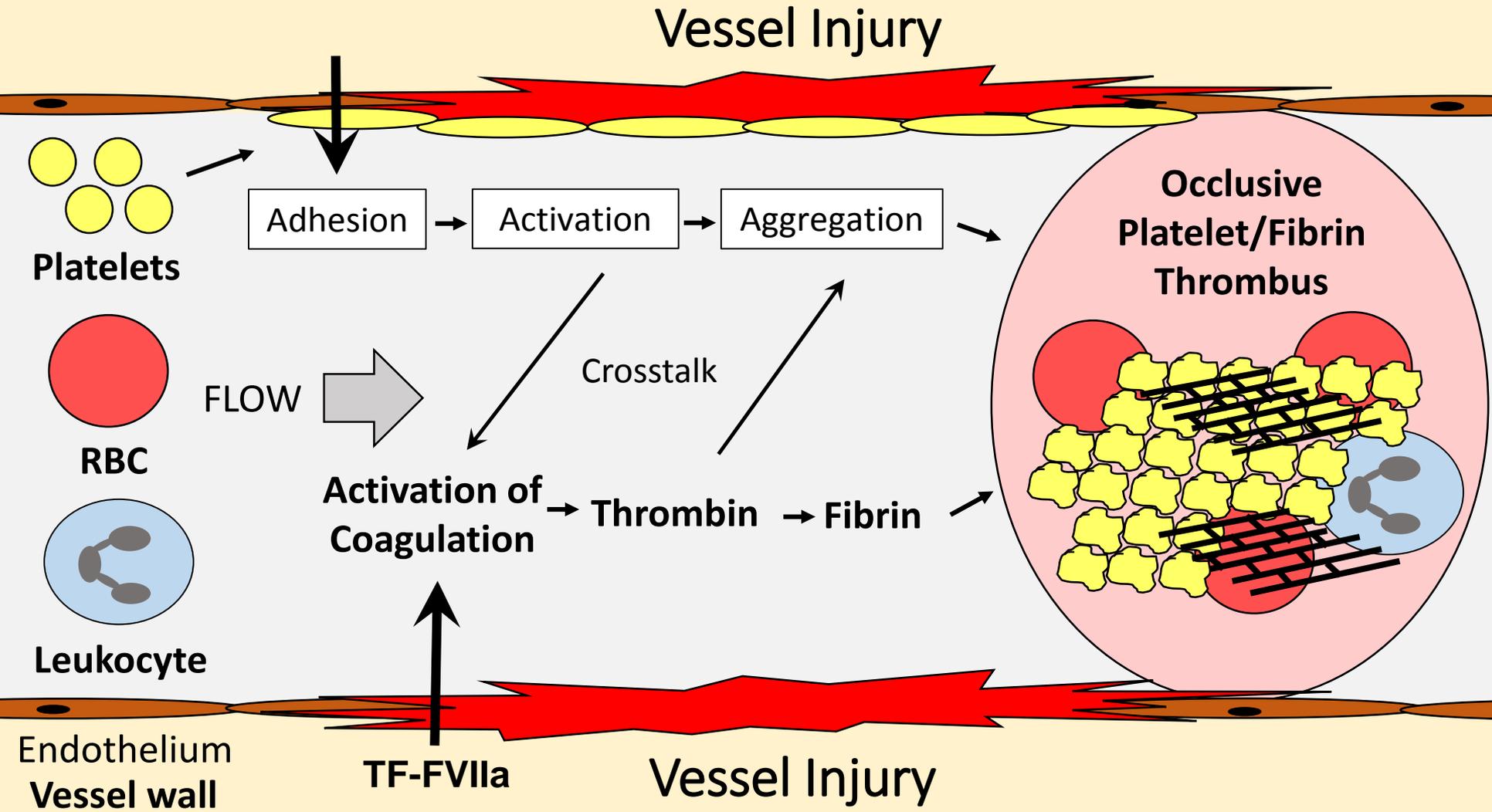
Any phenotype in PAR-1<sup>-/-</sup> mice cannot be due to a defect in thrombin activation of platelets

# The Clotting Cascade and Host Defense

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# Formation of an Occlusive Thrombus



# Ischemia-Reperfusion Injury

---

**Ischemia**

**Reperfusion**



**Hypoxic Damage**

**Inflammatory Damage**

**Apoptosis/necrosis**

**Chemokines, cytokines,  
oxidants**

**Leukocyte recruitment  
and activation**

**Coagulation and fibrin  
deposition**

**Complement activation**

# Conclusions

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- PAR-1 contributes to the antiviral response to CVB3 infection.
- PAR-1 enhances poly I:C induction of IFN $\beta$  expression in cardiac fibroblasts.
- PAR-1 inhibits CVB3 replication in cardiac myocytes.

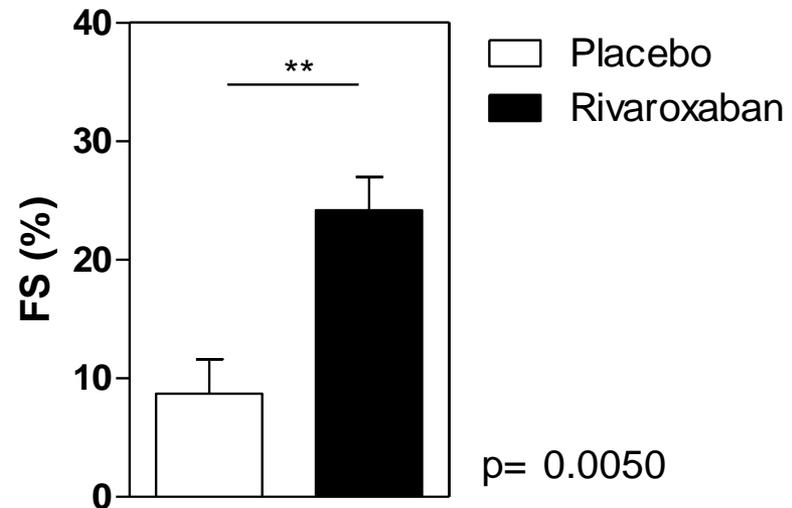
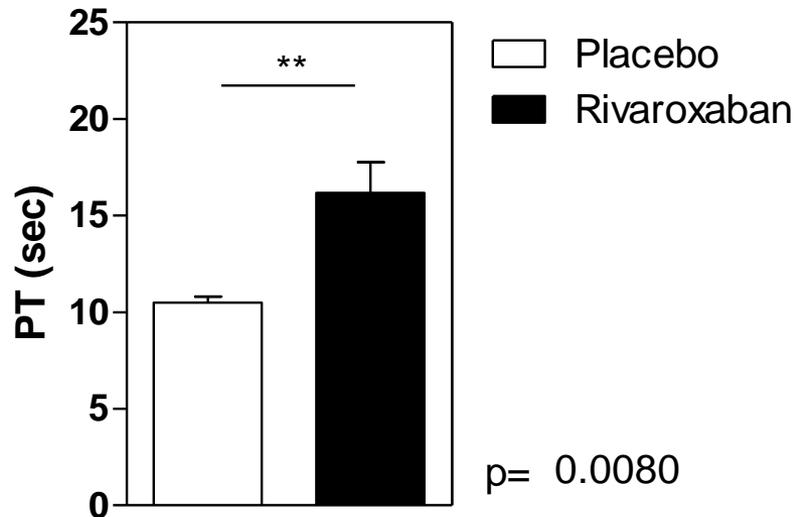
**I/R**

# Conclusion

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**Inhibition of TF, coagulation proteases or PAR-1 may reduce cardiac remodeling and heart failure in MI patients**

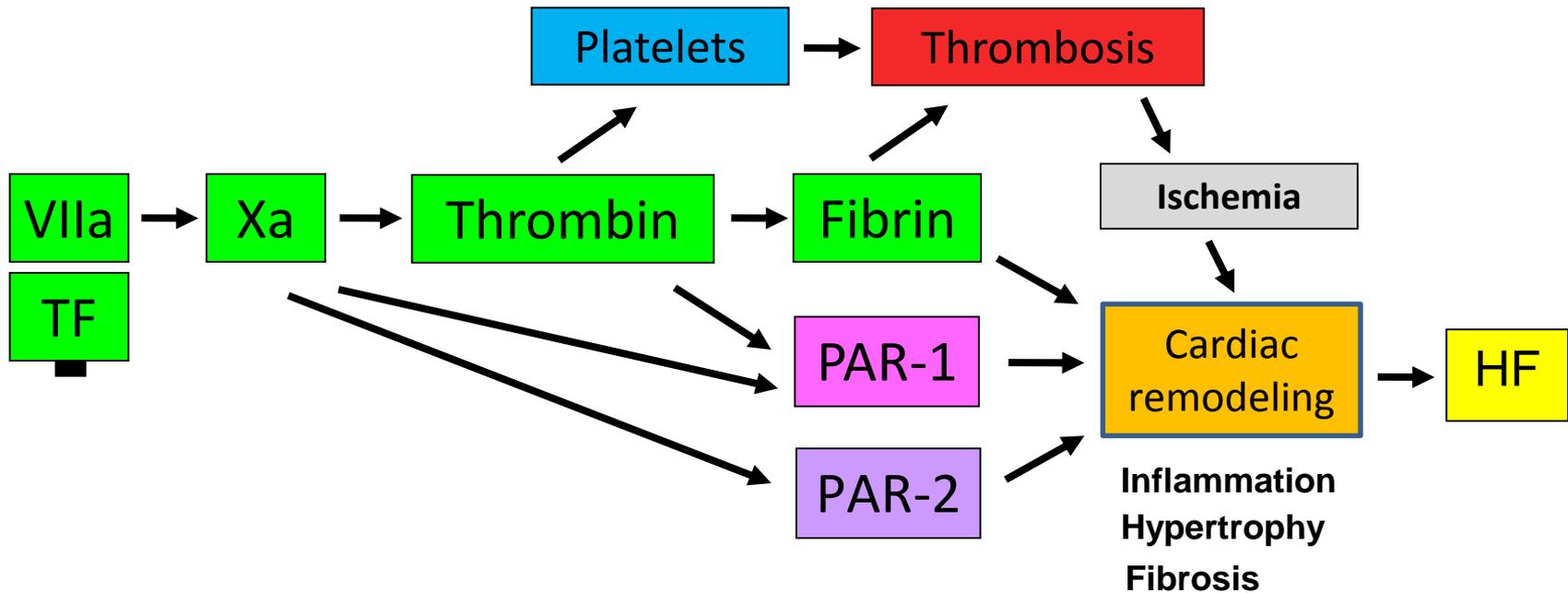
# Effect of Rivaroxaban on Cardiac Remodeling and Heart Failure in a Mouse Model of LAD Ligation



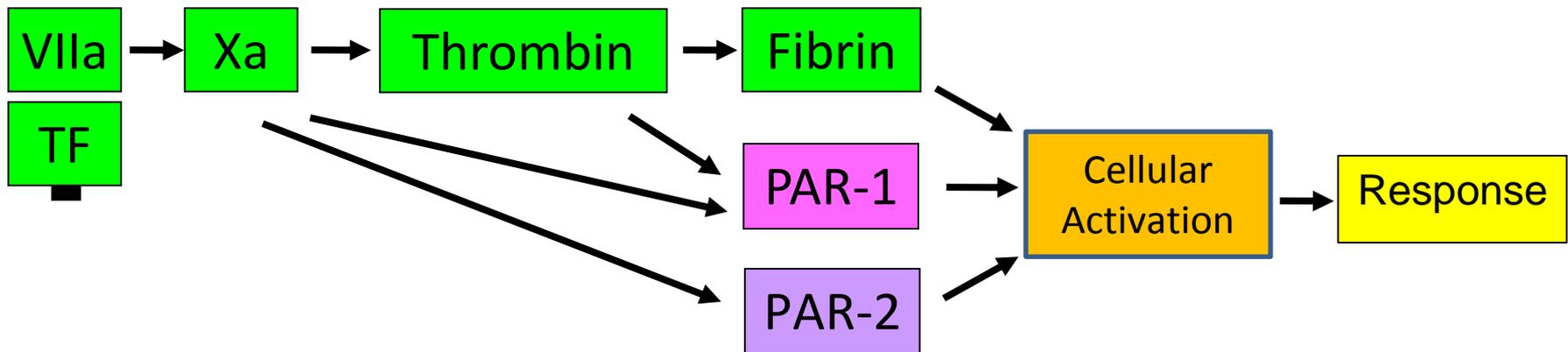
28 days after LAD ligation

Bode M unpublished results

# Coagulation Proteases and PARs in Cardiac Remodeling and Heart Failure

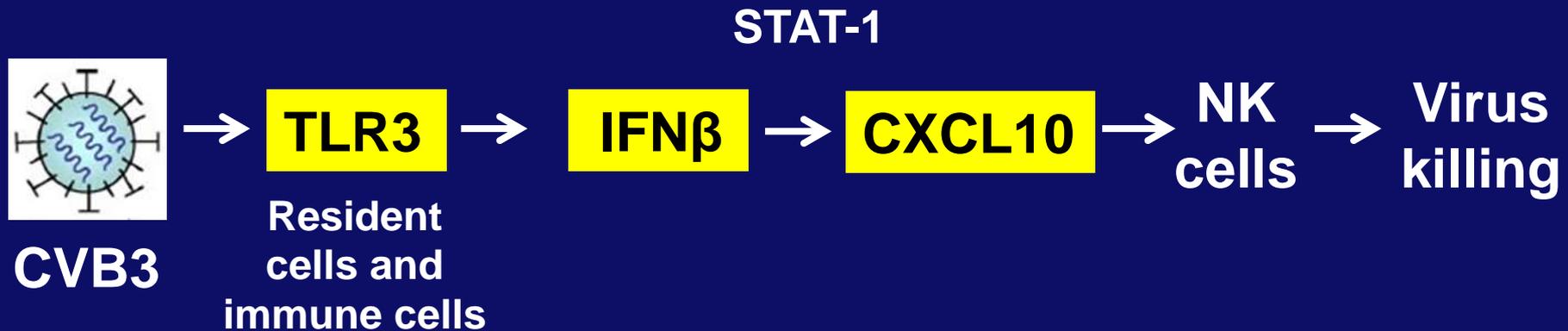


# Role of Protease-activated Receptors as Sensors of the Extracellular Environment

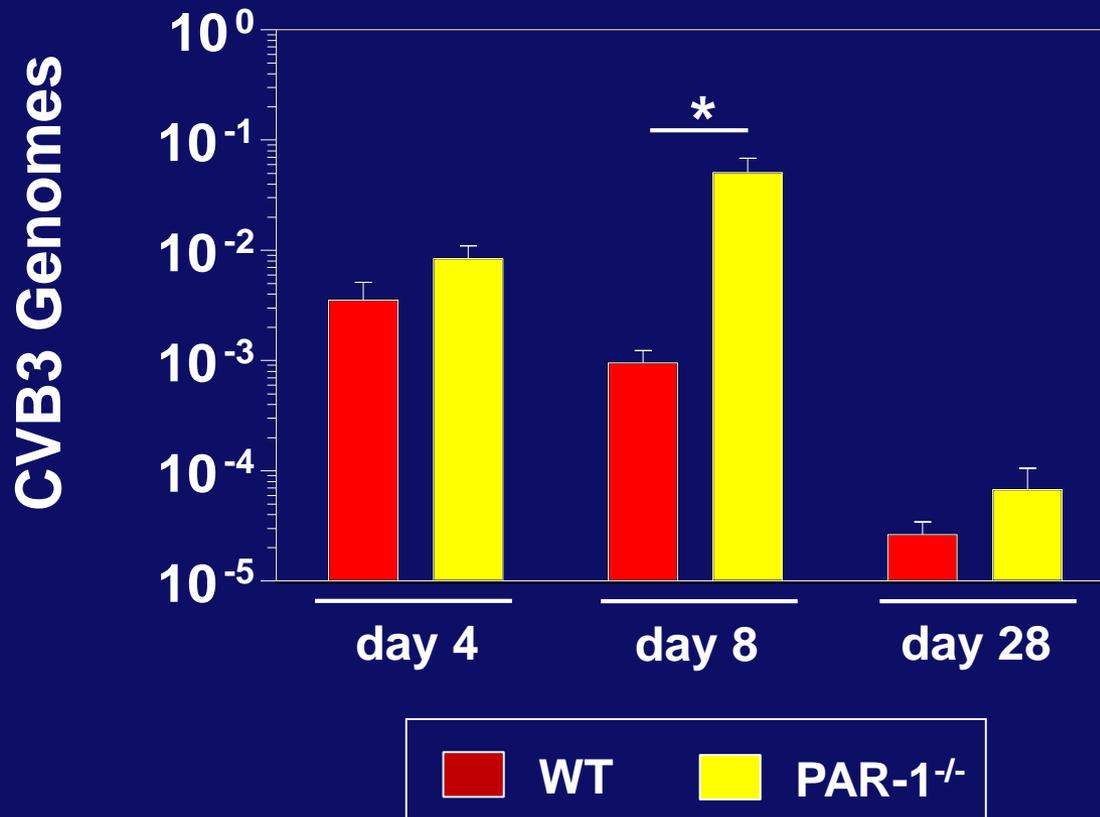


# The Innate Immune Response to Coxsackievirus B3 Infection

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# PAR-1 Deficiency is Associated with Increased Viral Load in the Heart after CVB3 infection

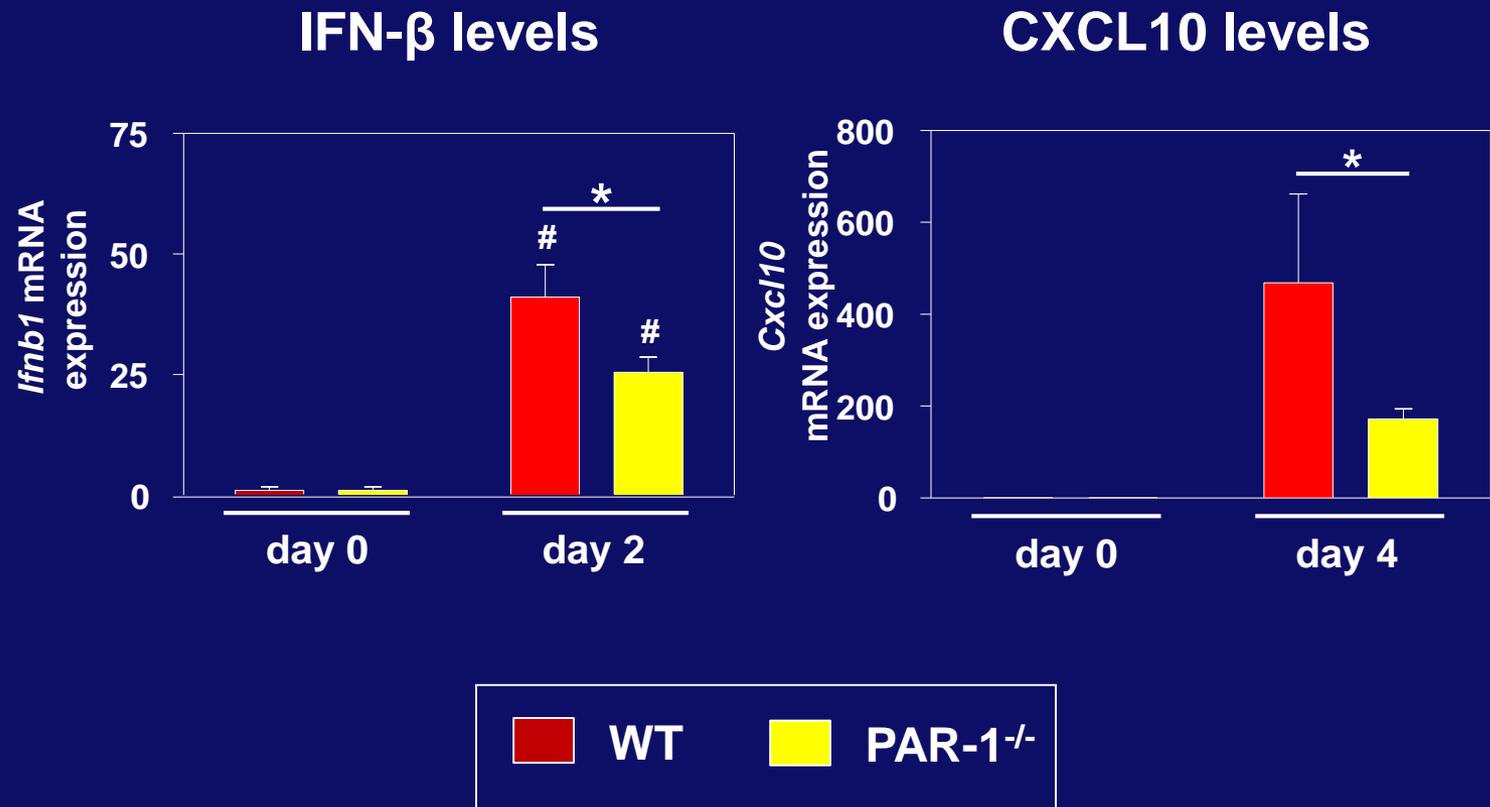


\*P<0.05

Two-Way ANOVA

Antoniak et al, JCI 2013

# PAR-1 Deficiency is Associated with Reduced IFN- $\beta$ and CXCL10 expression After CVB3 Infection



\*P < 0.05; #P < 0.05 vs. day 0  
Two-Way ANOVA

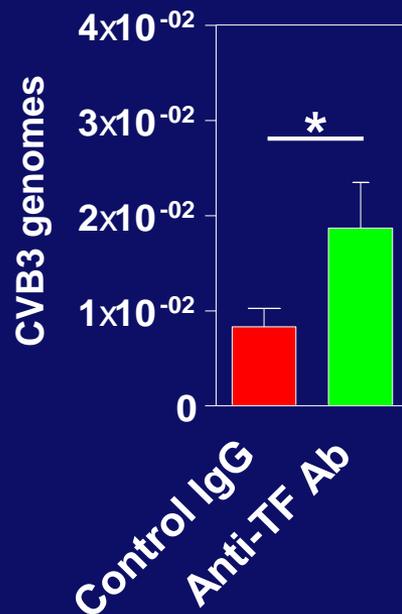
Antoniak et al, JCI 2013

# Effect of Inhibition of Tissue Factor or Thrombin on CVB3-induced Myocarditis

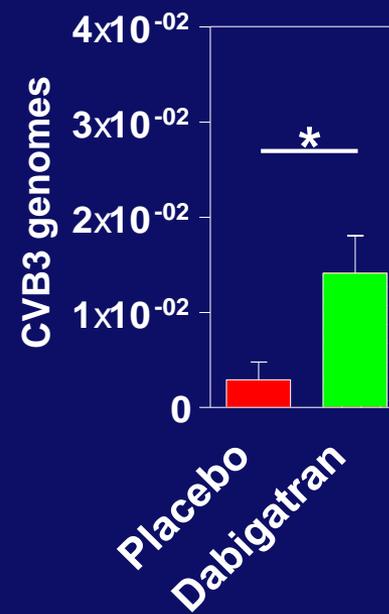
## Anti-TF Ab

## Dabigatran

day 8



day 8

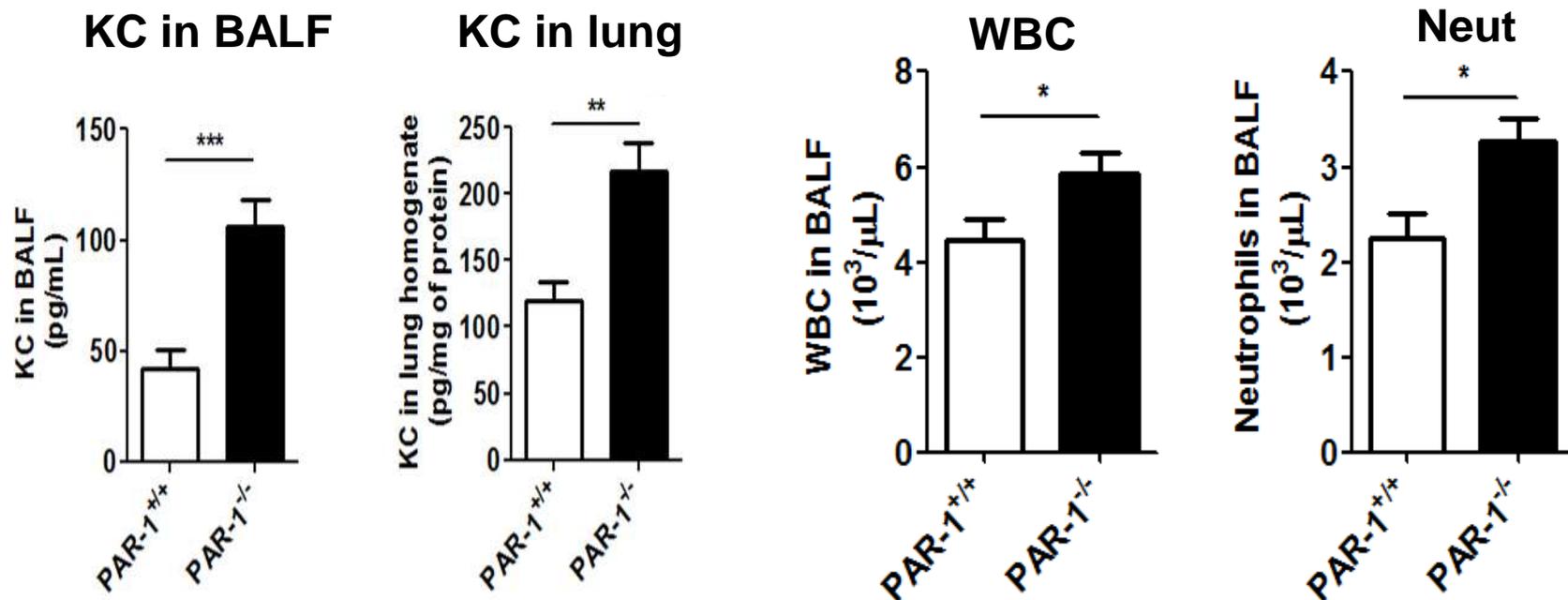


# Ongoing Studies

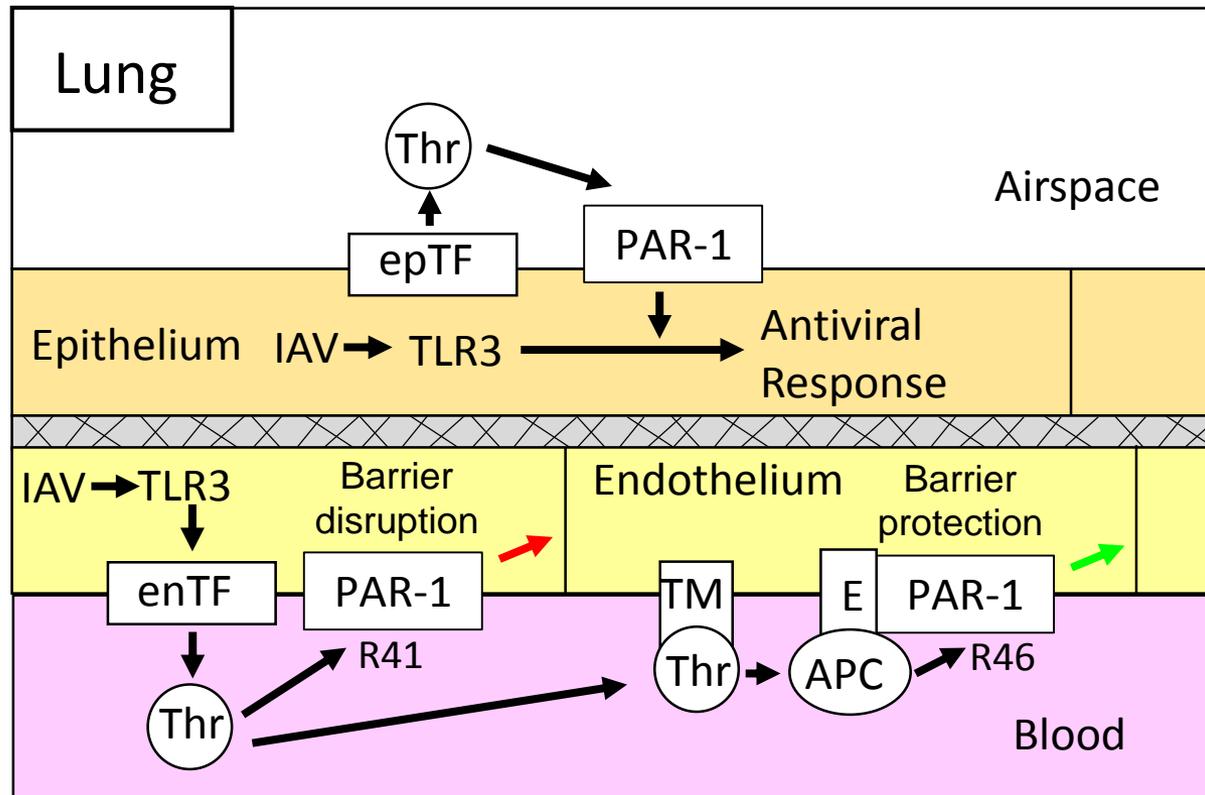
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- Determine the effect of the following on IAV-induced vascular permeability in the lung:
- (i) deletion of PAR-1 in endothelial cells.
  - (ii) EPCR deficiency.
  - (iii) mutation of the APC cleavage site in PAR-1 (R46Q).

# Effect of PAR-1 Deficiency on KC Expression and Leukocyte Recruitment after IAV Infection



# Roles of the TF-Thrombin-PAR-1 Pathways in Influenza A Infection



# Ongoing Studies

---

Determine the effect of deletion of PAR-1 in cardiac myocytes or cardiac fibroblasts on CVB3-induced myocarditis.

- PAR-1<sup>fl/fl</sup>,Mlc2v<sup>Cre</sup> (CMs)- 72% reduction in PAR-1 expression. Increased CVB3-induced myocarditis.
- PAR-1<sup>fl/fl</sup>,TCF21<sup>Cre-ERT2</sup> (CFs)

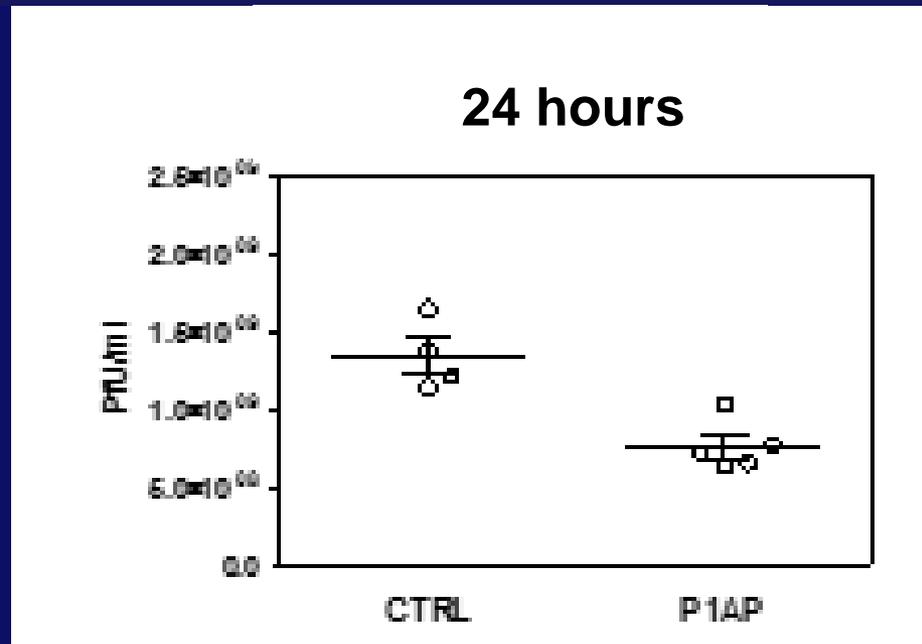
# Effect of PAR-1 Activation on Poly I:C Induction of IFN $\beta$ Expression in Cardiac Fibroblasts and Cardiac Myocytes



**How is CM PAR-1 protecting mice from CVB3-induced myocarditis?**

Bode M et al unpublished data

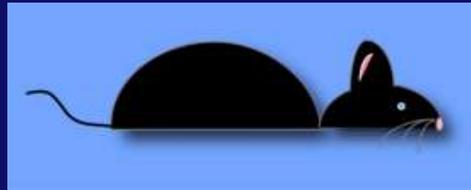
# PAR-1 Activation Inhibits CVB3 Replication in Cardiac Myocytes



Bode M et al unpublished data

# Mouse Models

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## **PAR-1<sup>fl/fl</sup> mice**

Cell type specific  
deletion of PAR-1  
(crossed with Cre<sup>+</sup>)

Palumbo J  
unpublished data

# Transgenic Mouse Lines

Table 1 Transgenic mouse lines

<u>Mouse line</u>	<u>Description</u>
PAR-1 <sup>-/-</sup>	Δ All cells
PAR-1 <sup>fl/fl</sup> ,Mlc2v <sup>cre</sup>	Δ CMs
PAR-1 <sup>fl/fl</sup> ,TCF21 <sup>cre-ERT2</sup>	Δ CFs
PAR-1 <sup>fl/fl</sup> , <u>LysM<sup>cre</sup></u>	Δ Myeloid cells
PAR-1 <sup>fl/fl</sup> , <u>Alb<sup>cre</sup></u>	Δ Hepatocytes
PAR-1 <sup>R41Q</sup>	No thrombin activation
PAR-1 <sup>R46Q</sup>	No APC activation
PAR-1 <sup>fl/fl</sup> , <u>Spc<sup>cre</sup></u>	Δ Lung epithelial cells
PAR-1 <sup>fl/fl</sup> ,VE-cad <sup>cre-ERT2</sup>	Δ EC*
<u>Procr<sup>Lox</sup></u>	Δ EPCR in all cells

\* Currently being generated.

# Generation of Transgenic Mice Expressing PAR-1 Mutants

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John Griffin

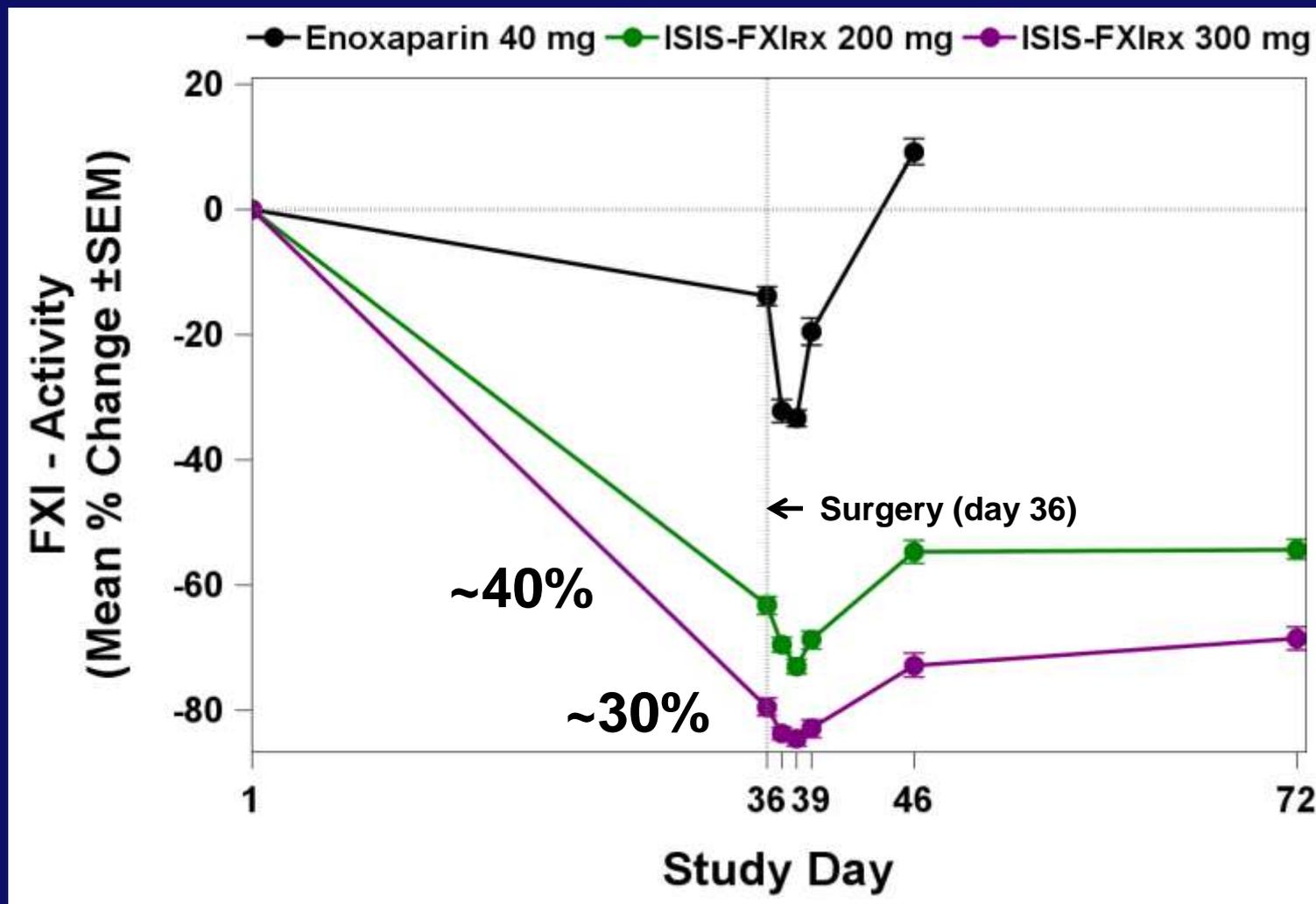
# Clinical Characteristics

	<b>Enoxaparin 40 mg (n =72)</b>	<b>ISIS-FXI<sub>Rx</sub> 200 mg (n = 144)</b>	<b>ISIS-FXI<sub>Rx</sub> 300 mg (n = 77)</b>
<b>Mean age – yr</b>	<b>64 ± 9</b>	<b>63 ± 9</b>	<b>63 ± 8</b>
<b>Female – no. (%)</b>	<b>60 (83%)</b>	<b>118 (82%)</b>	<b>60 (78%)</b>
<b>Mean Weight, kg (range)</b>	<b>87 (52, 132)</b>	<b>89 (52,124)</b>	<b>90 (52,130)</b>
<b>Creatinine clearance (ml/min)</b>	<b>111 ± 30</b>	<b>112 ± 31</b>	<b>116 ± 30</b>
<b>Mean factor XI activity (units/ml)</b>	<b>1.23 ± 0.21</b>	<b>1.20 ± 0.20</b>	<b>1.16 ± 0.22</b>

**ISIS-FXI<sub>Rx</sub> – FXI ASO**

**Buller H et al N Engl J Med 2015**

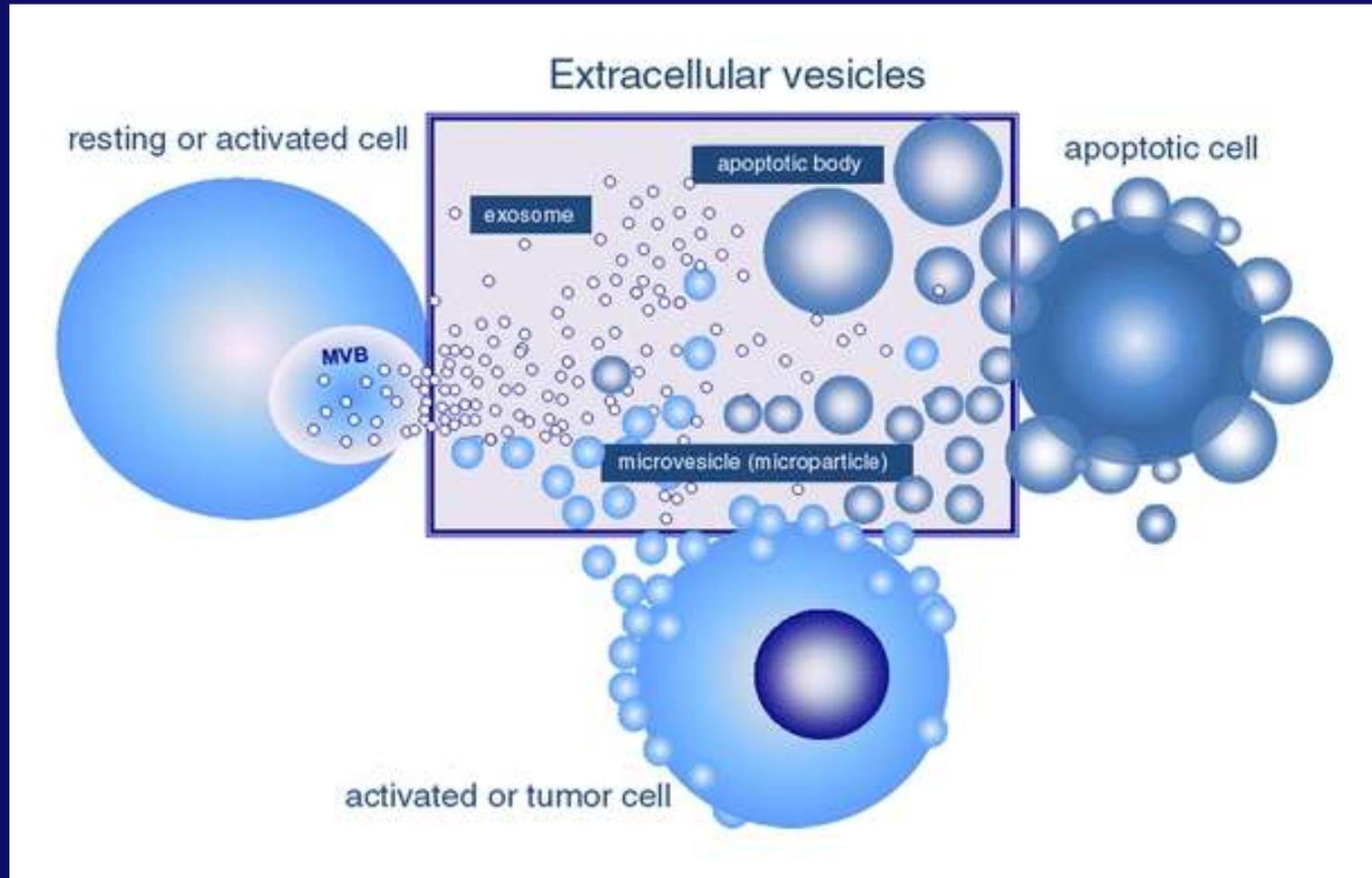
# Treatment With Ionis-FXI<sub>RX</sub> Produced a Dose-dependent and Sustained Decrease in Factor XI Activity





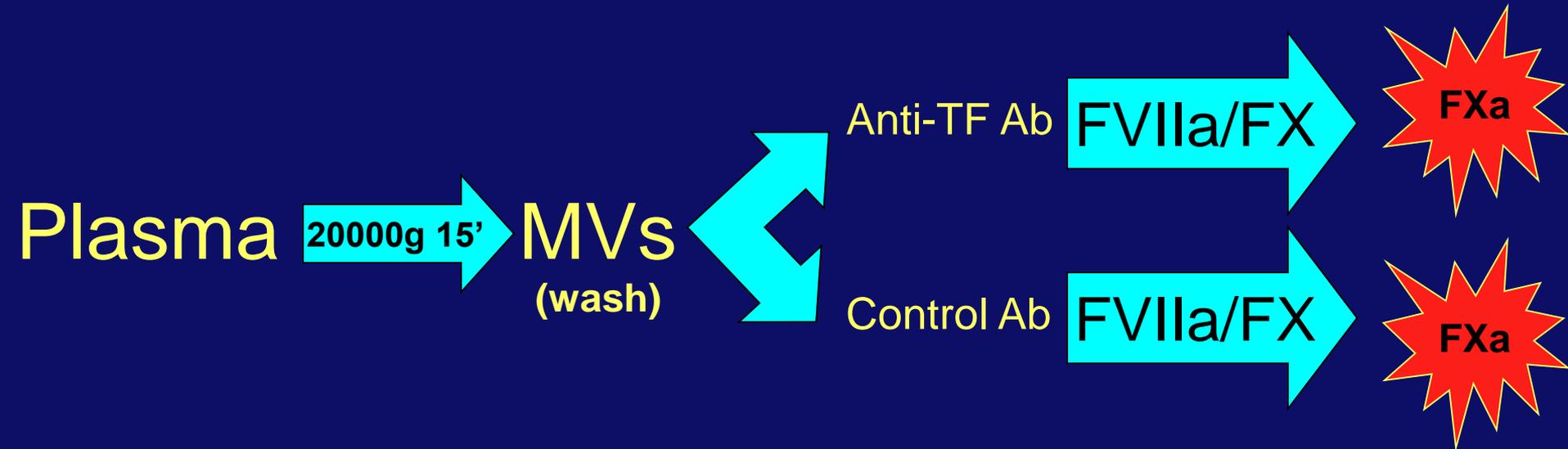
**WORLD THROMBOSIS DAY**  
**13 OCTOBER**

# Extracellular Vesicles



# Development of a New Assay to Measure Levels of MV TF Activity in Plasma

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**TF-dependent and TF-independent factor Xa activity**

Khorrana A et al JTH 2008, Wang J-G et al JTH 2009

Tesselaar M et al JTH 2007

**Healthy individuals do not have TF<sup>+</sup> MVs in the circulation**

**TF is expressed in a tissue-specific manner (e.g. high in heart and low in skeletal muscle)**

**Hypothesis:**

**TF provides additional hemostatic protection to select tissues**

**Analyze the hemostatic defects in low TF mice**

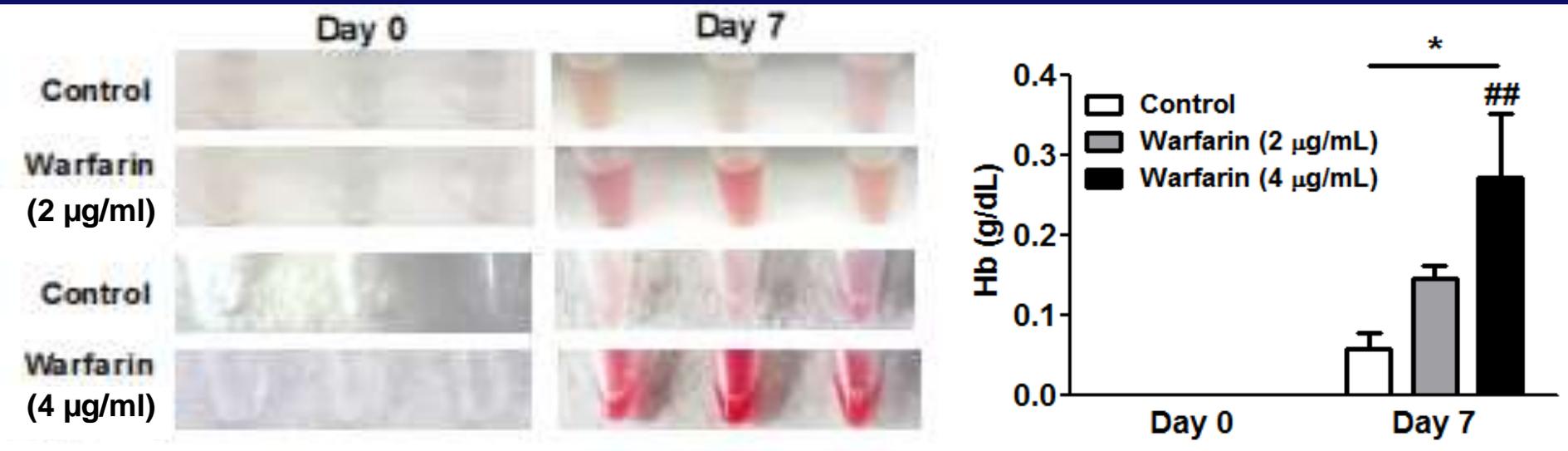
# Spontaneous Hemostatic Defects in Low TF Mice

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<b>Uterus</b>	Fatal hemorrhage post-partum	Erlich PNAS 1999
<b>Placenta</b>	Blood pools	Erlich PNAS 1999
<b>Heart</b>	Hemosiderin, fibrosis	Pawlinski PNAS 2002
<b>Lung</b>	Hemosiderin, fatal hemorrhages	Pedersen Blood 2005
<b>Testis</b>	Hemorrhage, calcification	Mackman JTH 2007
<b>Brain</b>	Occasional brain hemorrhage	Pawlinski T&H 2004

**Blood vessels in the heart and lung may be more prone to mechanical damage**

# Warfarin Increases Lung Hemorrhage After IAV Infection



# Conclusion

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**Anticoagulants may increase  
alveolar hemorrhage and  
morbidity in patients infected with  
influenza A**

# Cancer Cells and Procoagulant MVs

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## Tumor Shedding and Coagulation

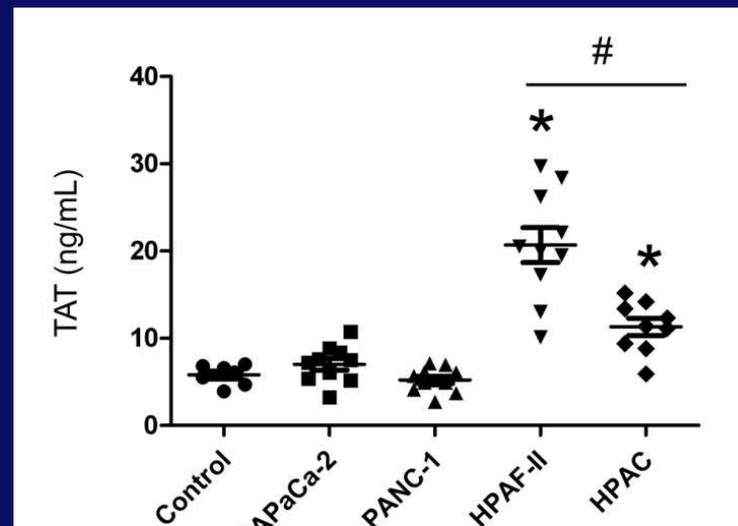
Dvorak HF et al Science 1981

Tumor cells shed plasma membrane vesicles when cultured in vitro. Shed vesicles carrying procoagulant activity may account for activation of the clotting system associated with malignancy

The procoagulant activity of tumor-derived MVs is due to TF

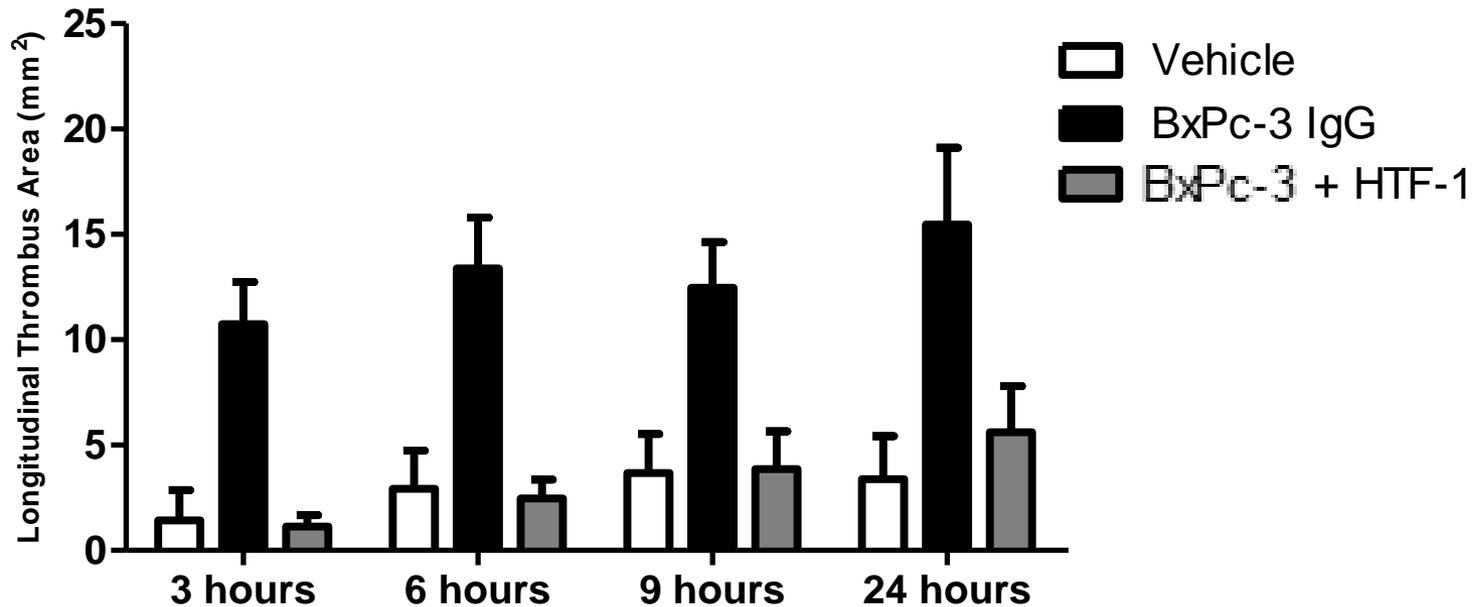
Dvorak et al Cancer Res 1983; Bastida et al Blood 1984; Yu and Rak JTH 2004

# Activation of Coagulation in Nude Mice containing Human Pancreatic Tumors



**An anti-human TF Ab reduces the activation of coagulation**

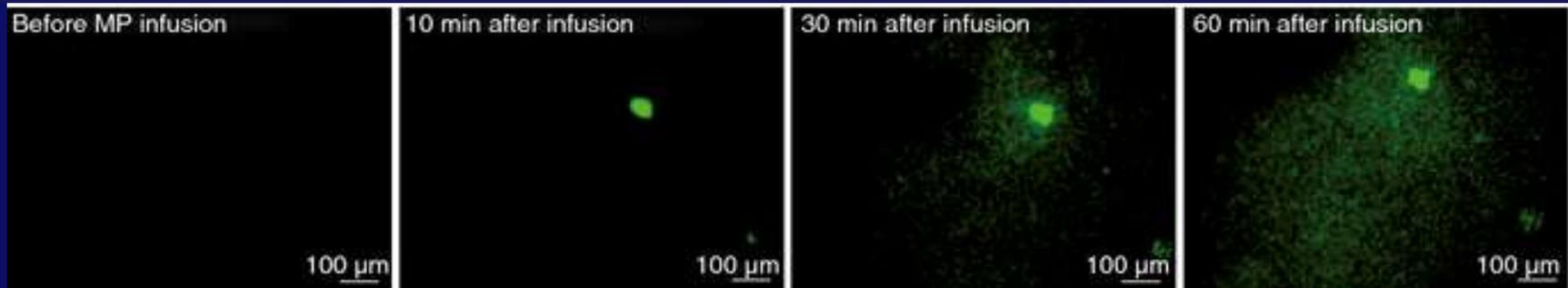
# Exogenous TF<sup>+</sup> Tumor MVs Enhance Thrombosis in the IVC Stenosis Model



**TF on tumor MV is required for the enhancement of thrombosis**

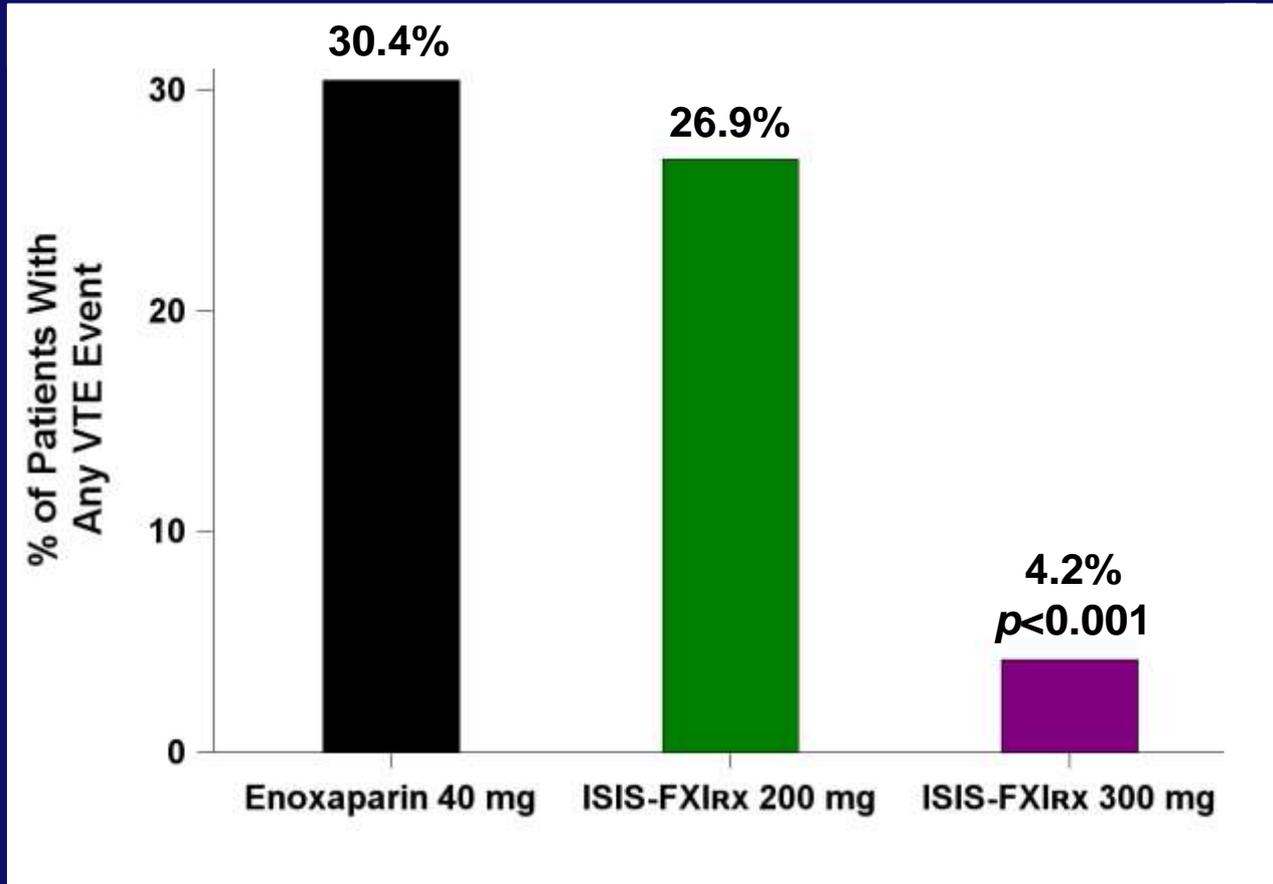
# Injected Tumor MVs Localize to the Site of Thrombosis in an IVC Stenosis Model

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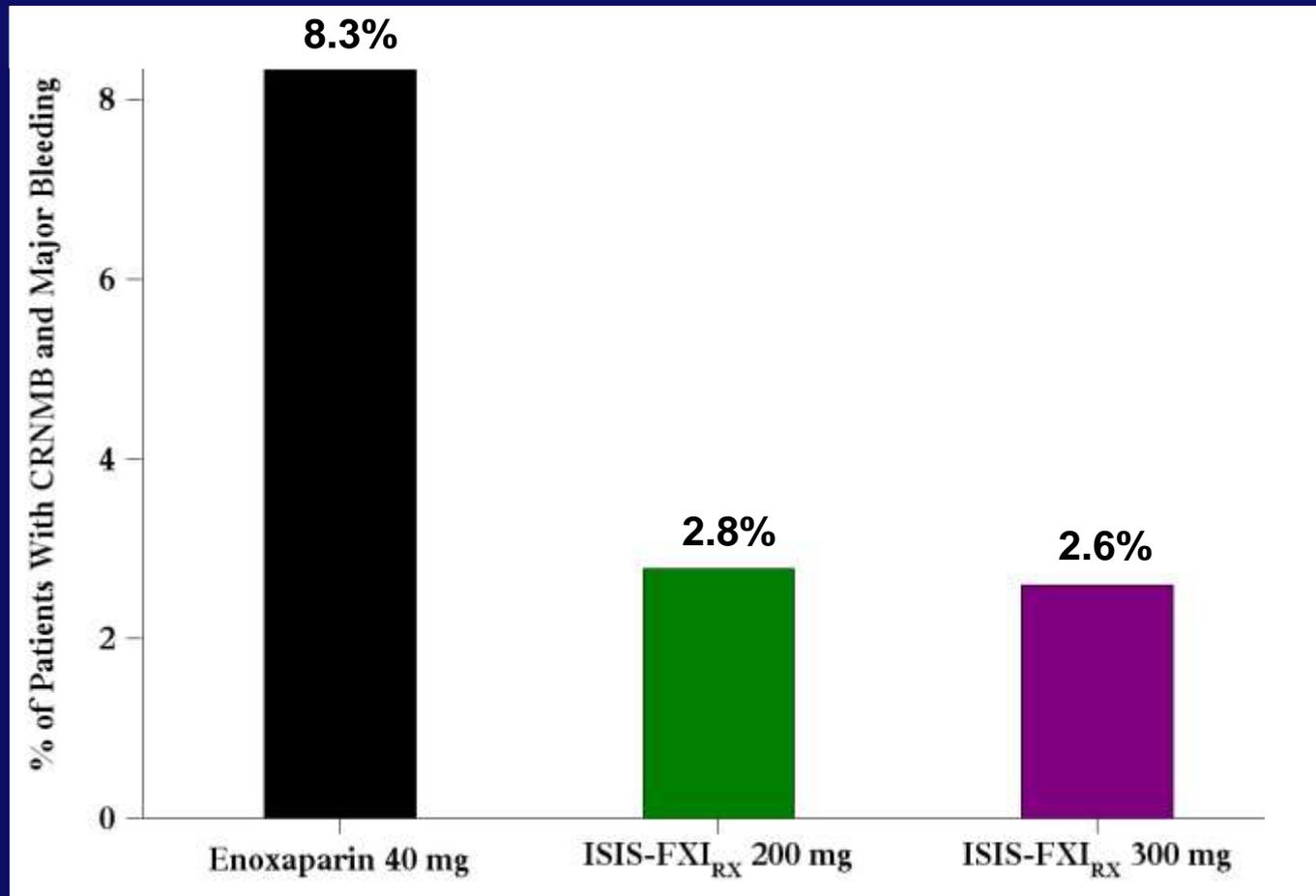
**Panc02 = mouse pancreatic cancer cell line**

# Primary Efficacy Outcome: Reduction in Incidence of VTE with Ionis-FXI<sub>Rx</sub> Compared with Enoxaparin



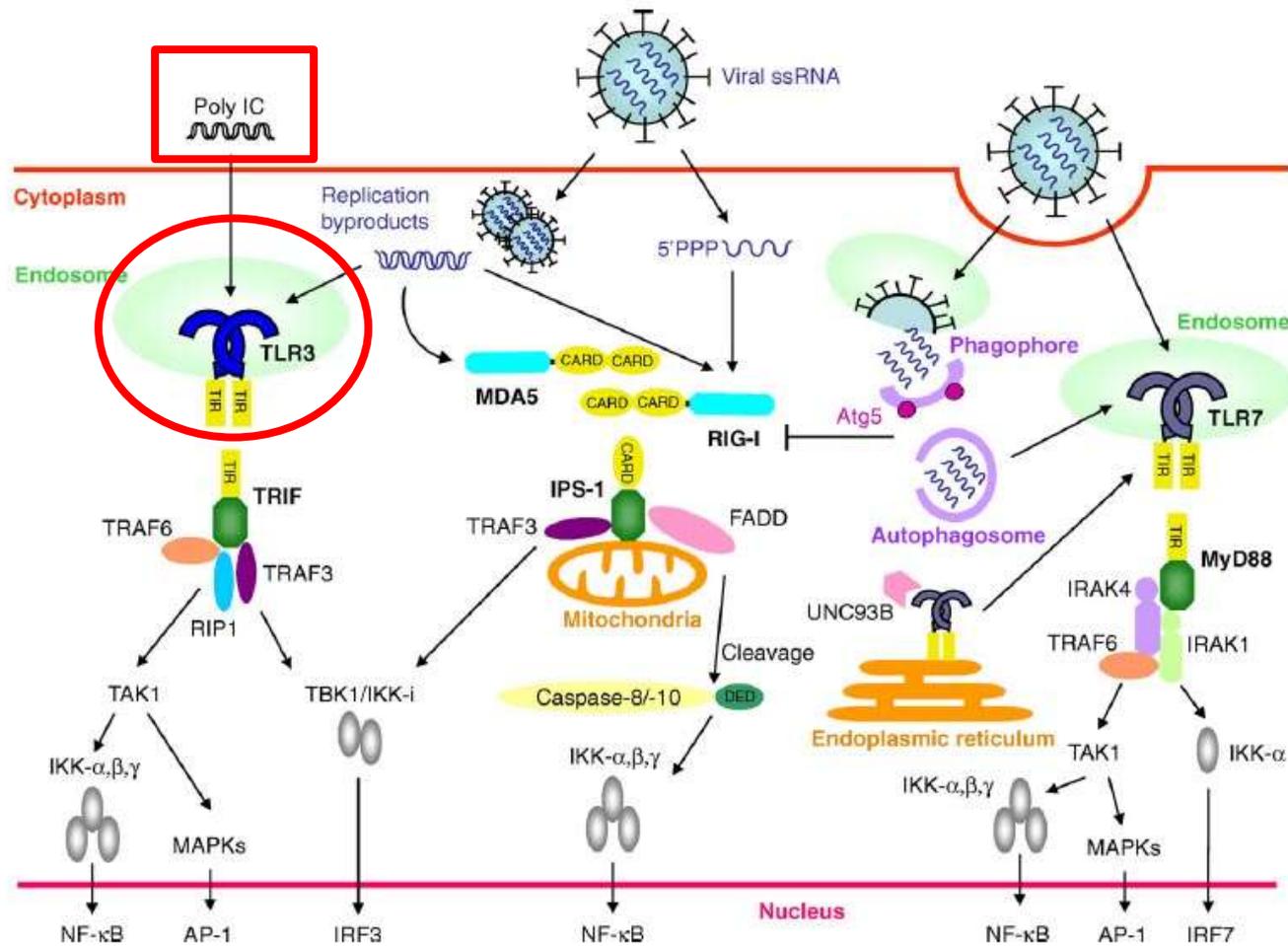
Buller H et al N Engl J Med 2015

# Clinically Relevant Bleeding Events with Ionis-FXI<sub>RX</sub> or Enoxaparin



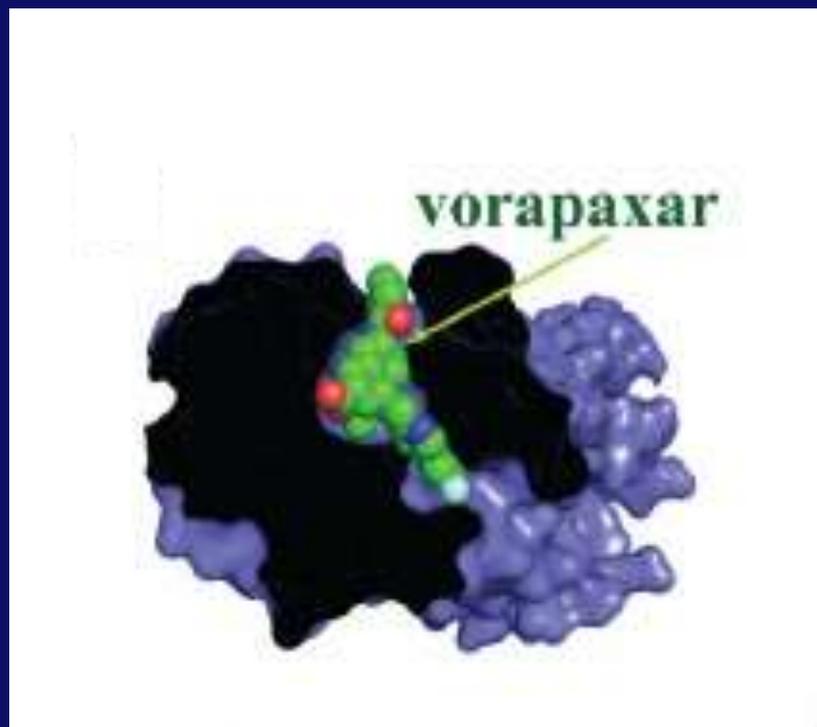
Buller H et al N Engl J Med 2015

# Detection of ssRNA Virus Infections by the Immune System



# Development of a New Antiplatelet Drug that Targets PAR-1

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**Vorapaxar (Zontivity) was approved by the FDA in 2014 for use with daily aspirin and/or clopidogrel**

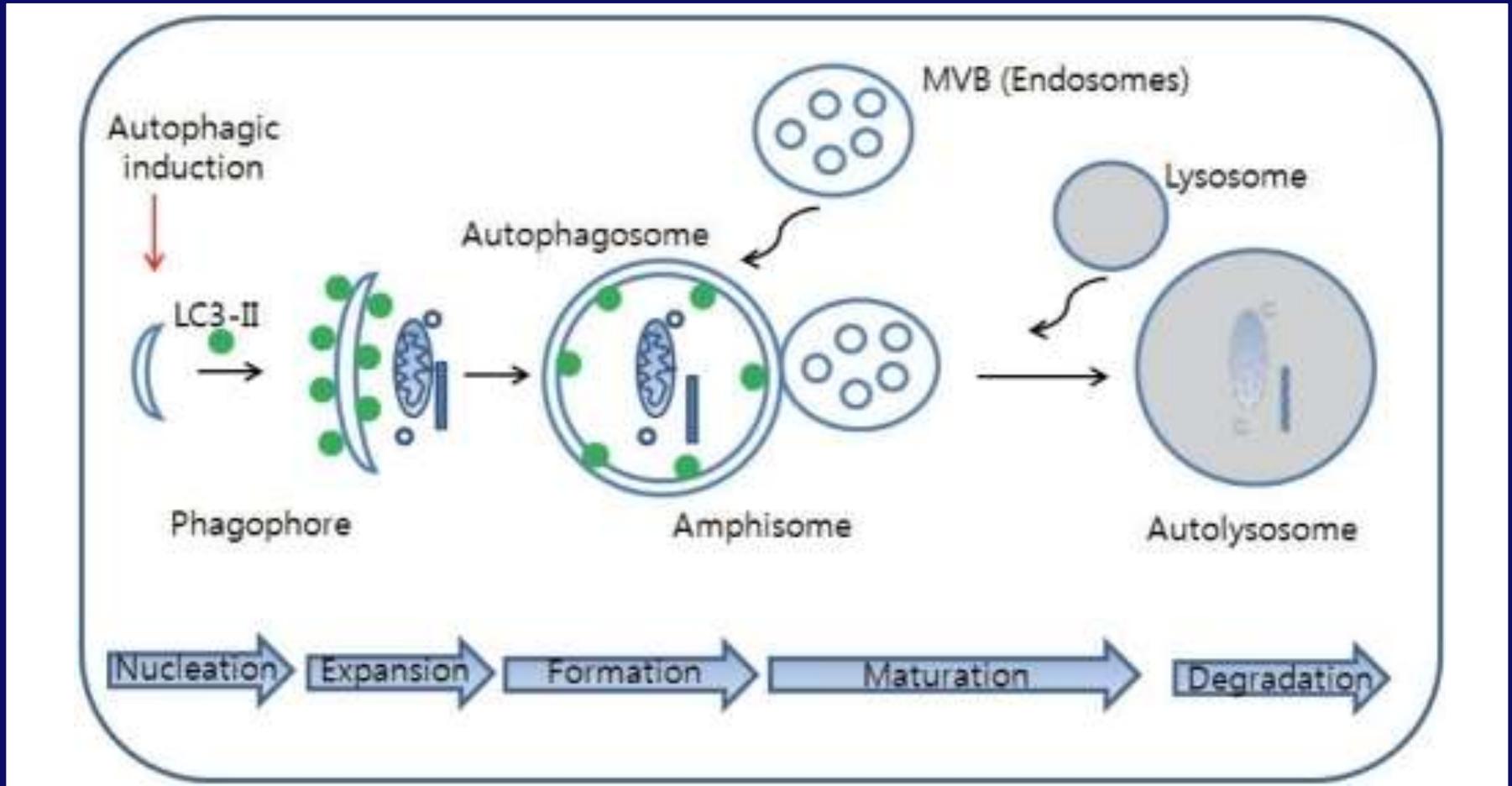
**Yang C et al Nature 2012**

# Virus Infection and Autophagy

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- Autophagy is usually used to destroy viruses.
- However, CVB3 is different because it utilizes autophagy to increase its replication.

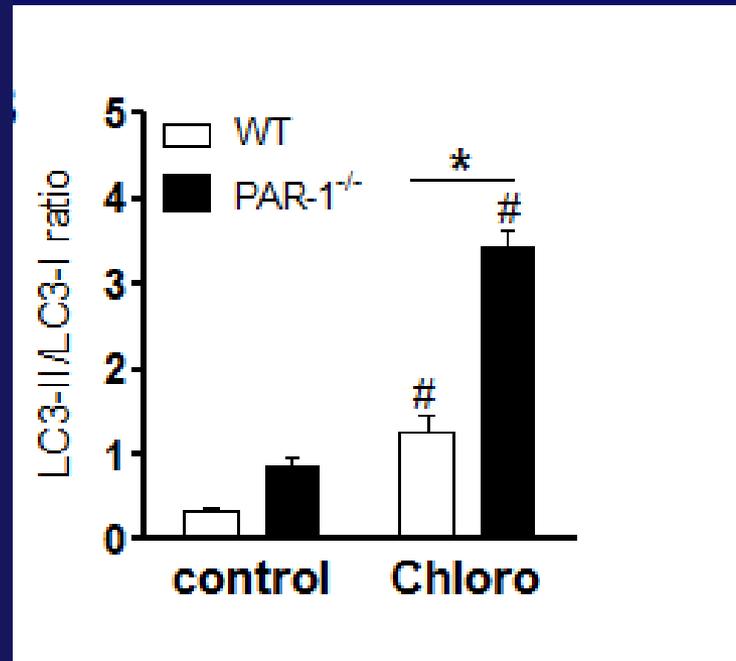
# Autophagy



Dr. Y. Ohsumi received the

**Hypothesis: PAR-1 reduces  
CVB3 replication by  
inhibiting autophagy**

# Effect of PAR-1 Deficiency on Autophagy



**Cloroquine inhibits autophagy and increases the accumulation of LC3-II and LC3-I**

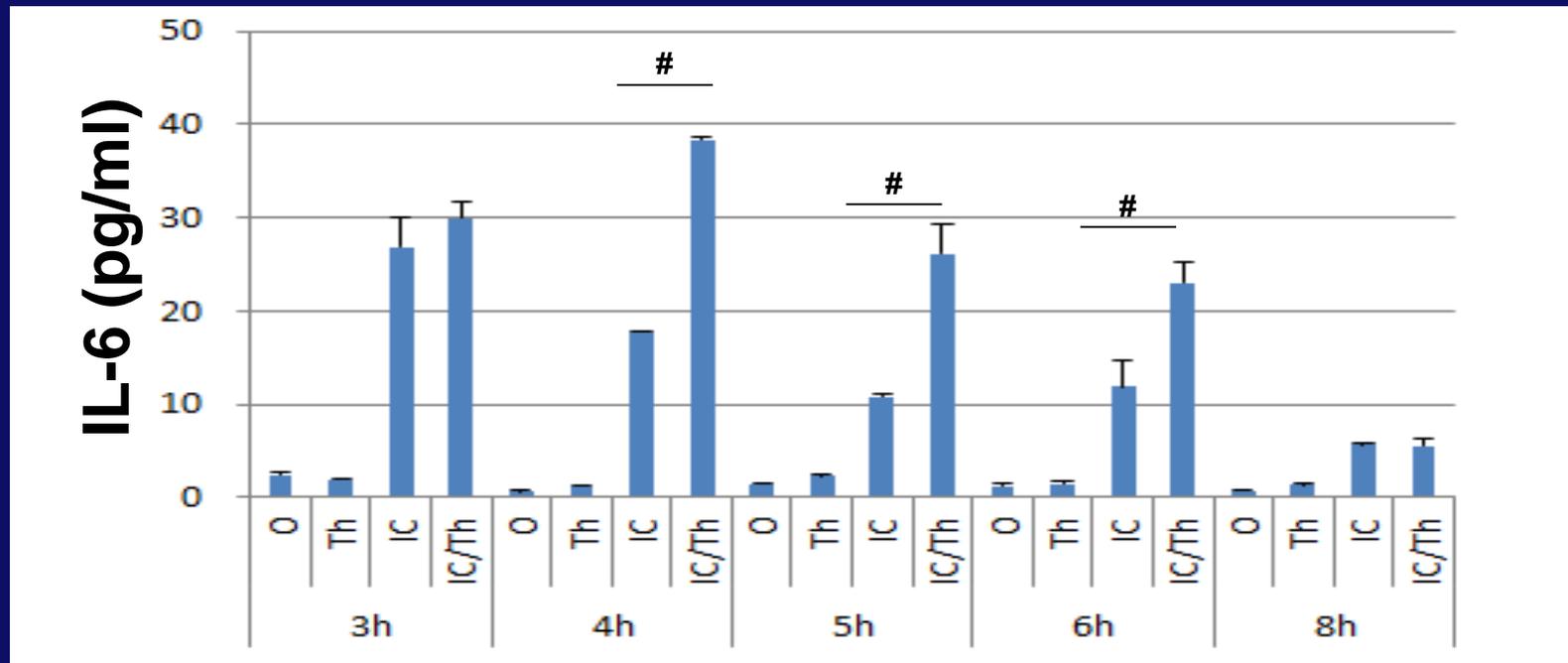
Vanja S et al. unpublished data

# Conclusion

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- **PAR-1 reduces CVB3 replication in CMs.**
- **Activation of autophagy increases CVB3 replication.**
- **PAR-1 deficiency is associated with increased autophagy.**

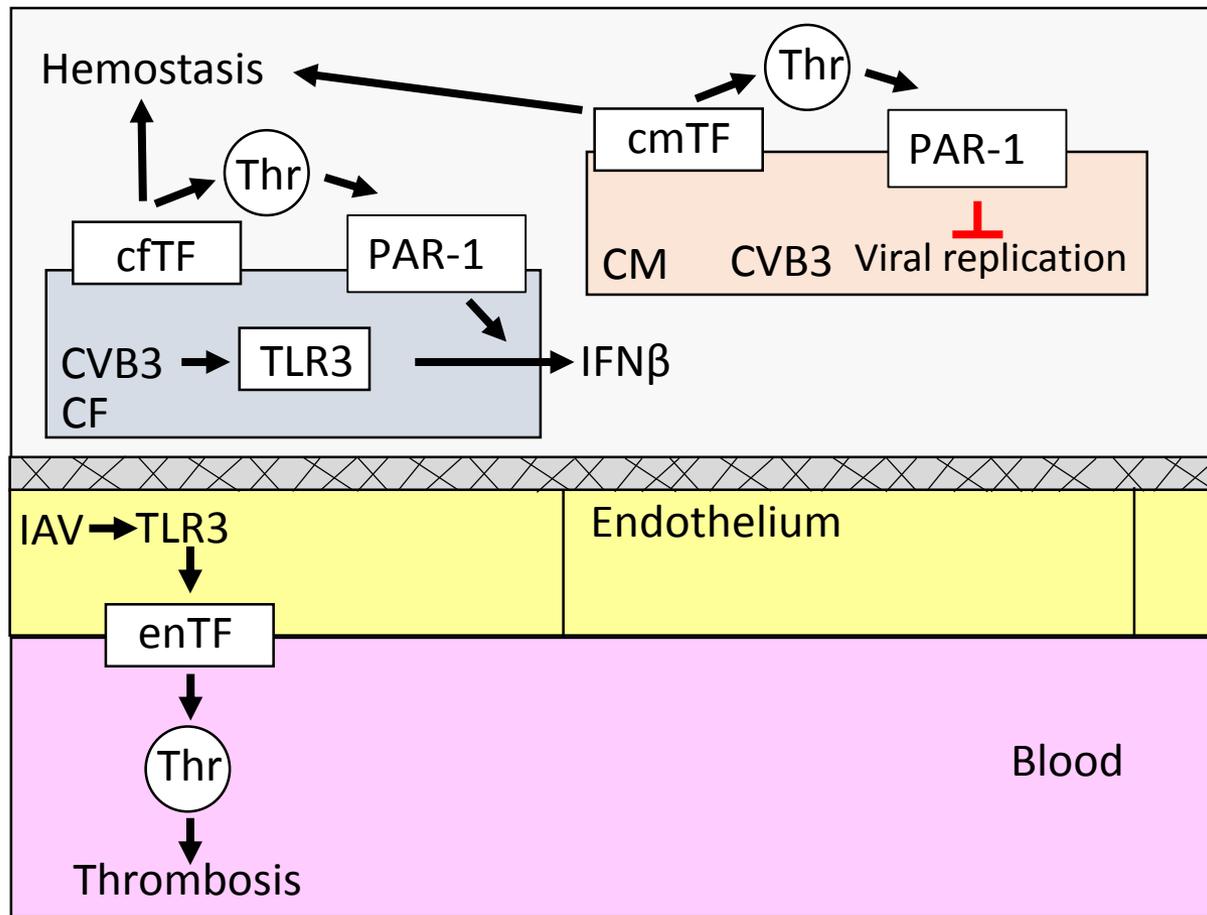
# PAR-1 Enhances Poly I:C Induction of IL-6 Expression in Primary Human Lung Epithelial Cells



No differences in poly I:C induction of IFN $\beta$ , CXCL10, OAS1, TNF $\alpha$  and MCP-1

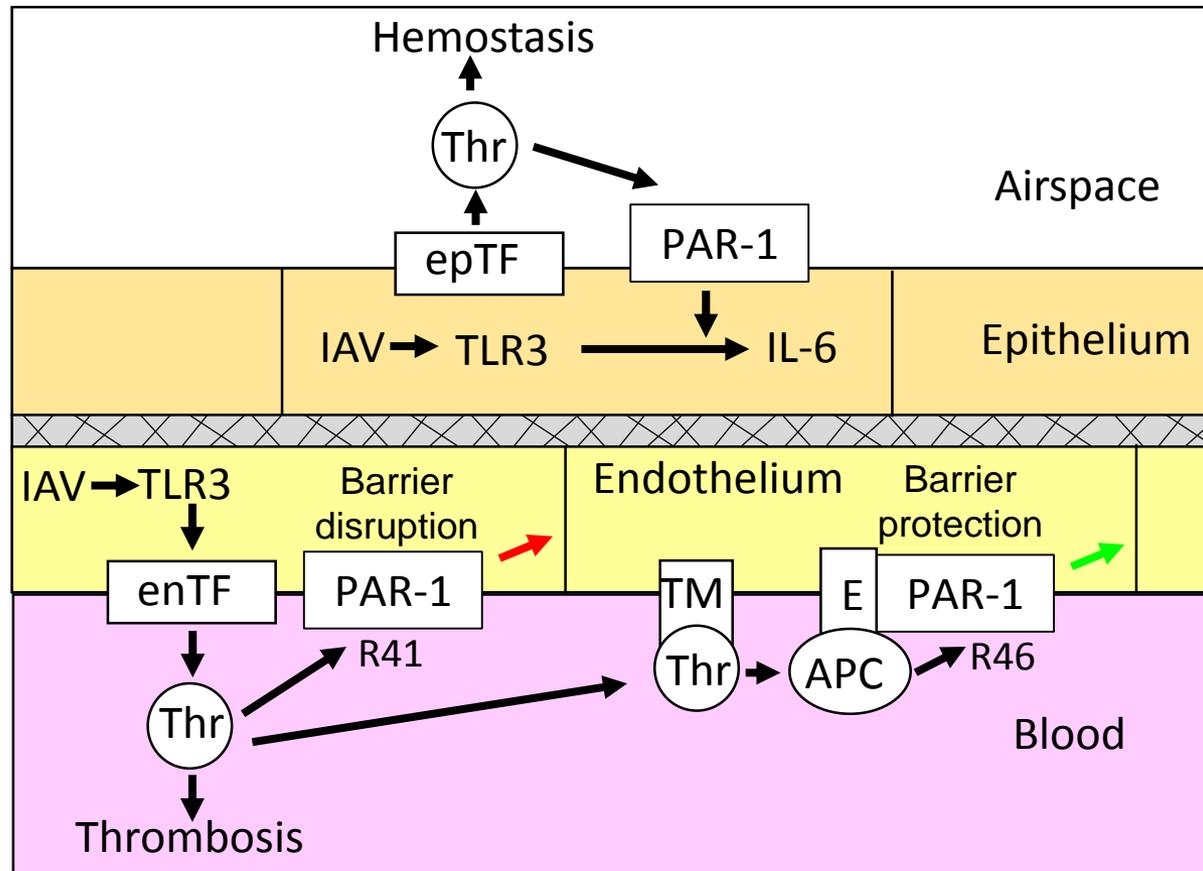
Tatsumi K et al. unpublished data

# Roles of the TF-Thrombin-PAR-1 Pathways in CVB3-induced Myocarditis



**Heart**

# Roles of the TF-Thrombin-PAR-1 Pathways in Influenza A Infection



**Lung**

# Viral Infections and the Clotting Cascade

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- Acute viral infections lead to strong activation of the clotting cascade and secondary hemorrhage (i.e. Ebola hemorrhagic fever).
- Inhibition of TF reduced death in a monkey model of Ebola.
- Chronic viral infections lead to a low level activation of the clotting cascade that increases cardiovascular disease (i.e. HIV).
- Two ongoing clinical trials are targeting FXa and PAR-1 in HIV patients.



# Coagulation and the Innate Immune Response to Pathogens

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- Formation of a clot reduces dissemination of pathogens
- Fibrin enhances activation of macrophages and cytokine expression by binding to Mac-1 (CD11b/CD18)
- Thrombin activation of PAR-1 enhances IFN- $\beta$  and CXCL10 expression

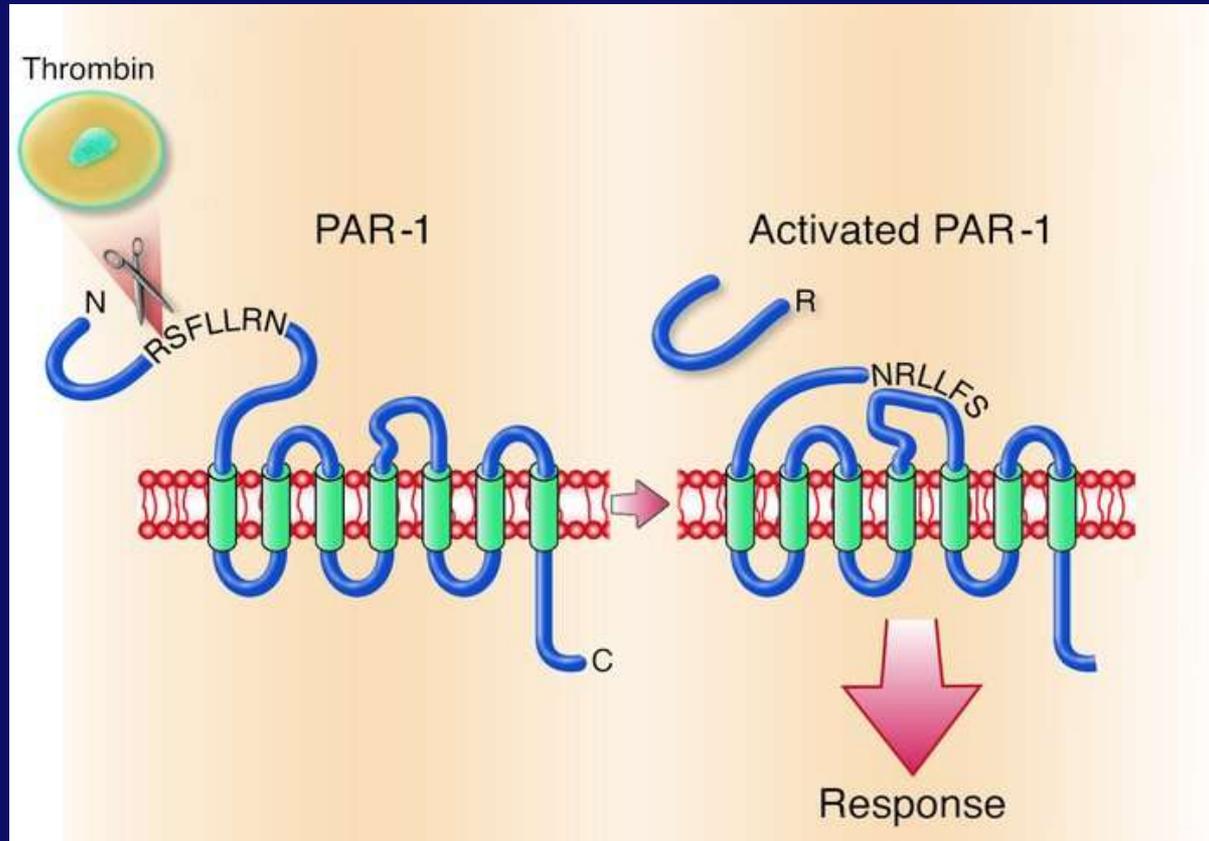
**General Hypothesis:  
The thrombin-PAR-1 pathway  
contributes to innate antiviral  
immunity**

# The Protease-Activated Receptor Family

	PAR-1	PAR-2	PAR-3	PAR-4
Primary activating proteases (EC <sub>50</sub> )	Thrombin (50 pM)	Trypsin (1 nM) Tryptase (1 nM)	Thrombin (0.2 nM)	Thrombin (5 nM) Trypsin (5 nM)
Secondary activating proteases	Granzyme A FXa Trypsin Plasmin MMP-1	FVIIa FXa MT-SP1		Cathepsin G
PAR-APs	SFLLRN TFLLRN	SLIGKV SLIGIRL SFLLRN		GYPGKF AYPGKF

Adapted from Major et al., 2003 *Arterioscler. Thromb. Vasc. Biol.*

# Thrombin Activation of PAR-1

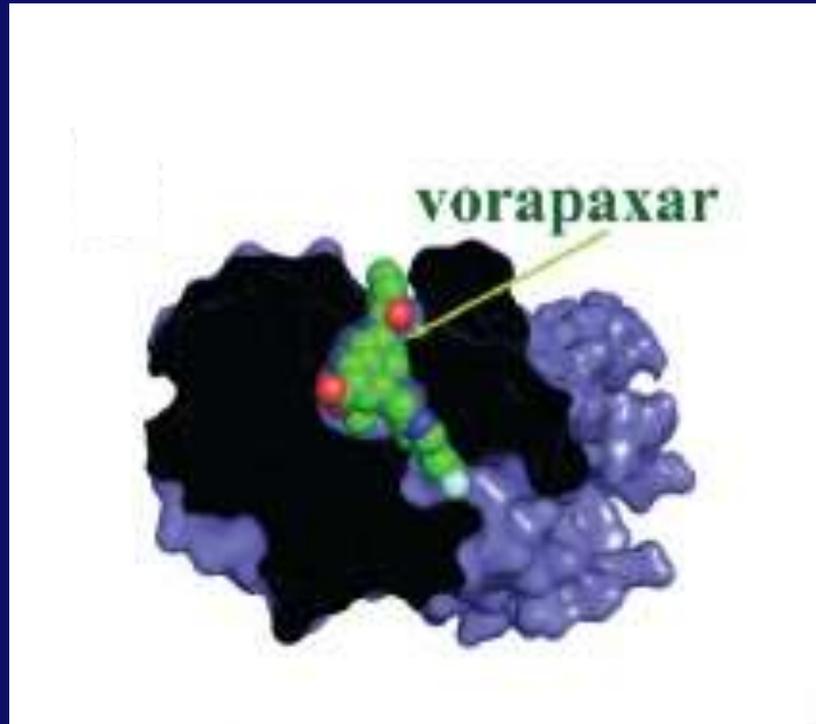


**PAR-1 is a unique GPCR because it is activated by proteolytic cleavage. It is the major thrombin receptor on human platelets.**

**Camerer and Coughlin JCI 2003**

# Development of a New Antiplatelet Drug that Targets PAR-1

---

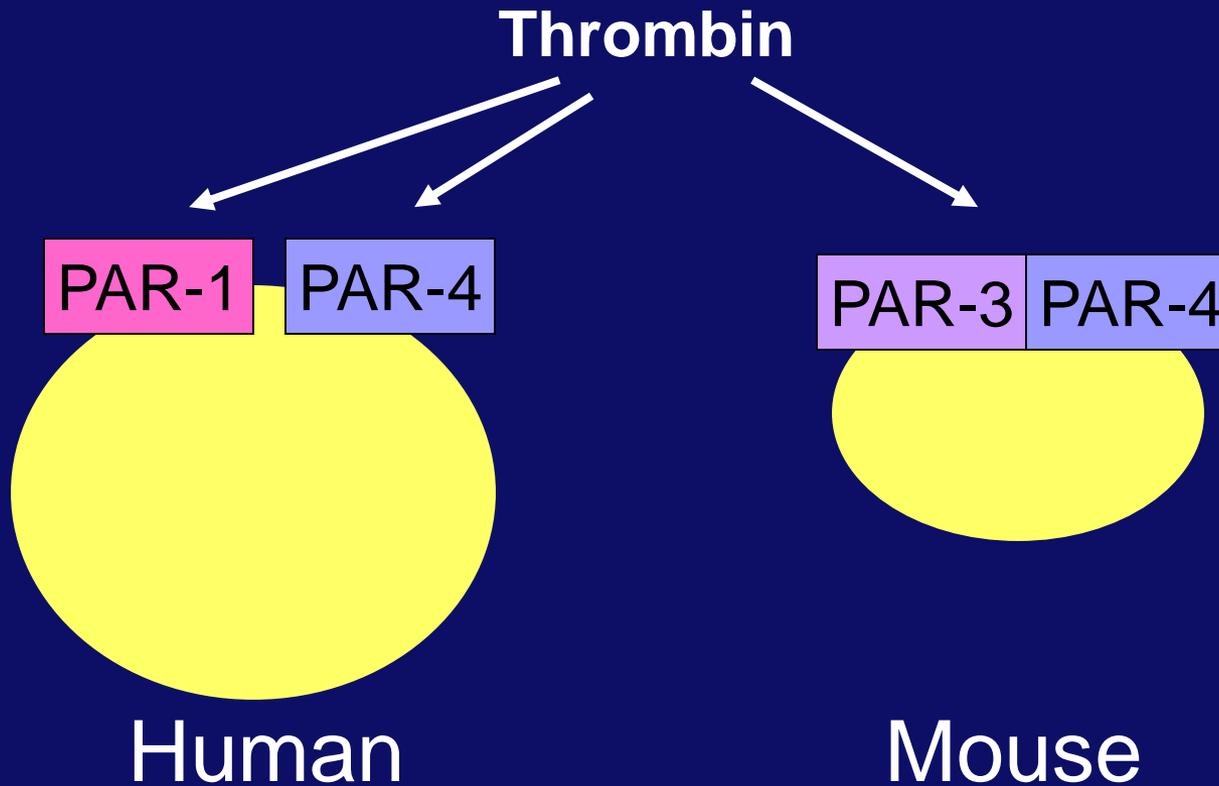


**Vorapaxar (Zontivity) was approved by the FDA in 2014 for use with daily aspirin and/or clopidogrel**

**Yang C et al Nature 2012**

# PAR Expression on Human and Mouse Platelets

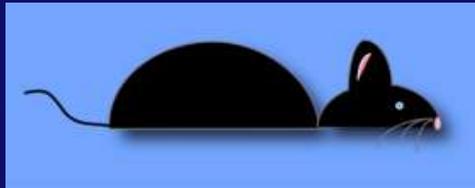
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Any phenotype in PAR-1<sup>-/-</sup> mice cannot be due to a defect in thrombin activation of platelets

# Mouse Models

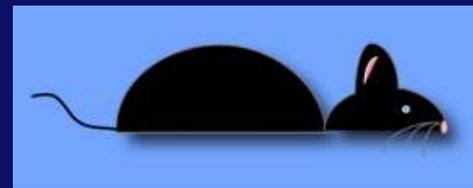
---



## PAR-1<sup>-/-</sup> mice

Cell type specific  
deletion of PAR-1  
(crossed with Cre<sup>+</sup>)

Darrow A et al  
Thromb & Hemost  
1996



## PAR-1<sup>fl/fl</sup> mice

Cell type specific  
deletion of PAR-1  
(crossed with Cre<sup>+</sup>)

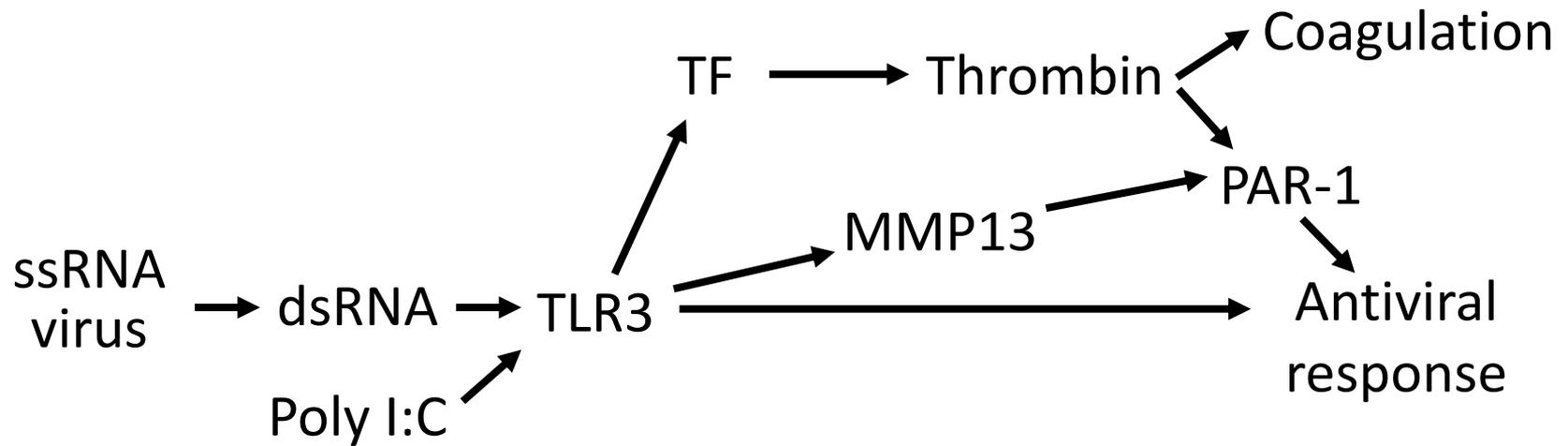
Palumbo J  
unpublished data

# Viral Models

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- **Coxsackievirus B3**  
(liver, spleen, pancreas and heart)
- **Influenza A virus**  
(lung)

# Role of the Thrombin-PAR-1 Pathway in Innate Antiviral Immunity



# Outline

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- **Poly I:C studies**
- **Coxsackievirus B3**
- **Influenza A virus**

**Poly I:C as a mimetic  
of an antiviral  
response**

# Role of PAR-1 in Poly I:C induction of IFN $\beta$ , CXCL10 and CXCL1 in Macrophages and Splenocytes

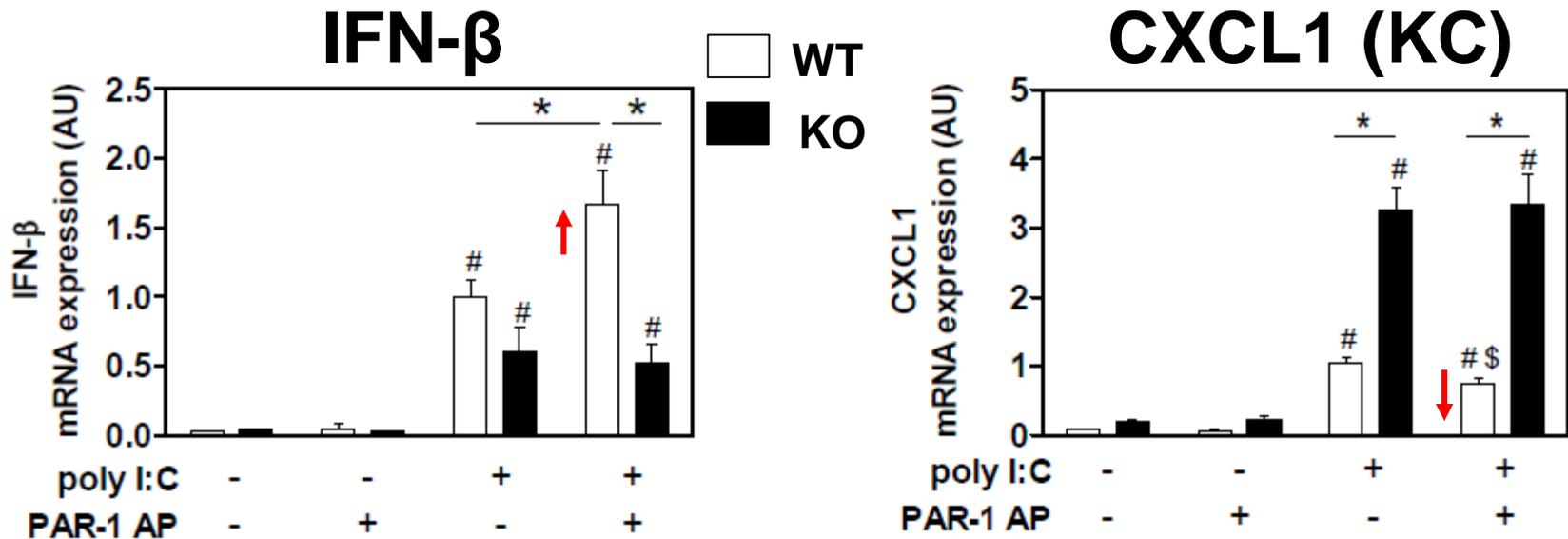
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Poly I:C was used to stimulate TLR3

PAR-1 agonist peptide (AP) was used to stimulate PAR-1

Use mouse bone marrow-derived macrophages (BMDM) and splenocytes

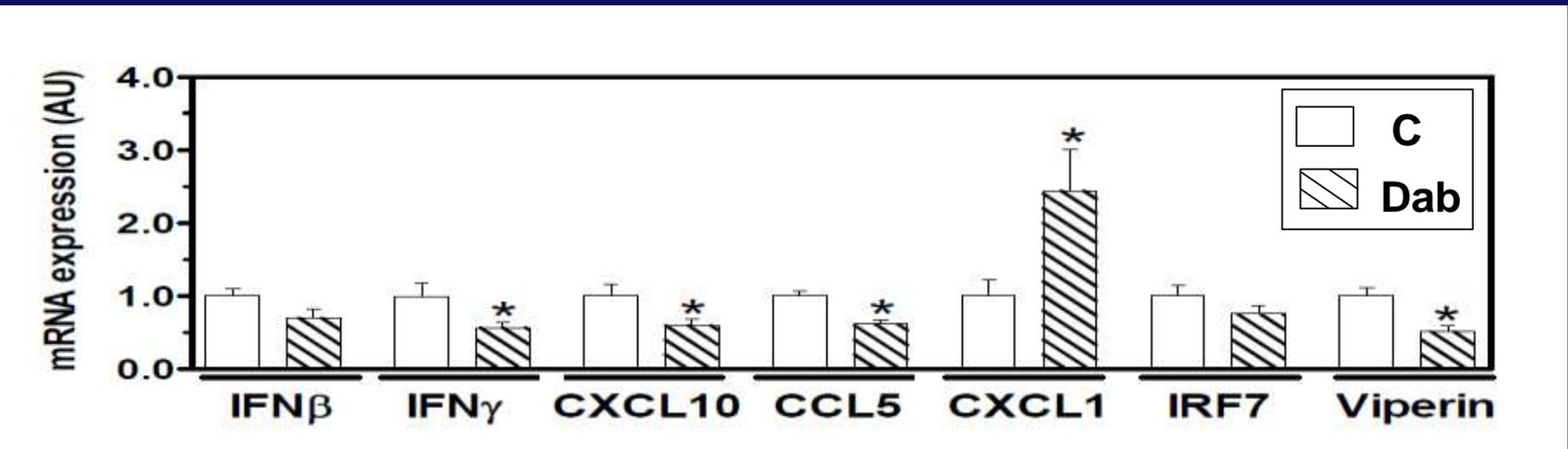
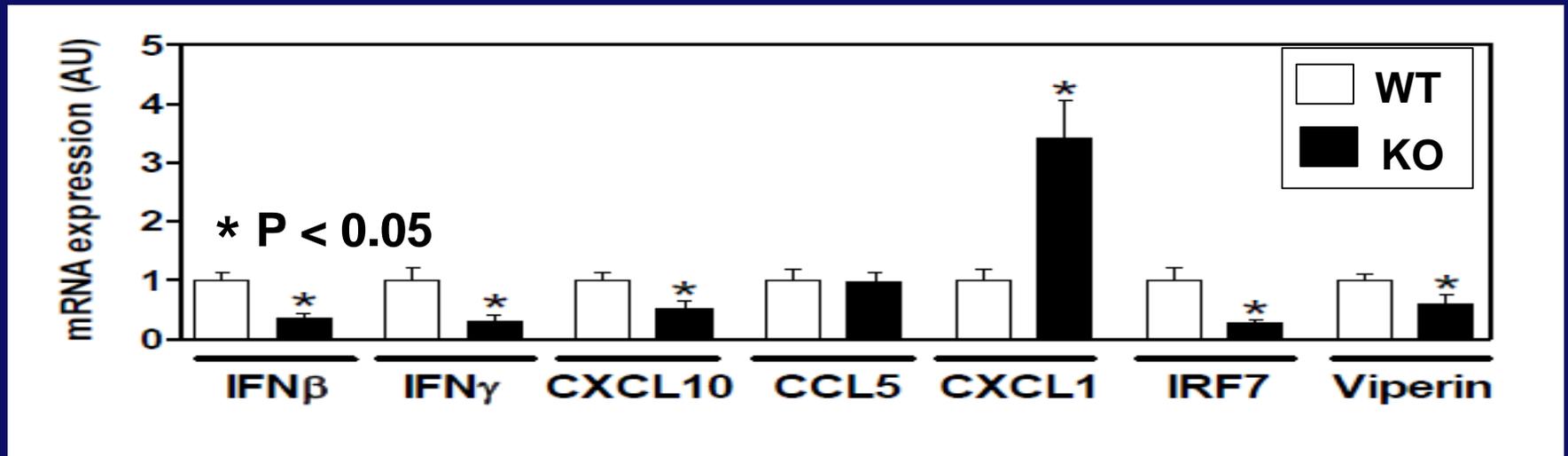
# Role of PAR-1 in Poly I:C Induction of Antiviral Genes in Splenocytes



Antoniak S et al unpublished data

**Effect of PAR-1 deficiency  
and thrombin inhibition in  
WT mice on poly I:C  
induction of the innate  
immune response in mice**

# Effect of PAR-1 Deficiency and Thrombin Inhibition in WT Mice on Poly I:C Induction of Antiviral Genes in the Spleen



Antoniak S et al unpublished data

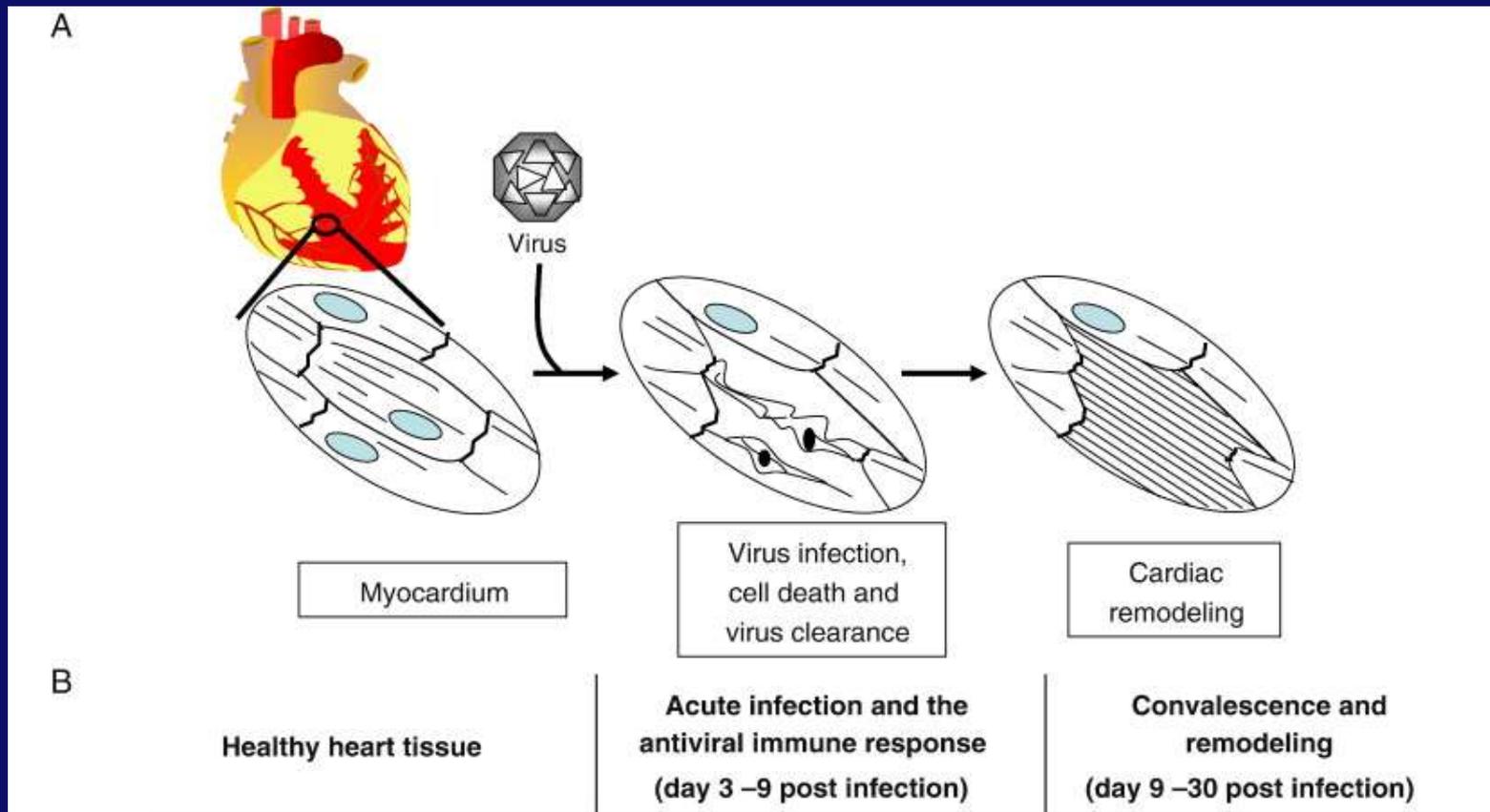
# Conclusion

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**PAR-1 deficient mice and WT mice treated with dabigatran have decreased levels of various proteins, such as CXCL10, and increased levels of CXCL1 compared with controls after poly I:C injection**

# Viral Myocarditis

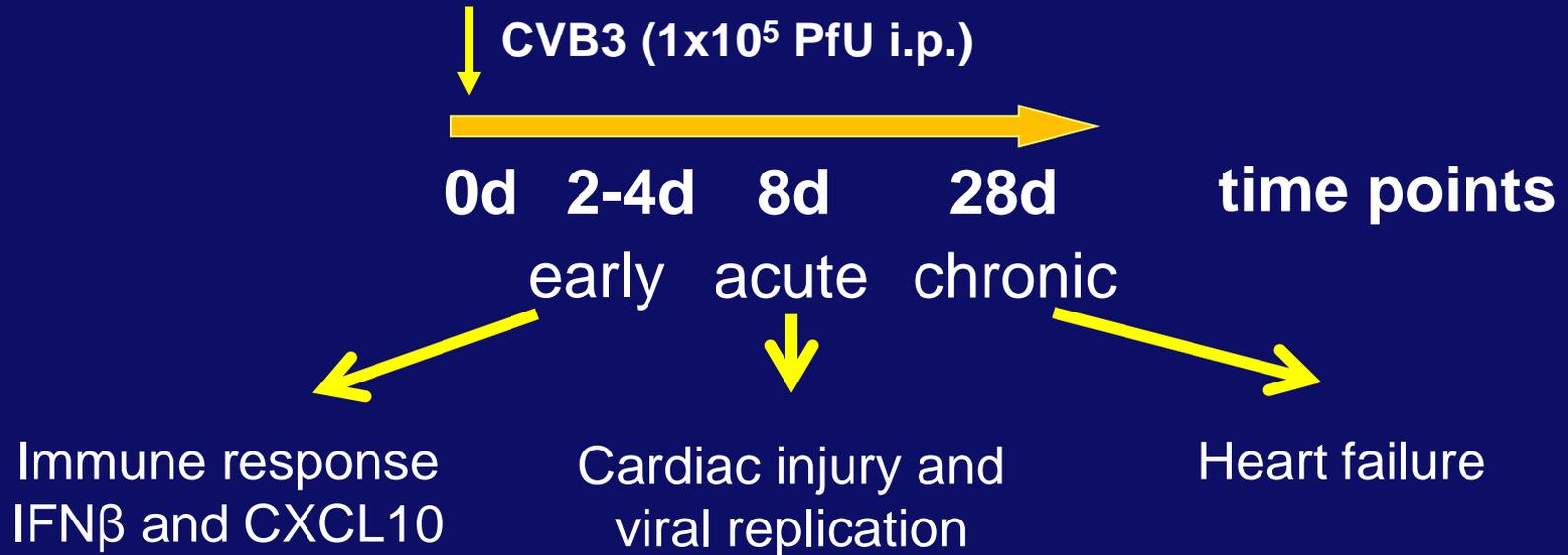
Accounts for up to 20% of sudden cardiac death among young people (<40 years of age)



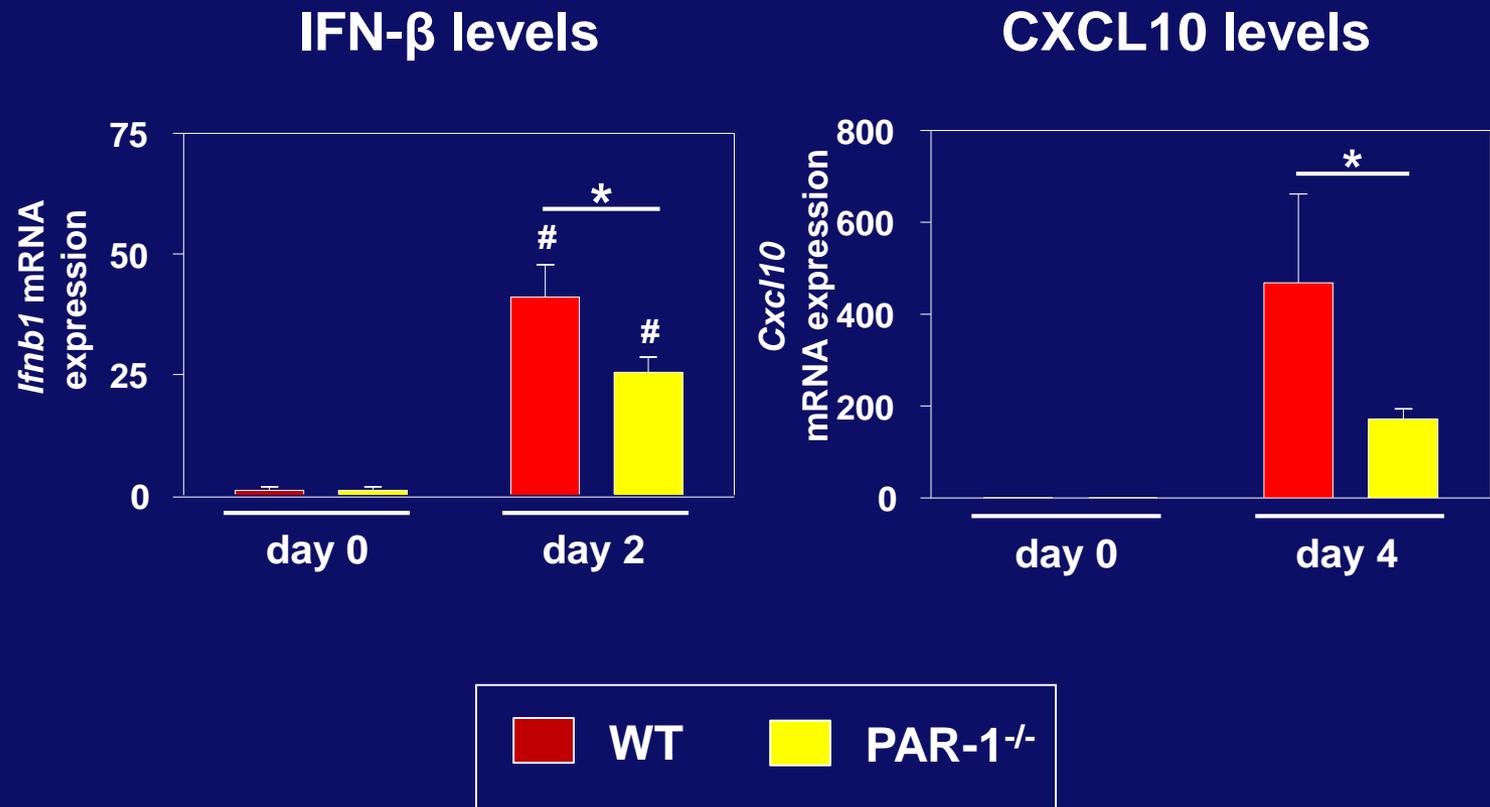
# Mouse Model of Coxsackievirus B3 Myocarditis

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- Coxsackievirus B3 (CVB3) = cardiotropic ssRNA(+) virus



# PAR-1 Deficiency is Associated with Reduced IFN- $\beta$ and CXCL10 expression After CVB3 Infection



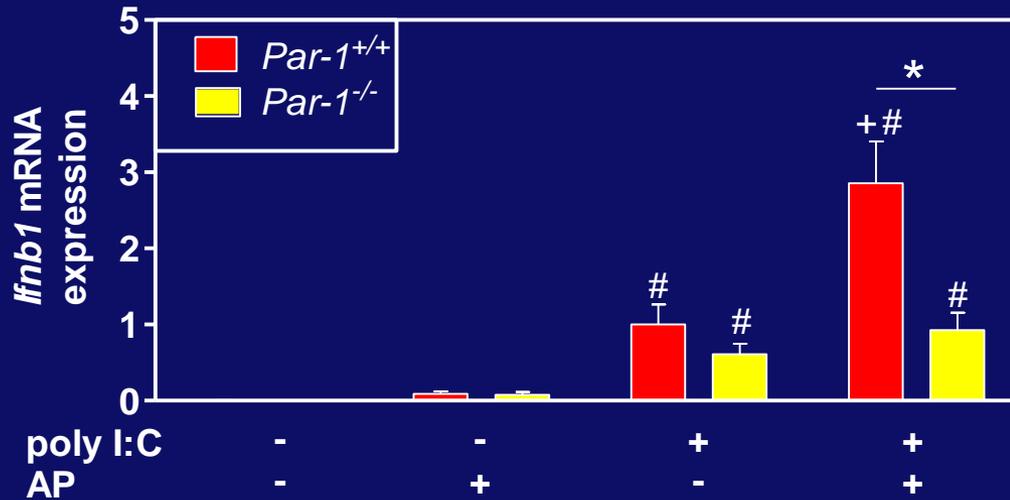
\*P<0.05; #P<0.05 vs. day 0  
Two-Way ANOVA

Antoniak et al, JCI 2013

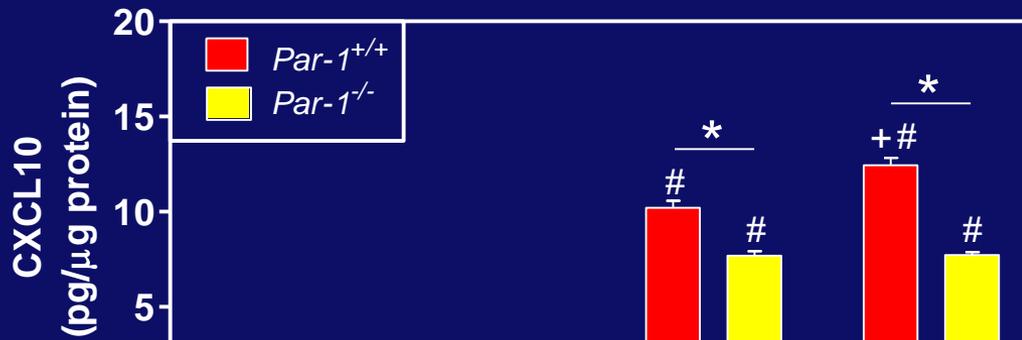
**BMT studies indicated that PAR-1  
on both hematopoietic and non-  
hematopoietic cells contributed  
to the protection from CVB3  
infection**

# PAR-1 Activation Enhances TLR3-dependent IFN- $\beta$ and CXCL10 Expression in Cardiac Fibroblasts

IFN- $\beta$   
mRNA



CXCL10  
Protein



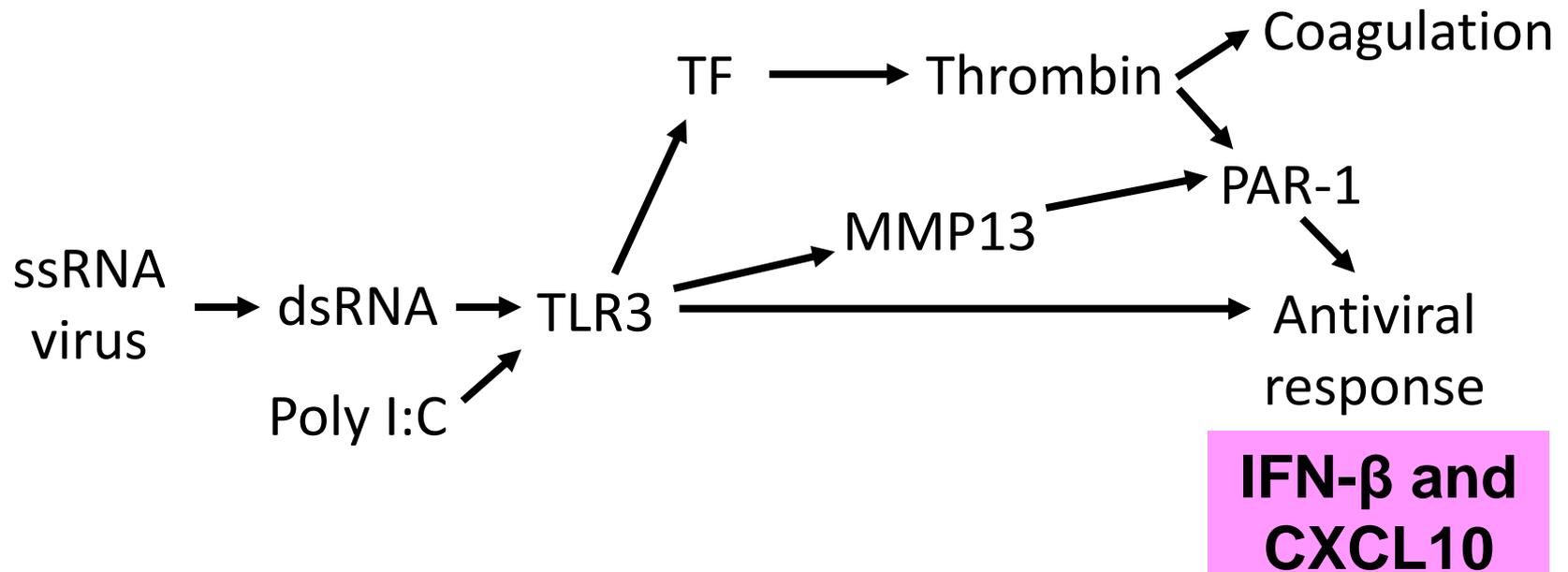
**Poly I:C induces MMP13 expression that activates PAR-1**

# Ongoing Studies: Effect of Deletion of PAR-1 in Myeloid Cells, Cardiac Fibroblasts or Cardiac Myocytes on CVB3-induced Myocarditis

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- PAR-1<sup>fl/fl</sup>,LysM-Cre (myeloid cells)
- PAR-1<sup>fl/fl</sup>,Mlc2v-Cre (CMs)
- PAR-1<sup>fl/fl</sup>,TCF21-iCre (CFs)

# Role of the Thrombin-PAR-1 Pathway in Innate Antiviral Immunity



# Influenza A Infection

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- Influenza A (IAV) is a respiratory pathogen
- IAV causes acute infection of the upper respiratory tract
- Severe IAV infection can cause infection of the lower respiratory tract, resulting in viral pneumonia
- Damage to the barrier formed by epithelial and endothelial cells in the pulmonary alveolus leads to respiratory dysfunction

# Mouse Model of Influenza A Infection (IAV)

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- Influenza A is a ssRNA virus
- Influenza A/Puerto Rico/8/1934 H1N1
- Mouse adapted and highly pathogenic
- Infected intranasally

Silvio Antoniak and Kohei Tatsumi

# Mouse Model of Influenza A Infection

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H1N1 (0.02  
HAU, 50 $\mu$ L i.n.)



0d

3 and 7d

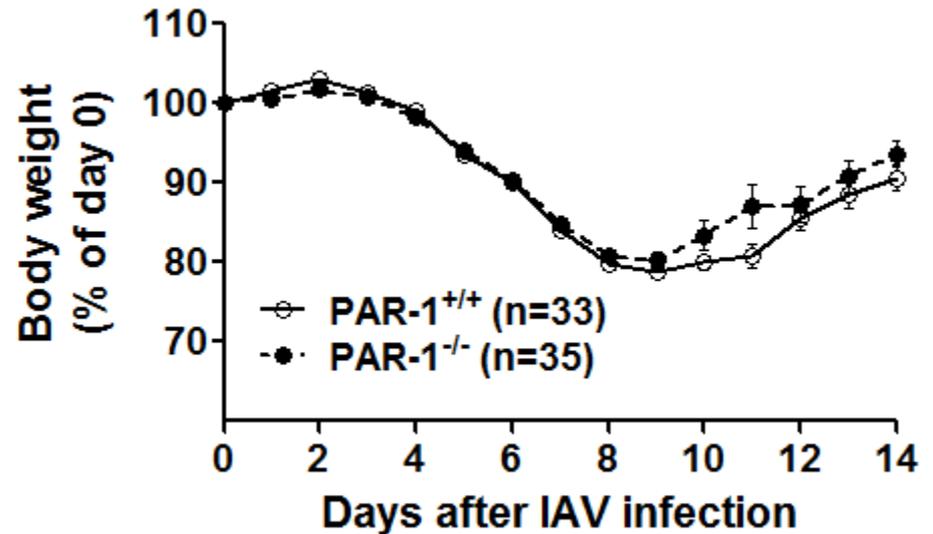
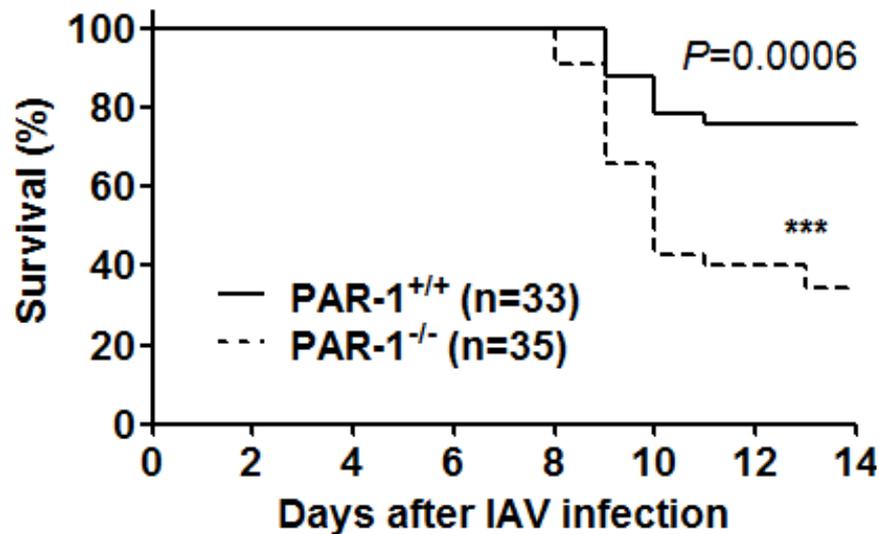
14d

time points

BALF and  
histology

Survival and  
weight loss

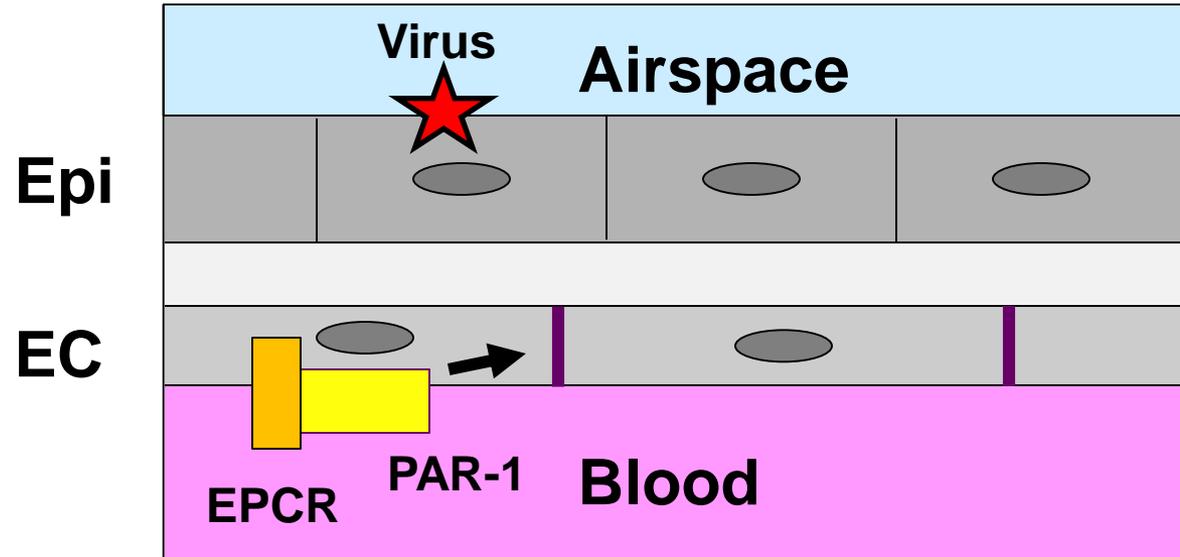
# PAR-1 Deficiency is Associated with Increased Mortality after Influenza A Infection



Tatsumi K et al. unpublished data

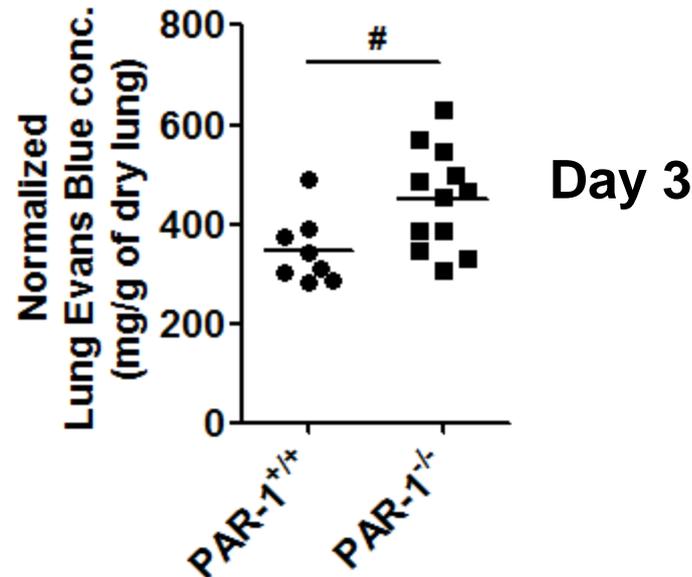
# Hypothesis 1:

APC/EPCR/PAR-1 maintains barrier function during IAV infection



# PAR-1 Deficient Mice have Increased Endothelial Permeability after IAV Infection

## Evans Blue Leakage

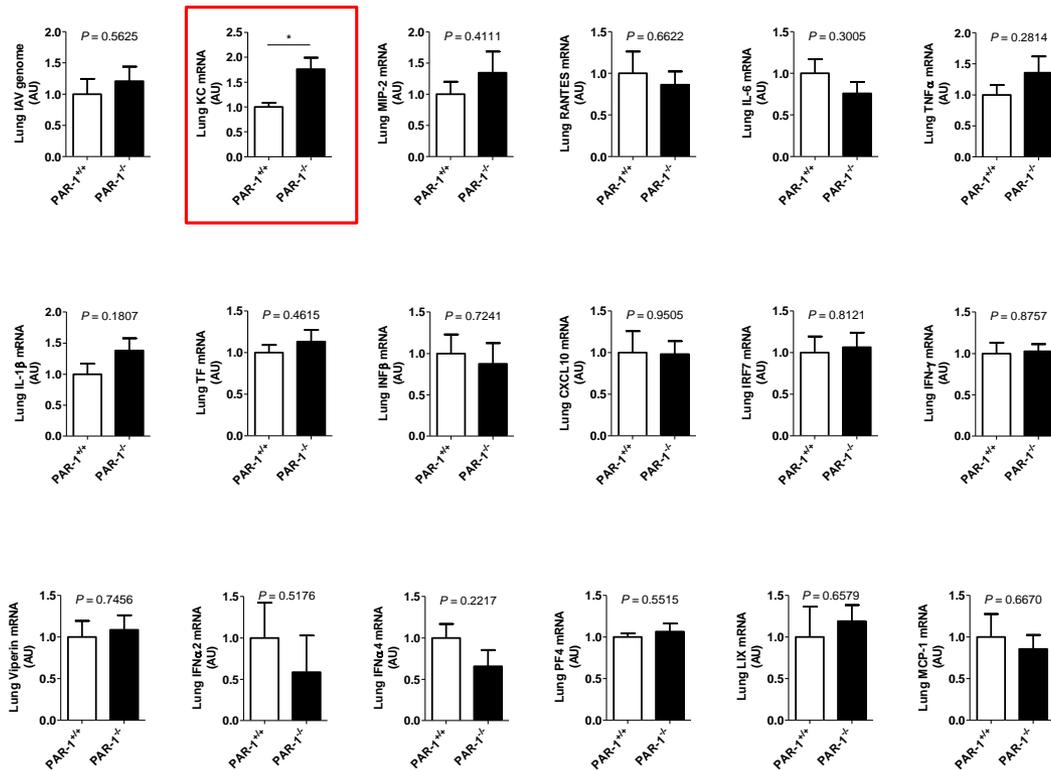


A decrease in barrier protection may contribute to the increased mortality in PAR-1<sup>-/-</sup> mice

## **Hypothesis 2:**

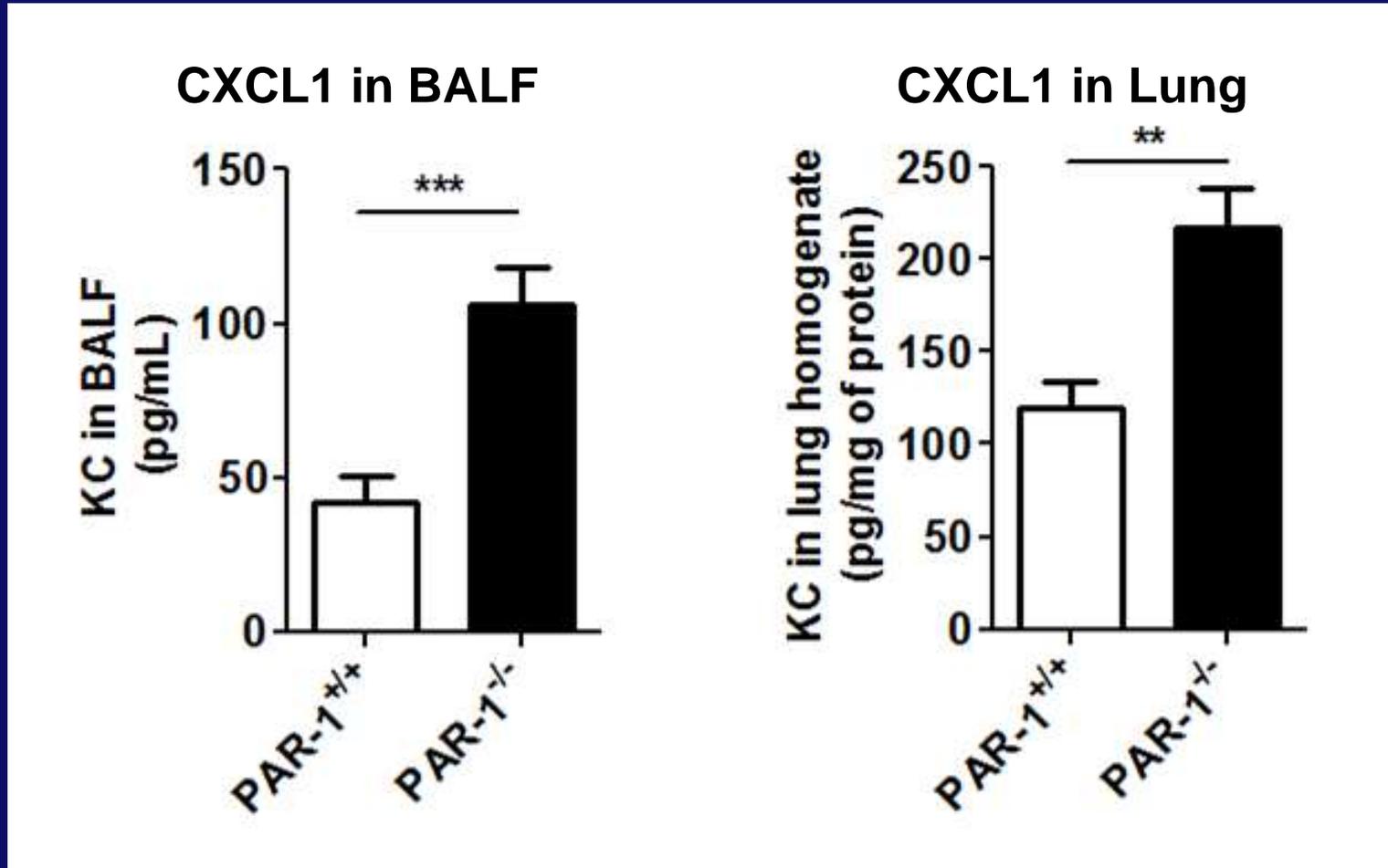
**PAR-1 deficiency is associated with a  
dysregulated inflammatory response after  
IAV infection**

# Effect of PAR-1 Deficiency on the Expression of Inflammatory Mediators in the Lung 3 days after IAV Infection



Tatsumi K et al. unpublished data

# PAR-1 Deficient Mice have Increased CXCL1 (KC) Expression in the Lungs 3 Days After IAV Infection



Tatsumi K et al. unpublished data

# PAR-1 Deficient Mice Have Increased Numbers of Neutrophils in the BALF 3 Days after IAV Infection

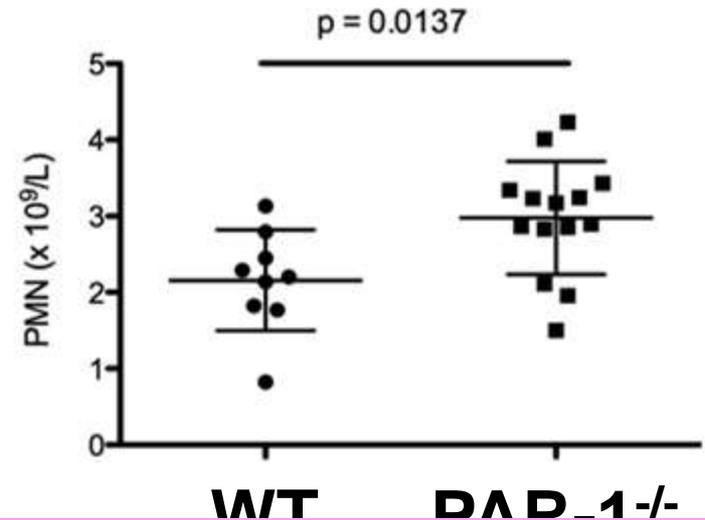
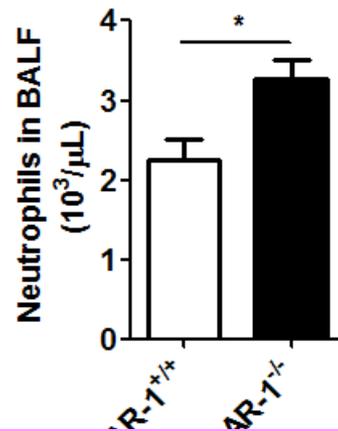
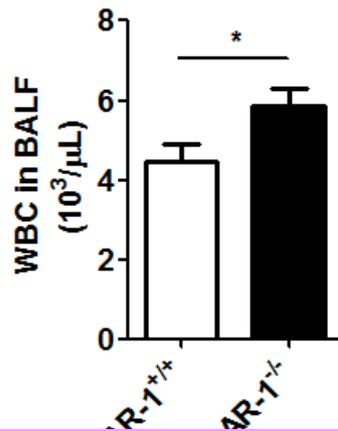
Hemavet

Cytospin

Neutrophils

WBC

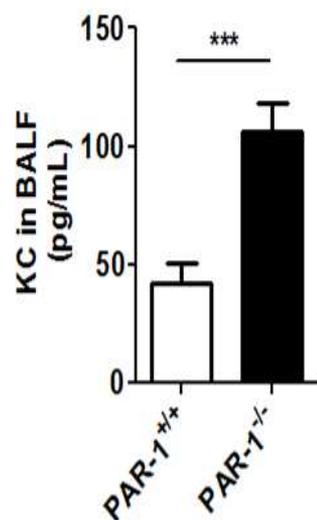
Neut.



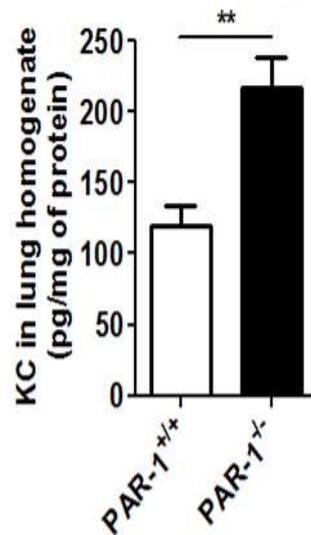
An increased in neutrophil infiltration into the lung may contribute to the increased mortality in PAR-1<sup>-/-</sup> mice

# Effect of PAR-1 Deficiency on KC Expression and Leukocyte Recruitment after IAV Infection

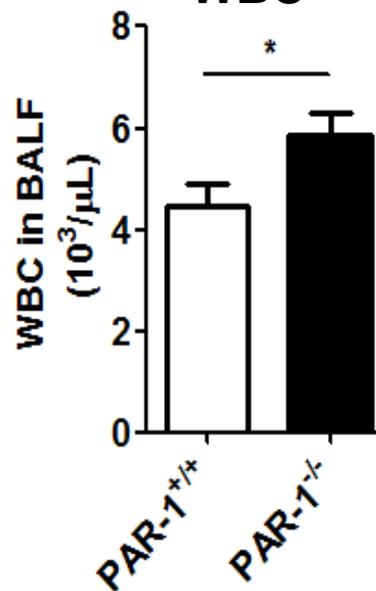
## KC in BALF



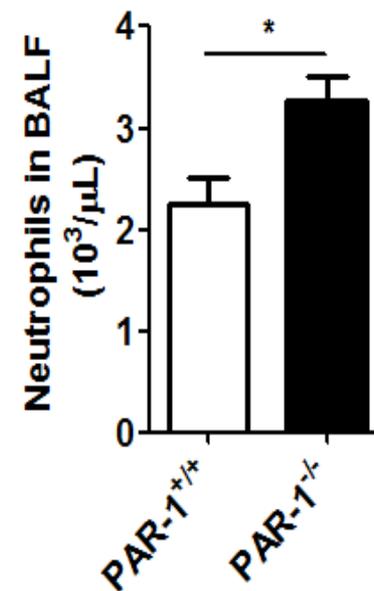
## KC in lung



## WBC



## Neut



# Ongoing Studies: Effect of Deletion of PAR-1 in Different Cell Types on IAV Infection

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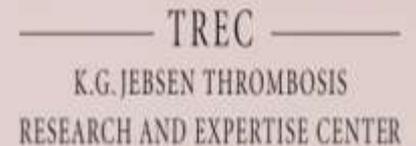
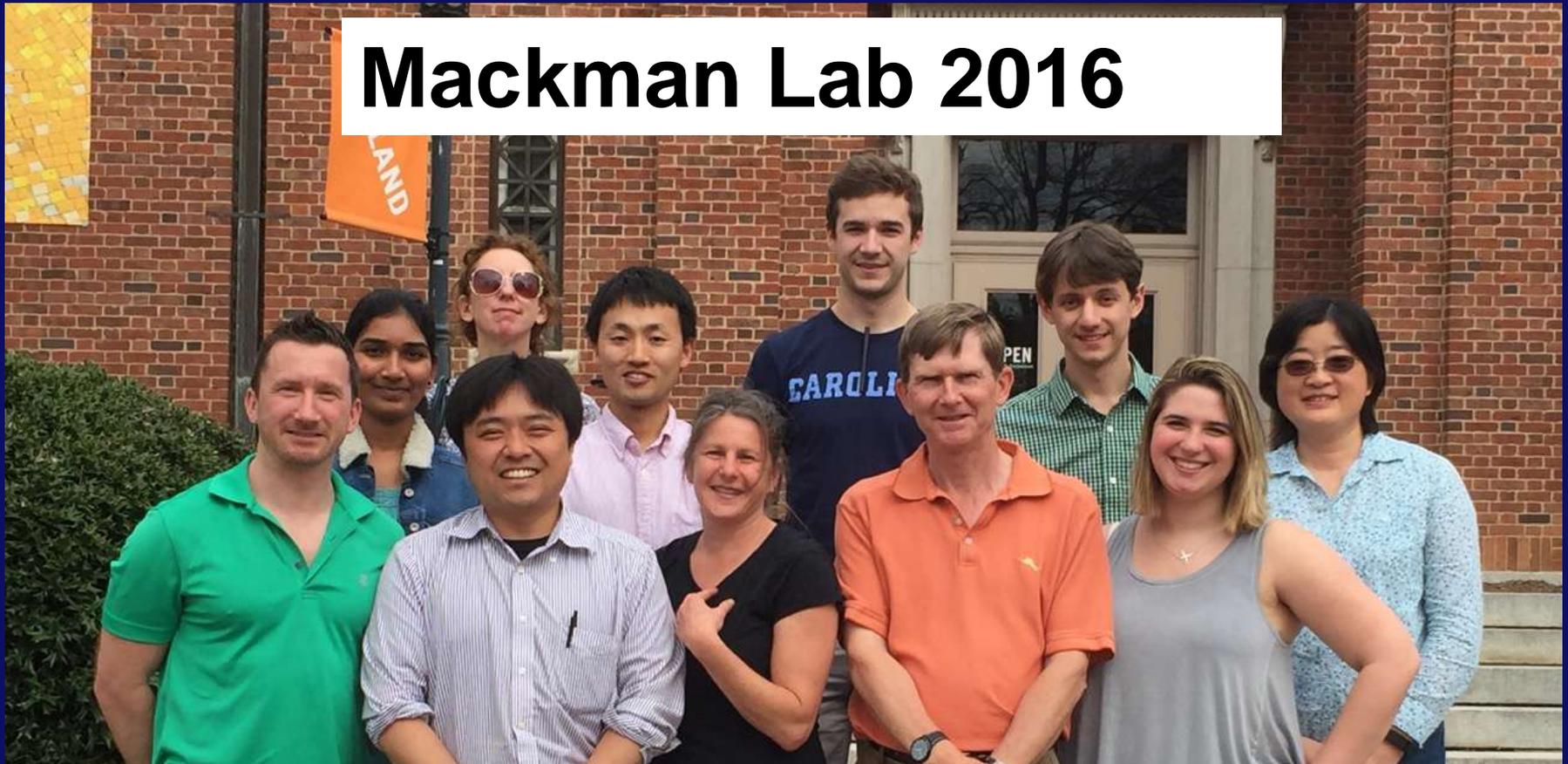
- PAR-1<sup>fl/fl</sup>, SpcCre- epithelial cell
- PAR-1<sup>fl/fl</sup>, LysMCre- myeloid cell
- PAR-1<sup>fl/fl</sup>, Tie2Cre- EC and hemat cell
- PAR-1<sup>fl/fl</sup>, iVE-cadherin- EC

# Conclusion

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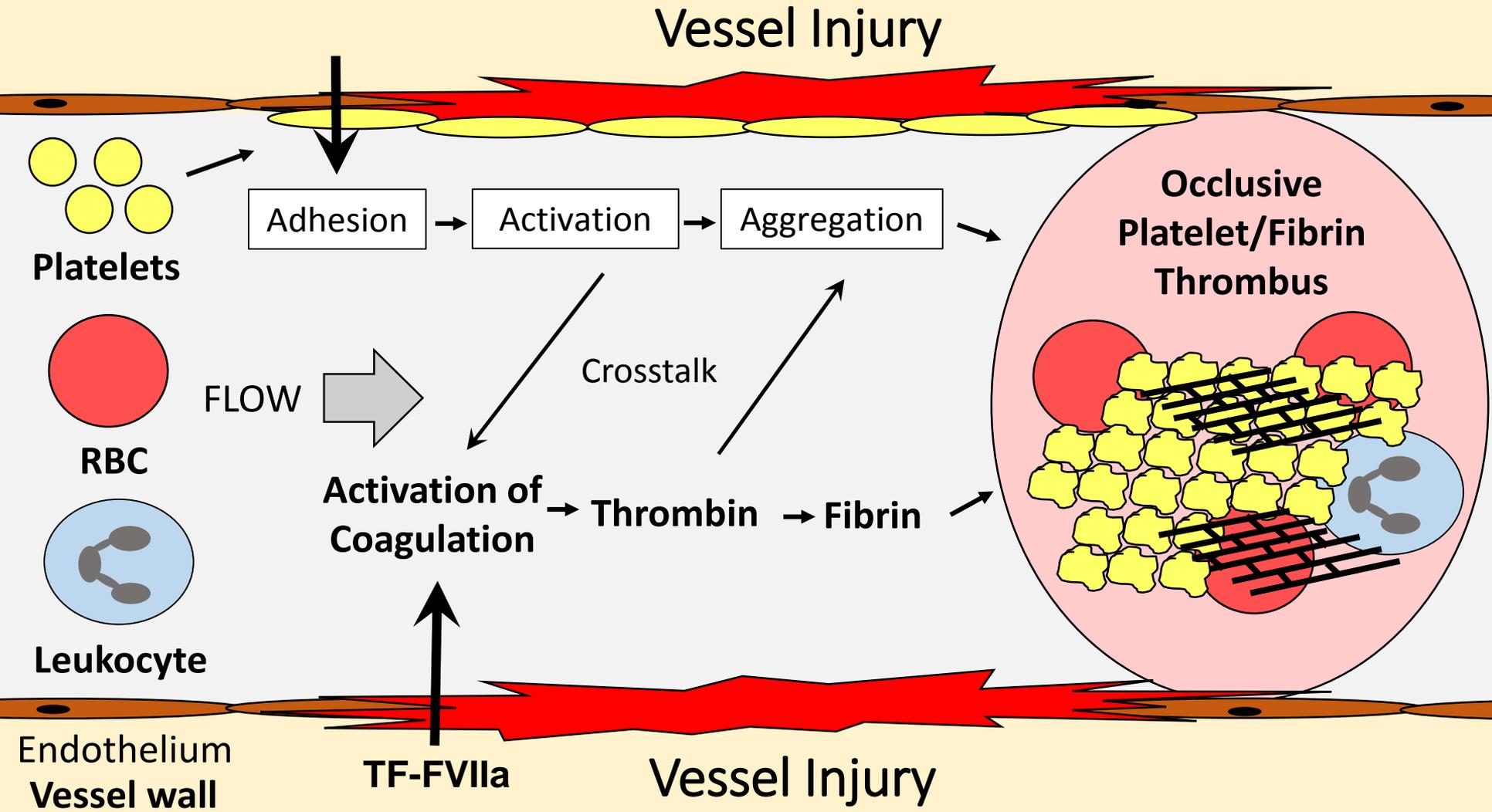
**PAR-1 deficient mice have increased mortality, vascular permeability and a dysregulated immune response after IAV infection**

# Mackman Lab 2016

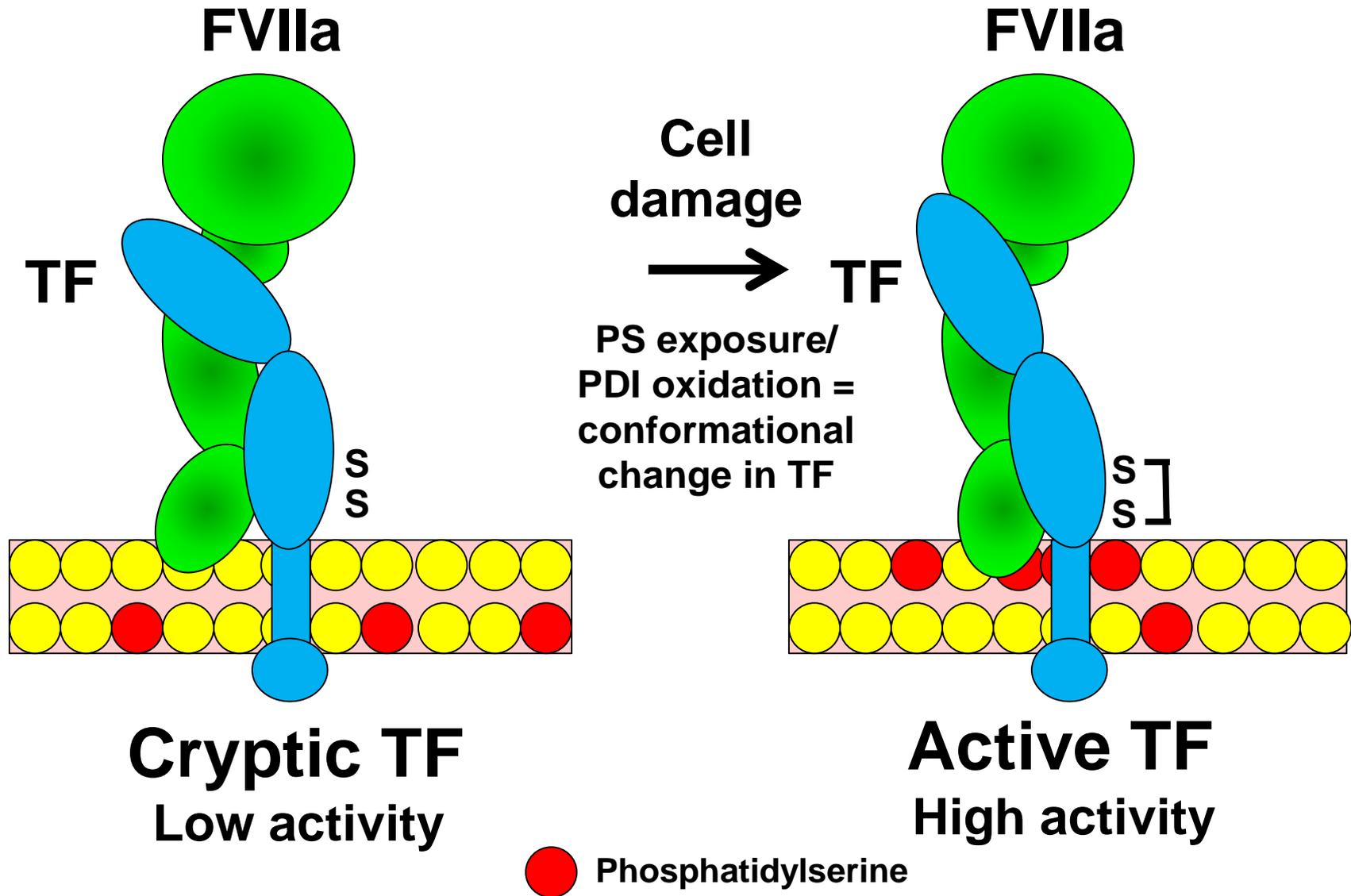




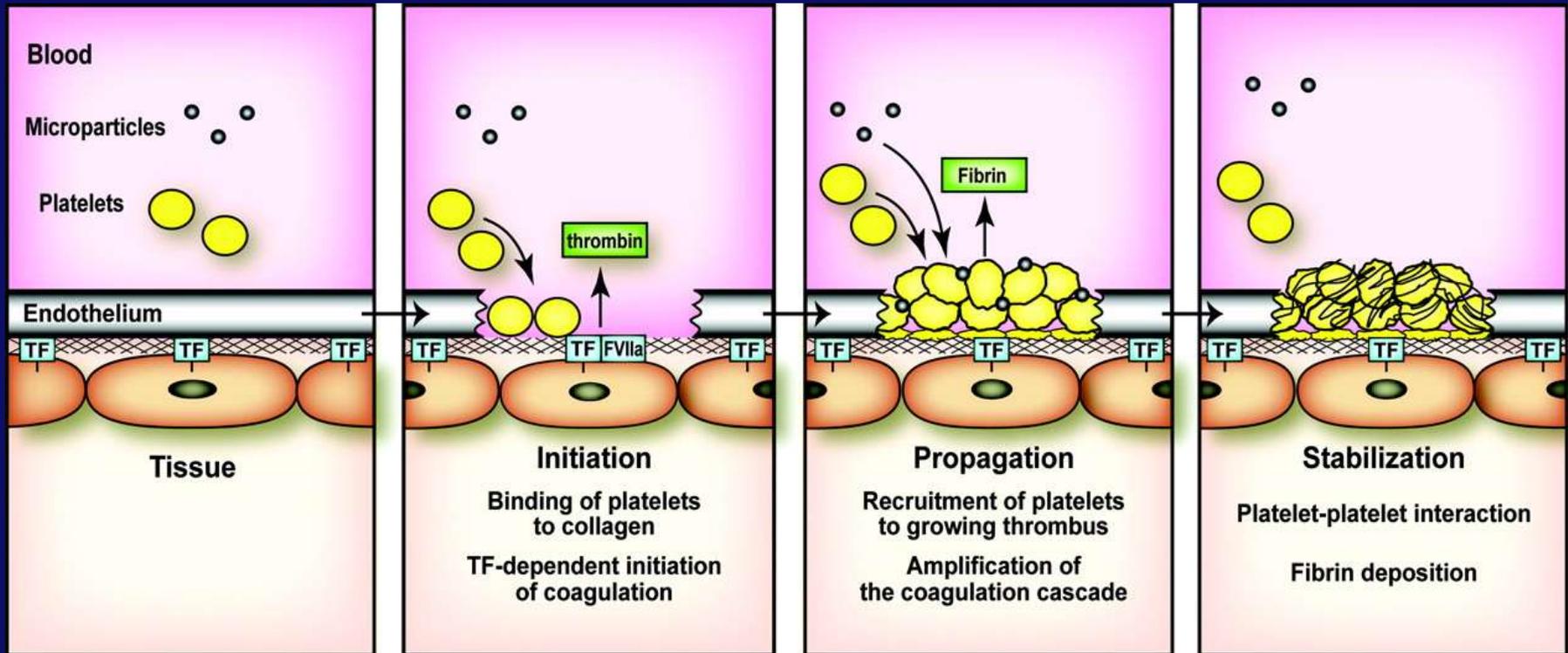
# Formation of an Occlusive Thrombus



# Regulation of TF Activity



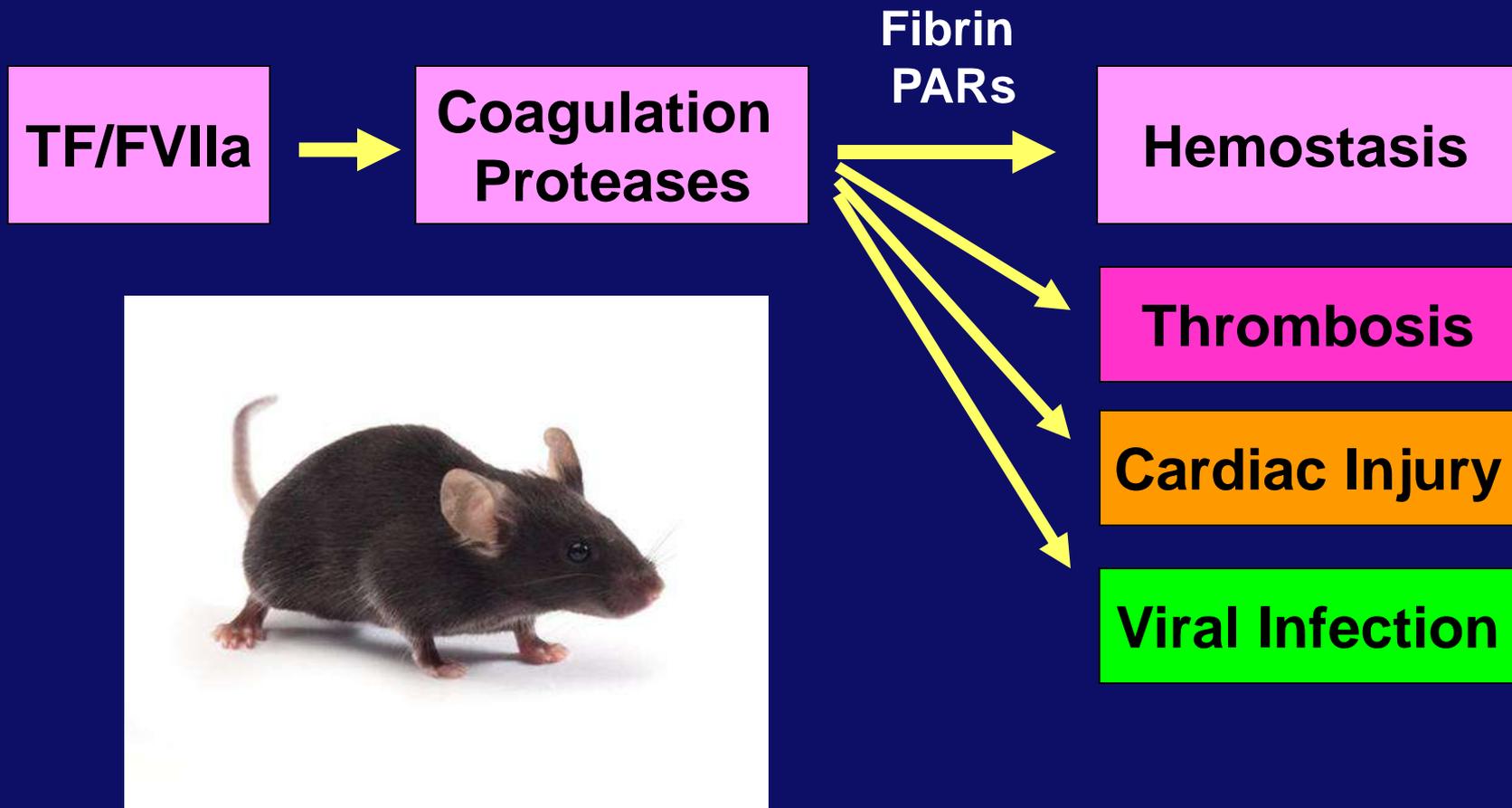
# Perivascular TF Maintains Hemostasis After Vessel Injury



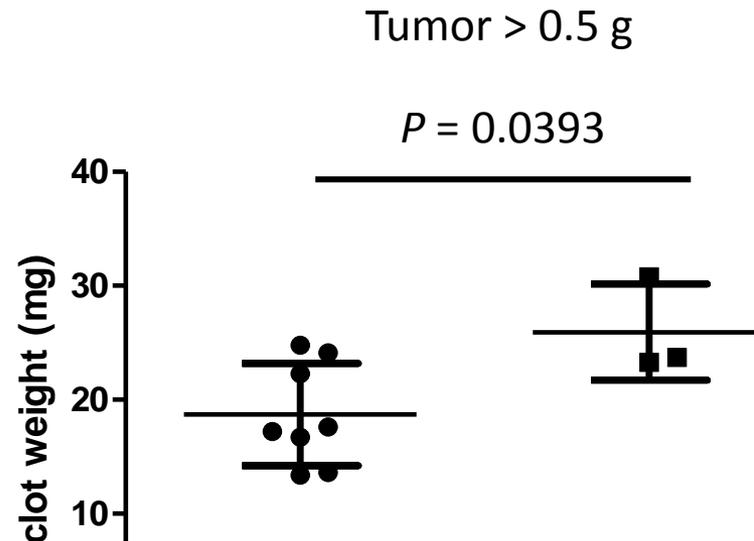
Mackman et al ATVB 2007

# Major Projects in the Mackman Lab

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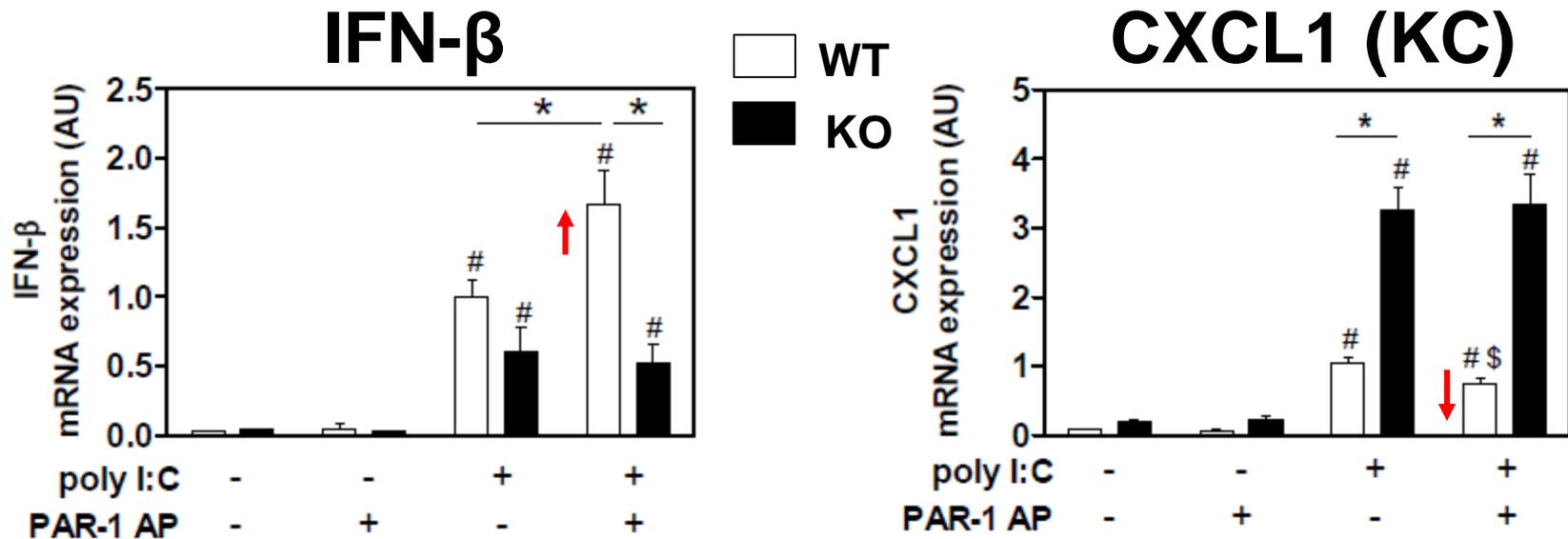
# Tumor Bearing Mice have Larger Thrombi in an IVC Full Ligation Model



**Determine the effect of inhibiting human TF on the enhanced thrombosis in tumor bearing mice**

Hisada Y unpublished data

# Role of PAR-1 in Poly I:C Induction of Antiviral Genes in Splenocytes



Antoniak S et al unpublished data

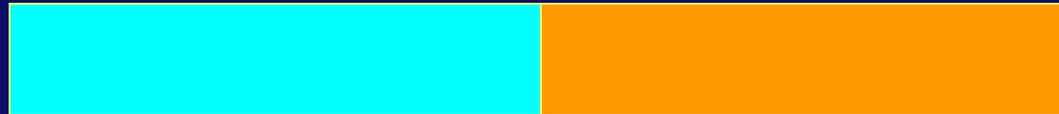
**Would inhibition of  
thrombin or PAR-1 in  
humans affect viral  
infections?**

# Ischemia-Reperfusion Injury

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**Ischemia**

**Reperfusion**



**Hypoxic Damage**

**Inflammatory Damage**

**Apoptosis/necrosis**

**Chemokines, cytokines,  
oxidants**

**Leukocyte recruitment  
and activation**

**Coagulation and fibrin  
deposition**

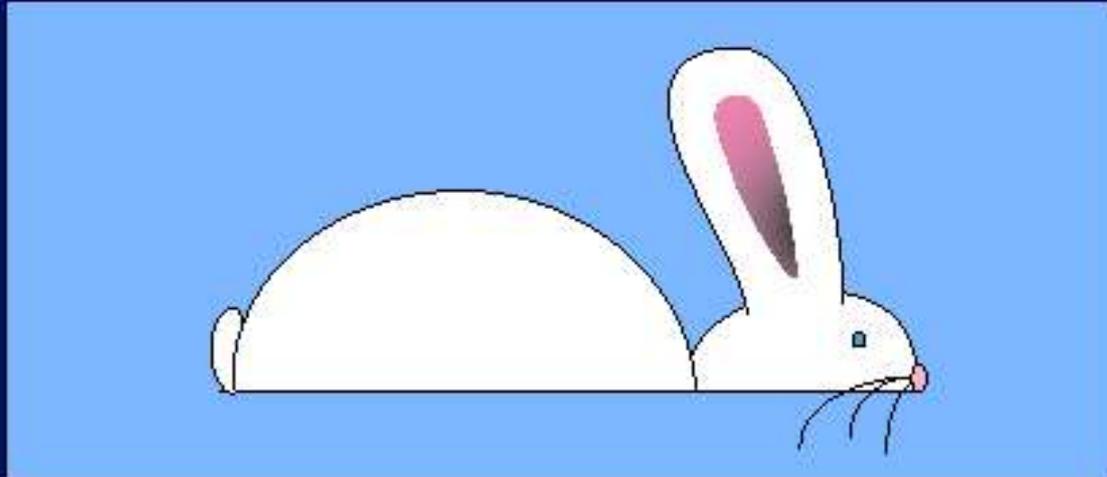
**Complement activation**

# Hypothesis

**Inhibition of TF and coagulation proteases will reduce infarct size after cardiac I/R injury in animal models**

# Rabbit Model of Cardiac I/R Injury

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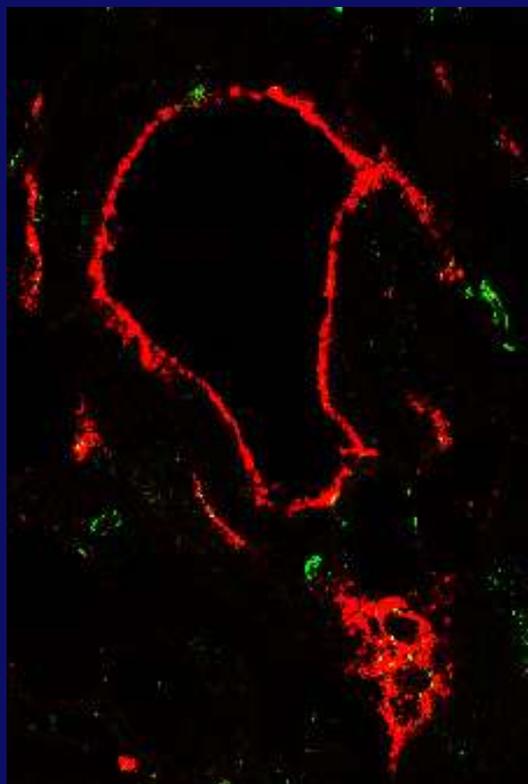
**Ischemia = 45 min**

**Reperfusion = 120 min**

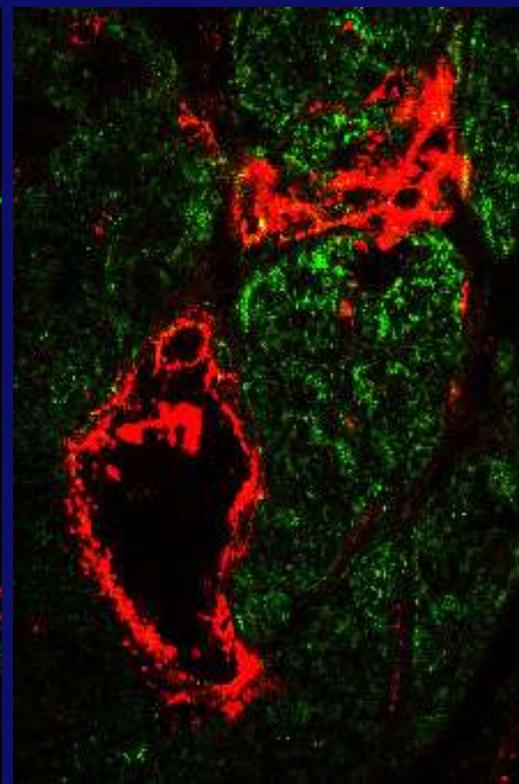
# Cardiac I/R Injury Damages the Endothelial Barrier and Leads to Extravascular Fibrin Deposition

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Rabbit  
model



sham



I/R

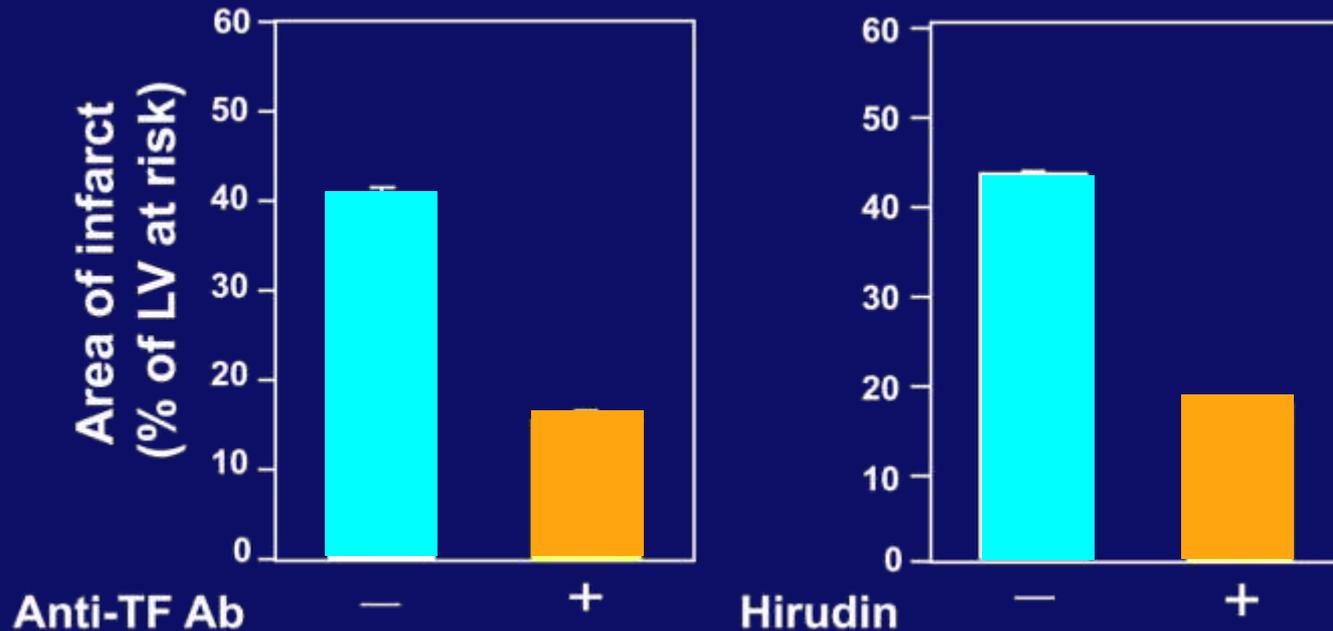
Red = vWF

Green = fibrin

Erlich et al., *Am J Path* 2000 (Cover)

# Effect of Inhibition of TF or Thrombin on Infarct Size in a Rabbit Cardiac I/R Model

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Inhibition of TF or thrombin also reduced inflammation

Erlich et al., *Am J Path* 2000

# Effect of Inhibition of Thrombin or a Deficiency of PAR-1 or PAR-2 on Infarct Size in a Mouse Cardiac I/R Model

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**Pawlinski R et al Circ  
2007- Thrombin  
inhibition or PAR-1  
deficiency.**

**Antoniak S et al ATVB  
2010- PAR-2 deficiency**

# Clinical Studies with Rivaroxaban

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**ATLAS ACS-2-TIMI-51 trial- the factor Xa inhibitor rivaroxaban reduced death due to cardiovascular causes (MI and stroke) in patients with a recent ACS event** Mega J et al NEJM 2012.

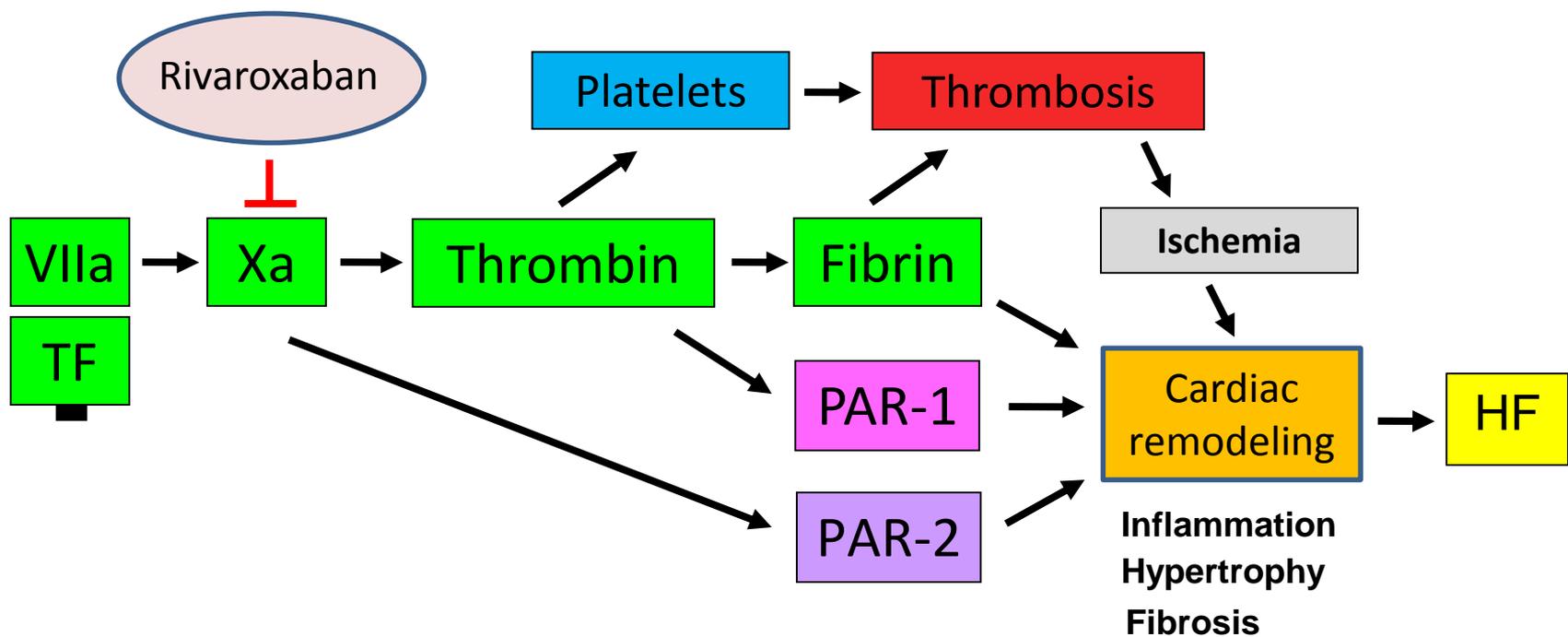
**In a subgroup analysis of HF patients, treatment with rivaroxaban 2.5 mg twice daily plus antiplatelet therapy demonstrated decreased rates of cardiovascular events and death compared with antiplatelet therapy alone.**

**COMMANDER-HF is a phase III clinical trial to evaluate the effect of the new oral anticoagulant rivaroxaban in patients with chronic heart failure.**

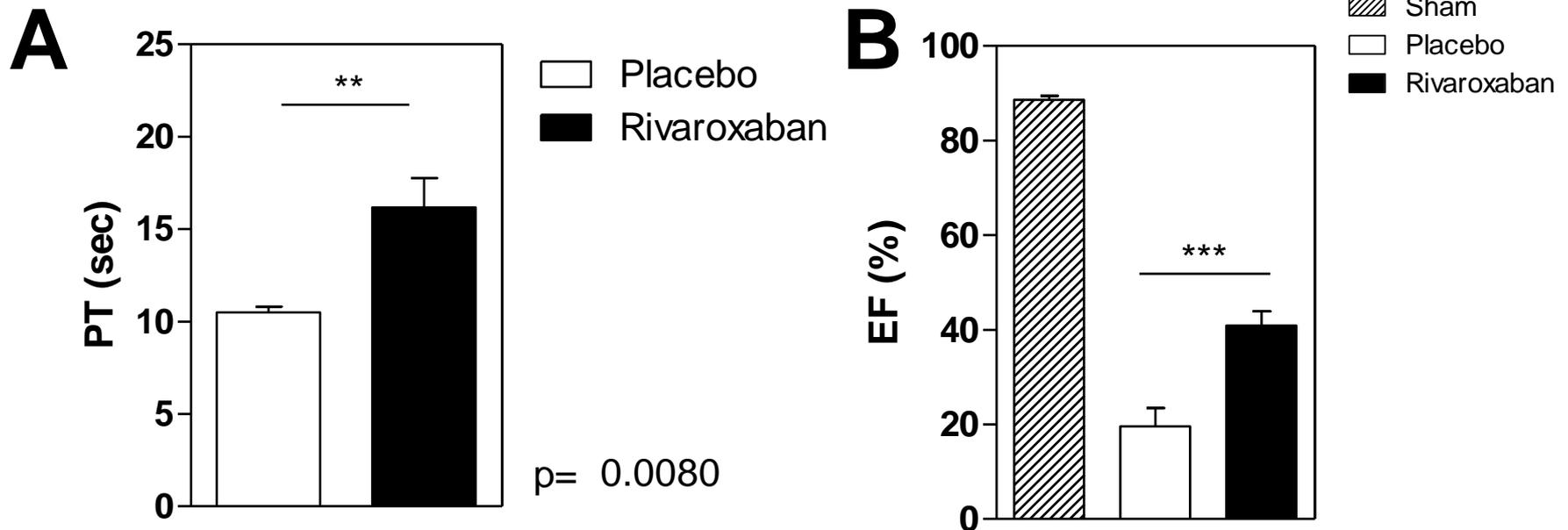
# Hypothesis

**Inhibition of FXa with  
rivaroxaban will reduce cardiac  
remodeling and heart failure in  
mice**

# Effect of Rivaroxaban on Cardiac Remodeling and Heart Failure



# Effect of Rivaroxaban on Cardiac Remodeling and Heart Failure in a Mouse Model of LAD Ligation



28 days after LAD ligation

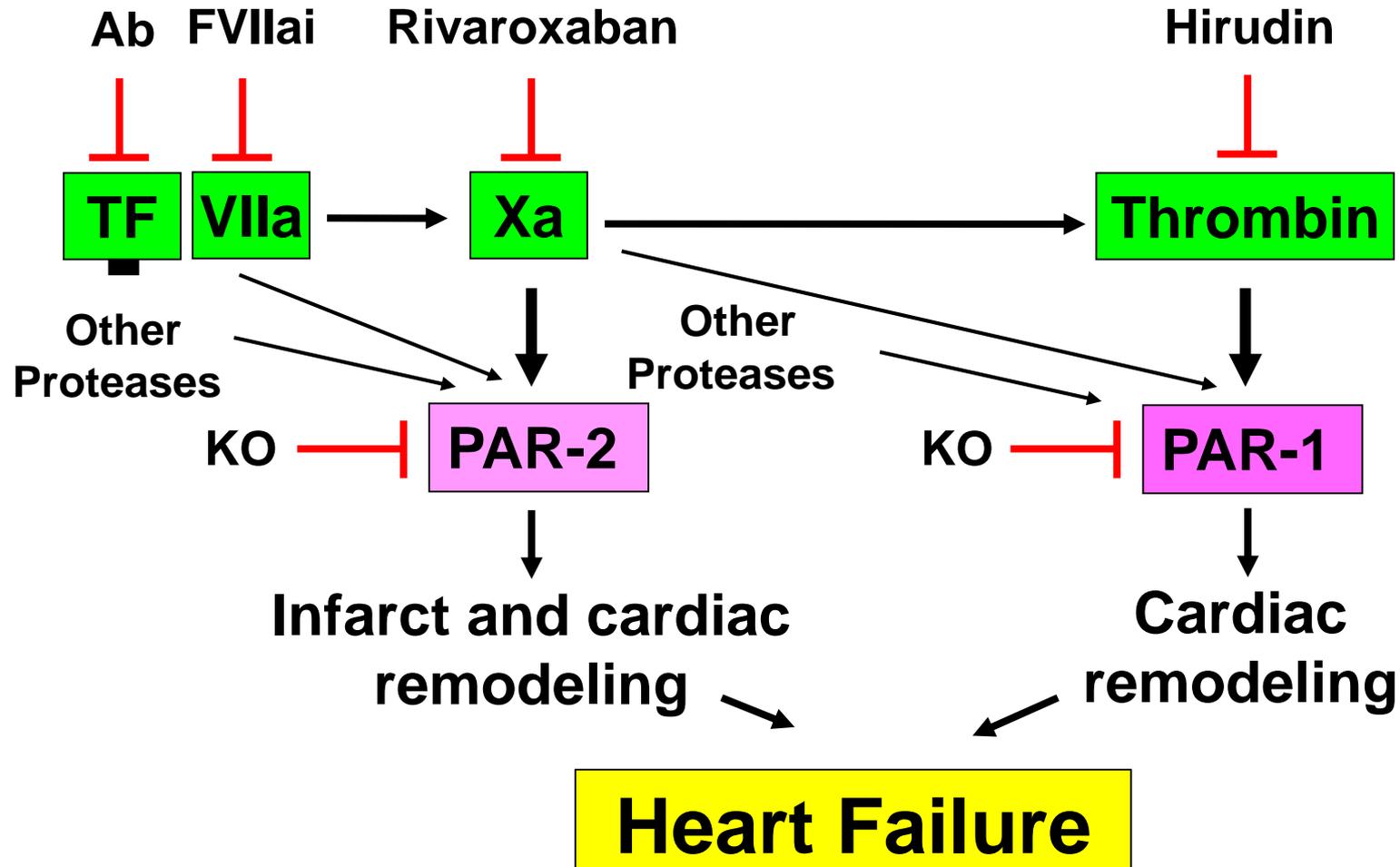
Bode M unpublished results

# Conclusion

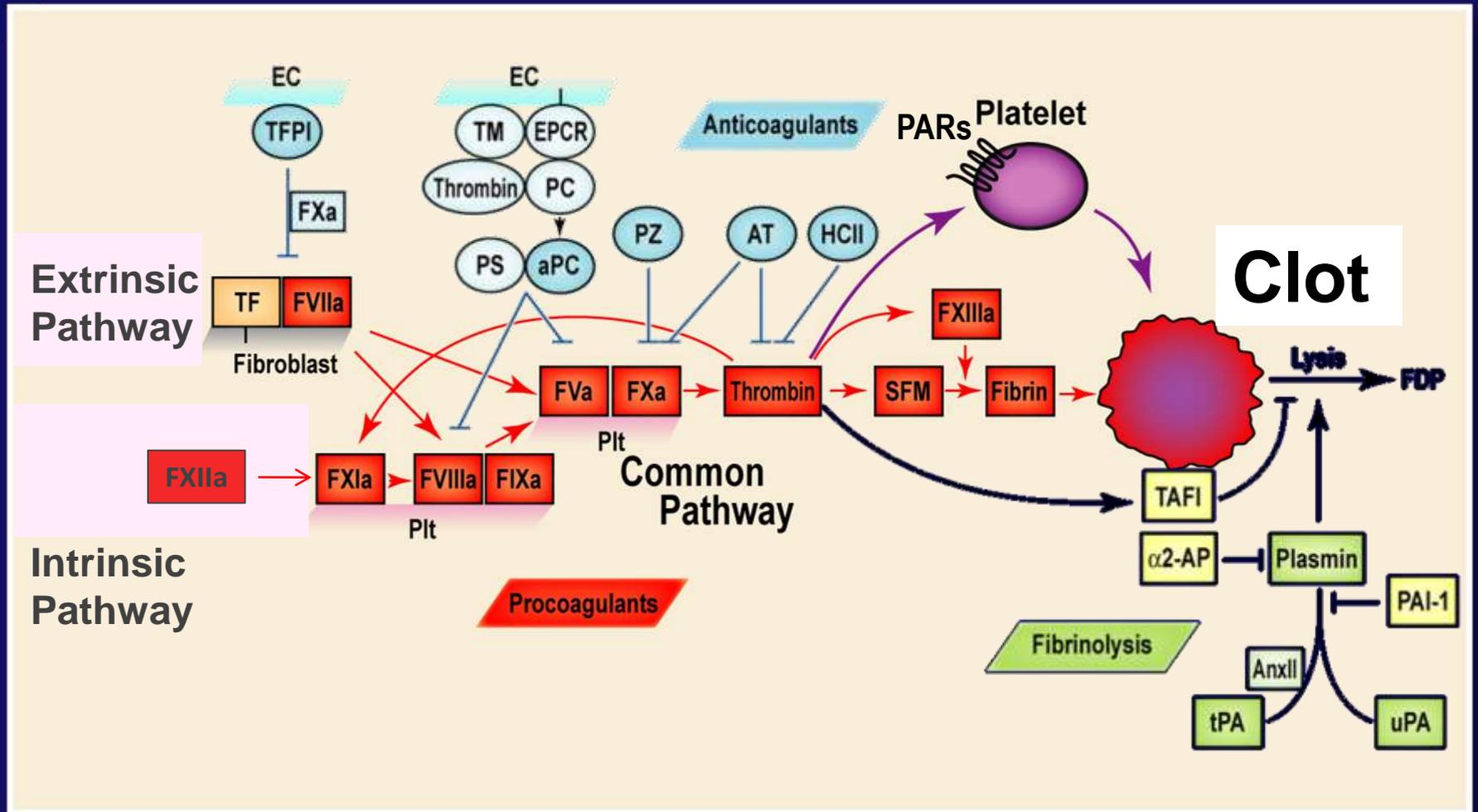
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**Inhibition of TF, coagulation proteases or PAR-1 may reduce cardiac remodeling and heart failure in MI patients**

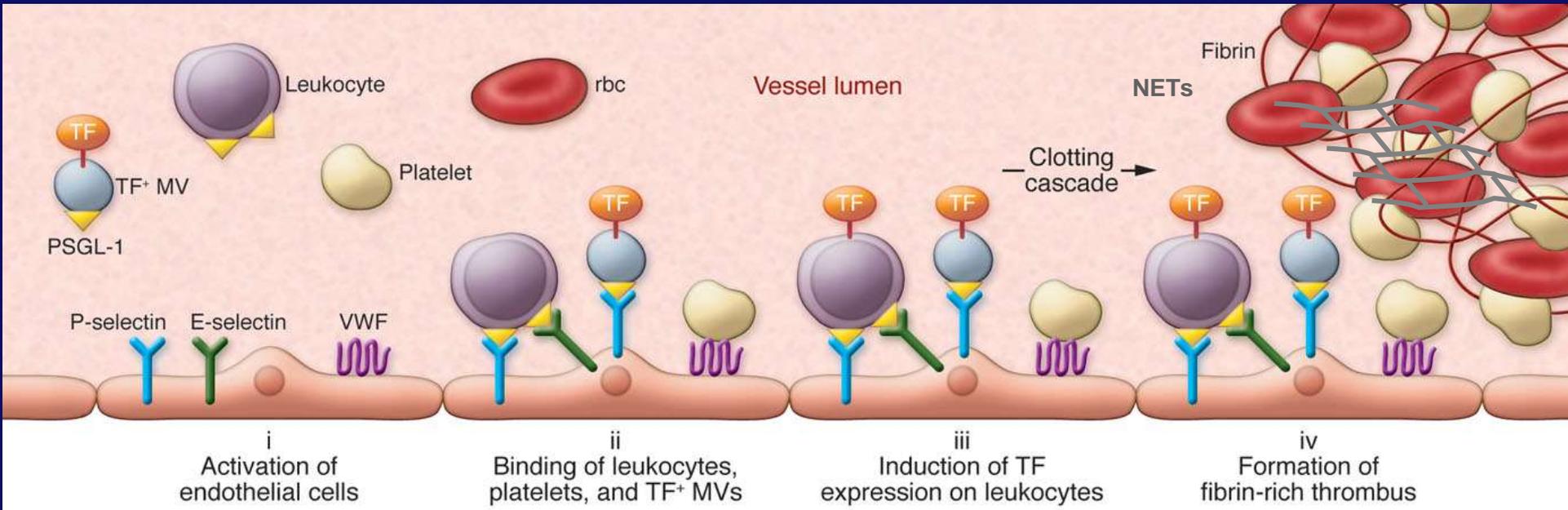
# Role of TF, Coagulation Proteases and PARs in Cardiac I/R Injury



# Formation and Lysis of a Clot

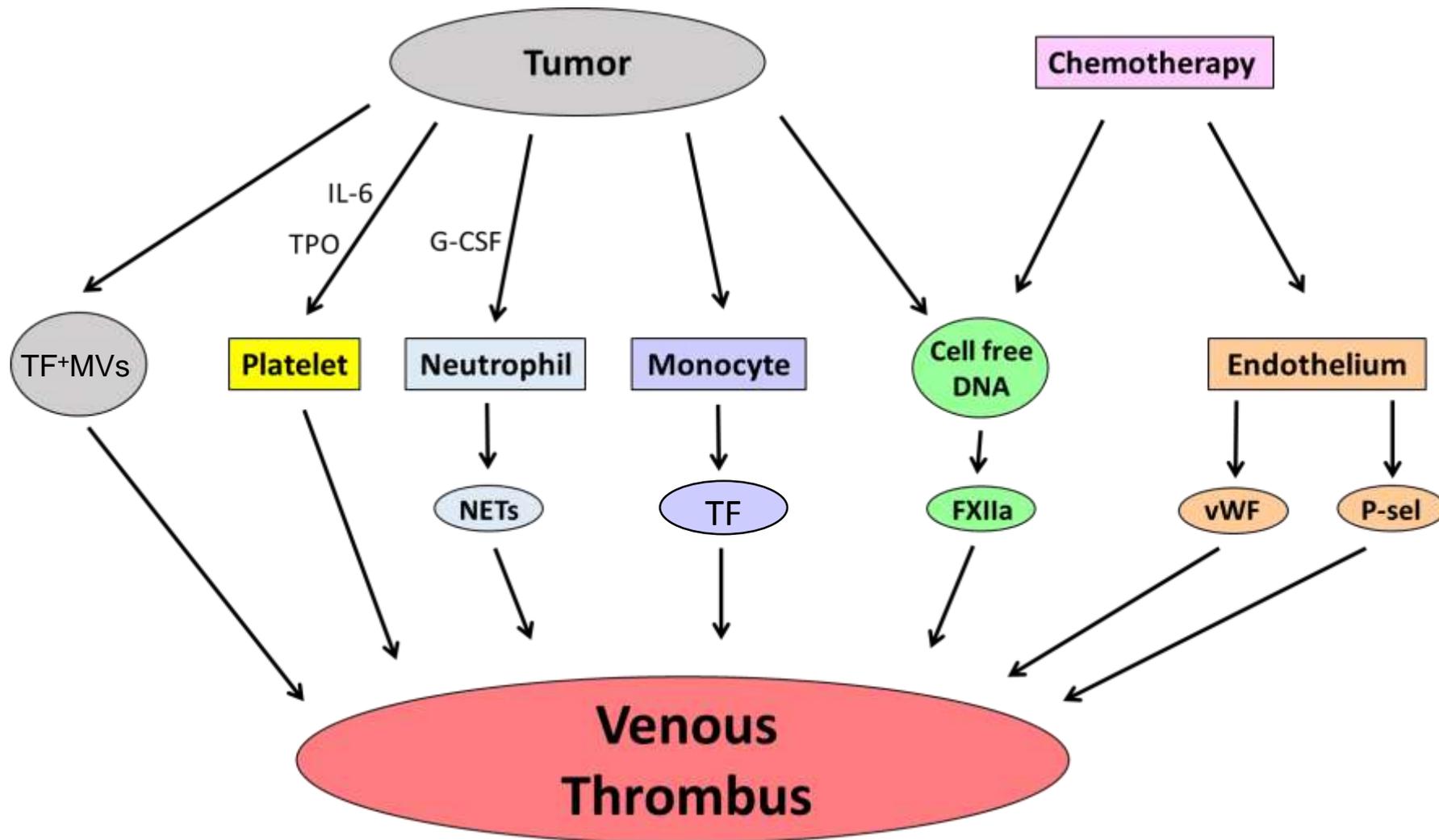


# Mechanisms of Venous Thrombosis

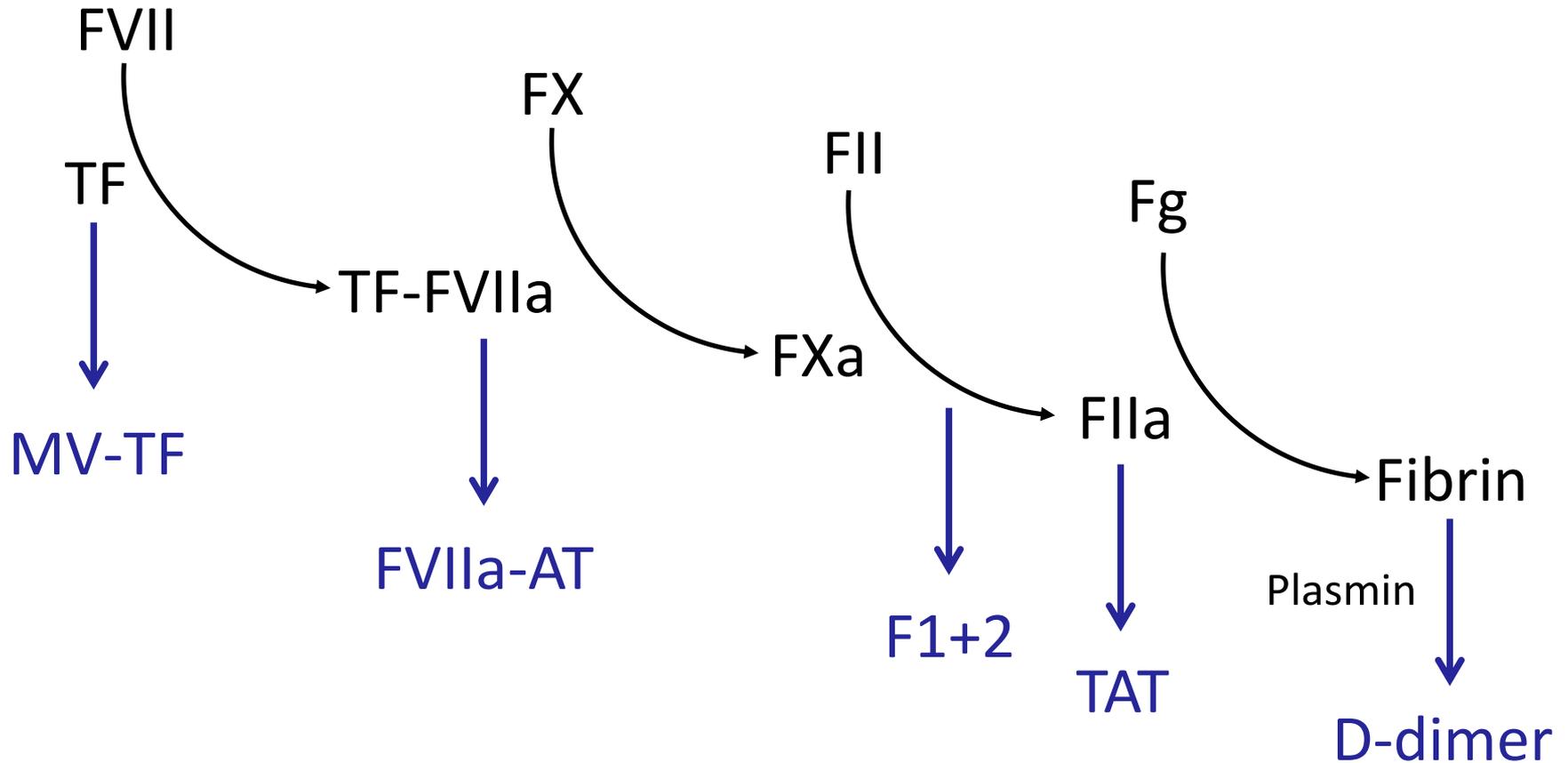


Mackman JCI 2012

# Possible Mechanisms of VTE in Cancer Patients



# Biomarkers Associated with Activation of the Clotting Cascade





## Factor XI Antisense Oligonucleotide for Prevention of Venous Thrombosis

Harry R. Büller, M.D., Claudette Bethune, Ph.D., Sanjay Bhanot, M.D., Ph.D., David Gailani, M.D., Brett P. Monia, Ph.D., Gary E. Raskob, Ph.D., Annelise Segers, M.D., Peter Verhamme, M.D., and Jeffrey I. Weitz, M.D.

2015;372:232-240

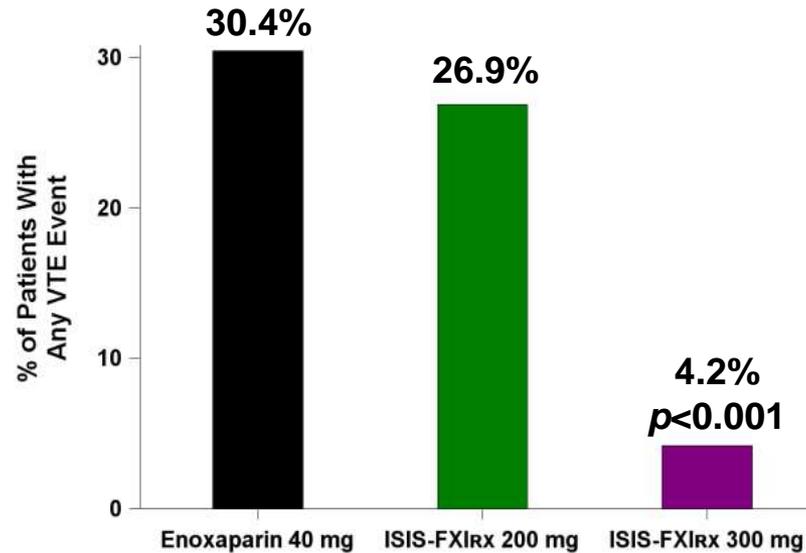
- Effect of decreased levels of FXI on VTE in pts undergoing total knee replacement
- Two doses of a FXI ASO were used: 200 mg ~41% FXI; 300 mg ~22% FXI.
- Enoxaparin was used as a control.

Slides from Dr. J. Weitz

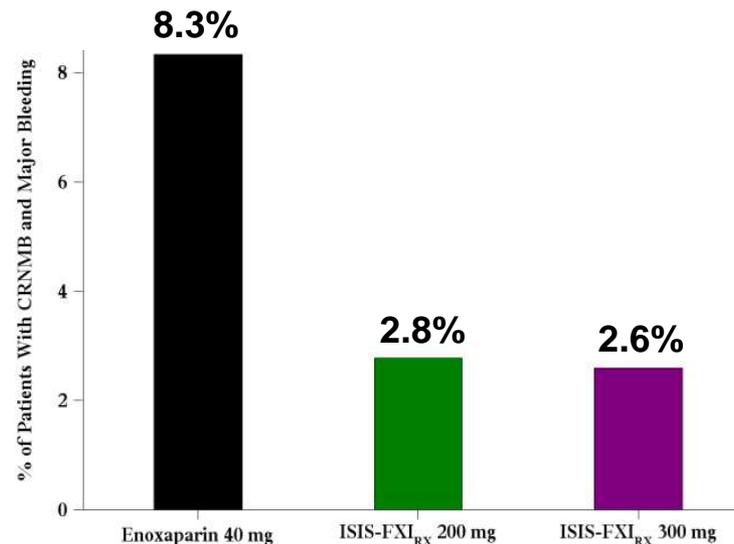
- **Primary efficacy outcome was incidence of venous thromboembolism (VTE) (asymptomatic and symptomatic)**
- **Principal safety outcome was bleeding**

# Primary Efficacy and Safety Outcomes

VTE



Bleeding



# Conclusion

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**FXI appears to be an attractive  
new target for preventing VTE  
without increase bleeding**

# Measurement of TF Antigen and Activity in Whole Blood, Plasma and Microparticles

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- Plasma- TF antigen
- MPs- TF antigen flow cytometry
- MPs- TF antigen impedance

TF  
antigen

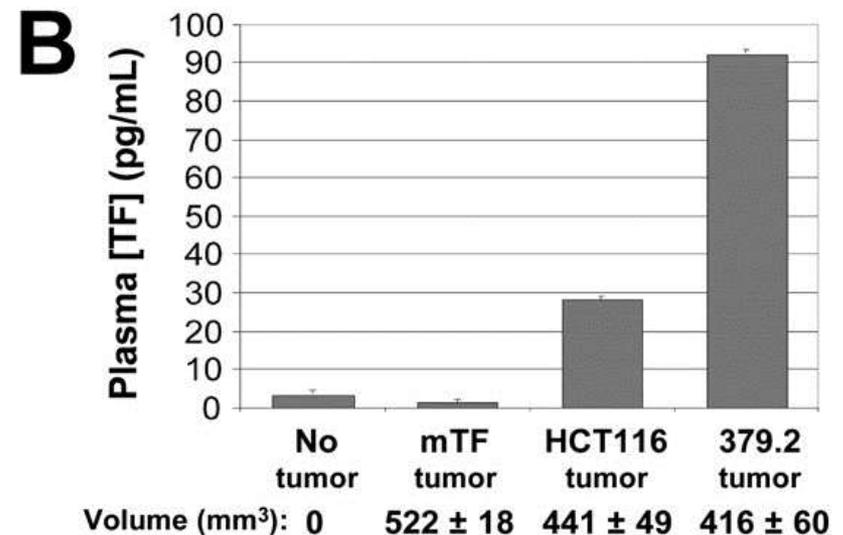
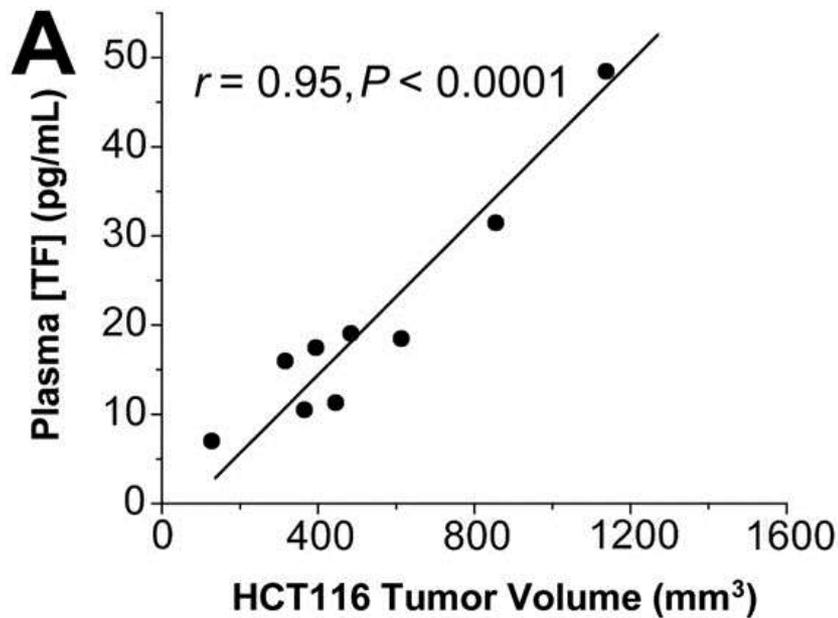
- Whole blood- TF activity
- Plasma- TF activity

- MP (pellet)- TF activity \*
- MP (capture)- TF activity
- Fibrin generation

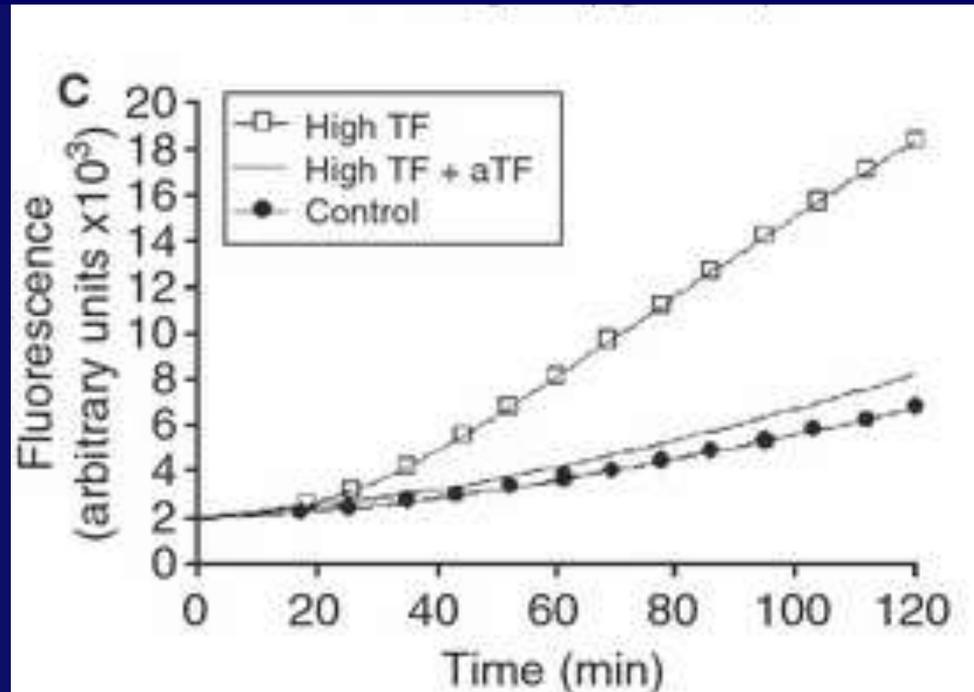
TF  
activity

# Mouse Studies

# Tumors Release Human TF into the Circulation in SCID Mice containing Human Tumors



# Tumor-derived Human TF is Functional in an In Vitro Assay

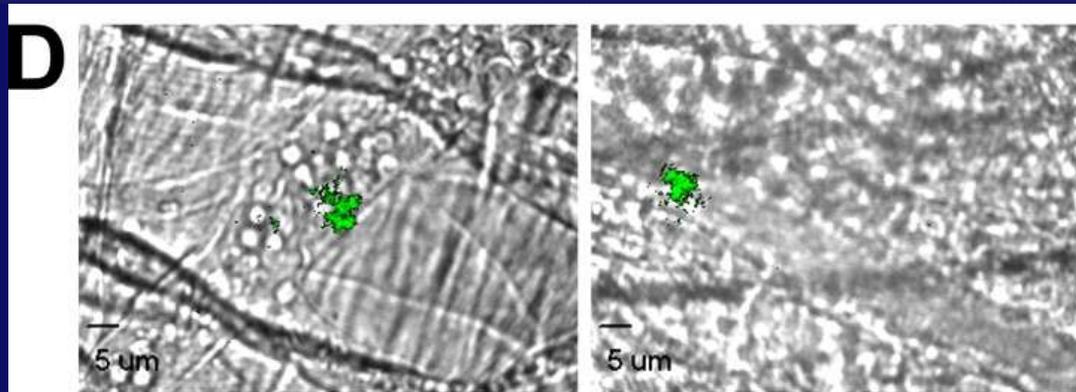


Human pancreatic L3.6pl cells grown orthotopically in nude mice

Davila et al JTH 2008

# Tumor-derived MPs Accumulate at the Site of Thrombus Formation In Vivo

Mouse Panc02 pancreatic cells labeled with pEGFP



Venule

Arteriole

JEM

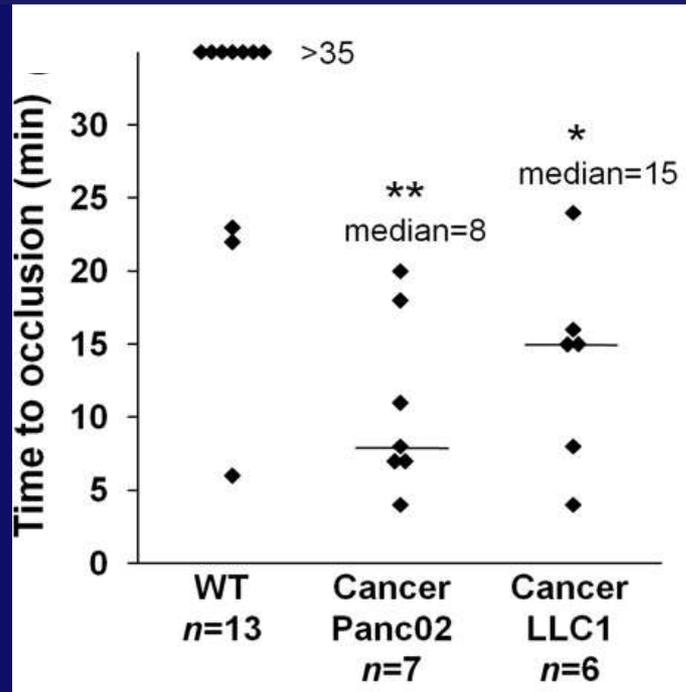
Thomas et al JEM 2009

# Tumor-bearing Mice have Larger Thrombi in a Mesenteric Vessel Model

Panc02 = mouse pancreatic carcinoma

LLC = mouse lewis lung carcinoma

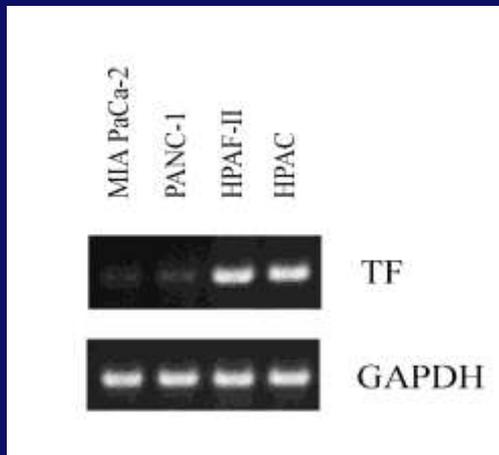
S.C.



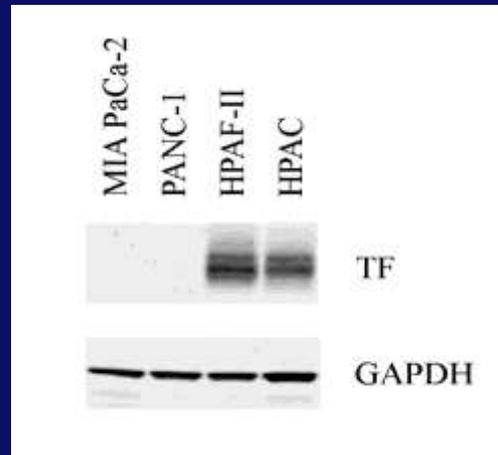
Thomas et al JEM 2009

# TF Expression in Various Human Pancreatic Cell Lines In Vitro

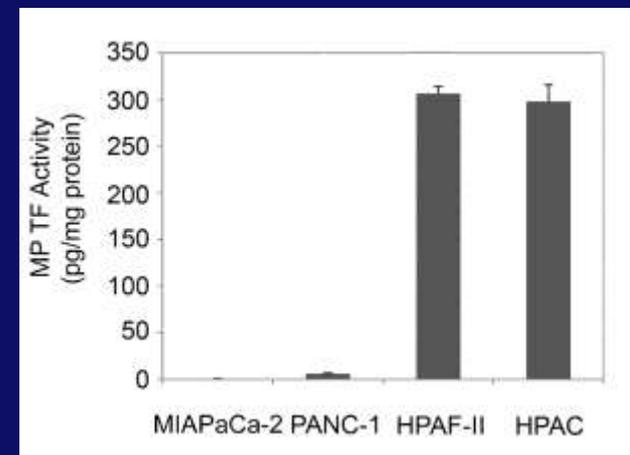
## TF mRNA



## TF Protein



## MP TF Activity



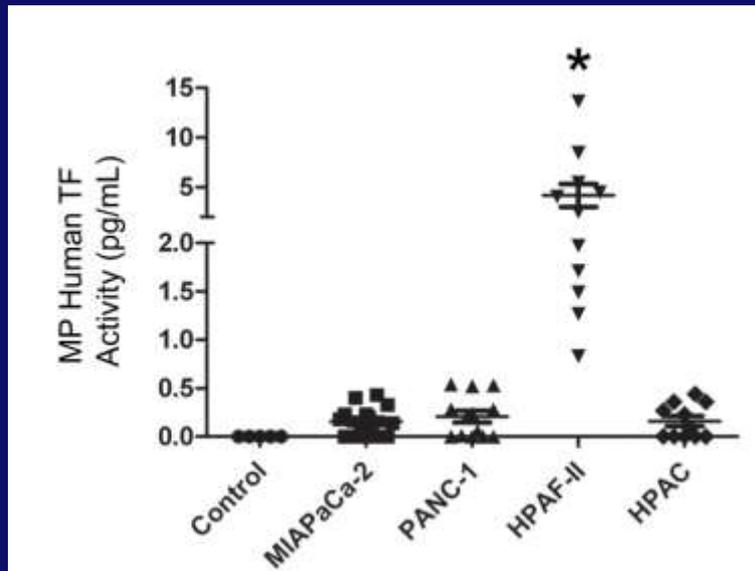
**Cells**

**Medium**

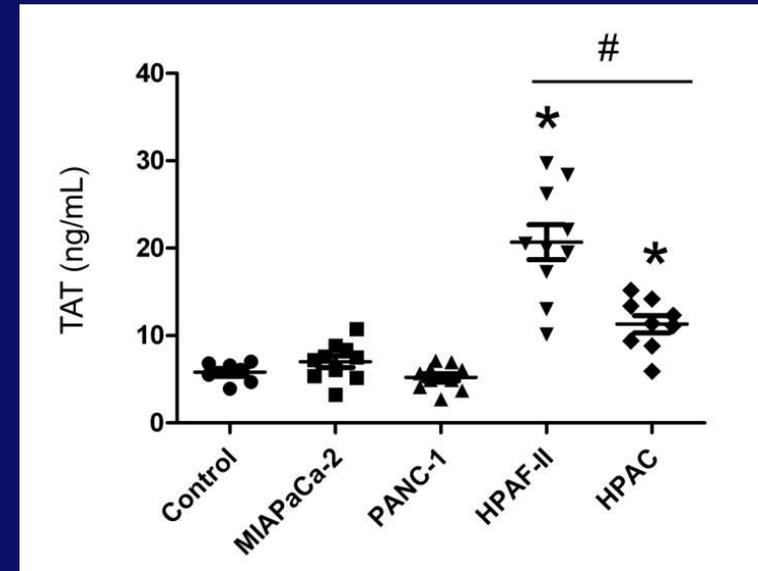
Wang et al unpublished data

# MP TF Activity and Activation of Coagulation in Nude Mice containing Human Pancreatic Tumors

## MP TF activity



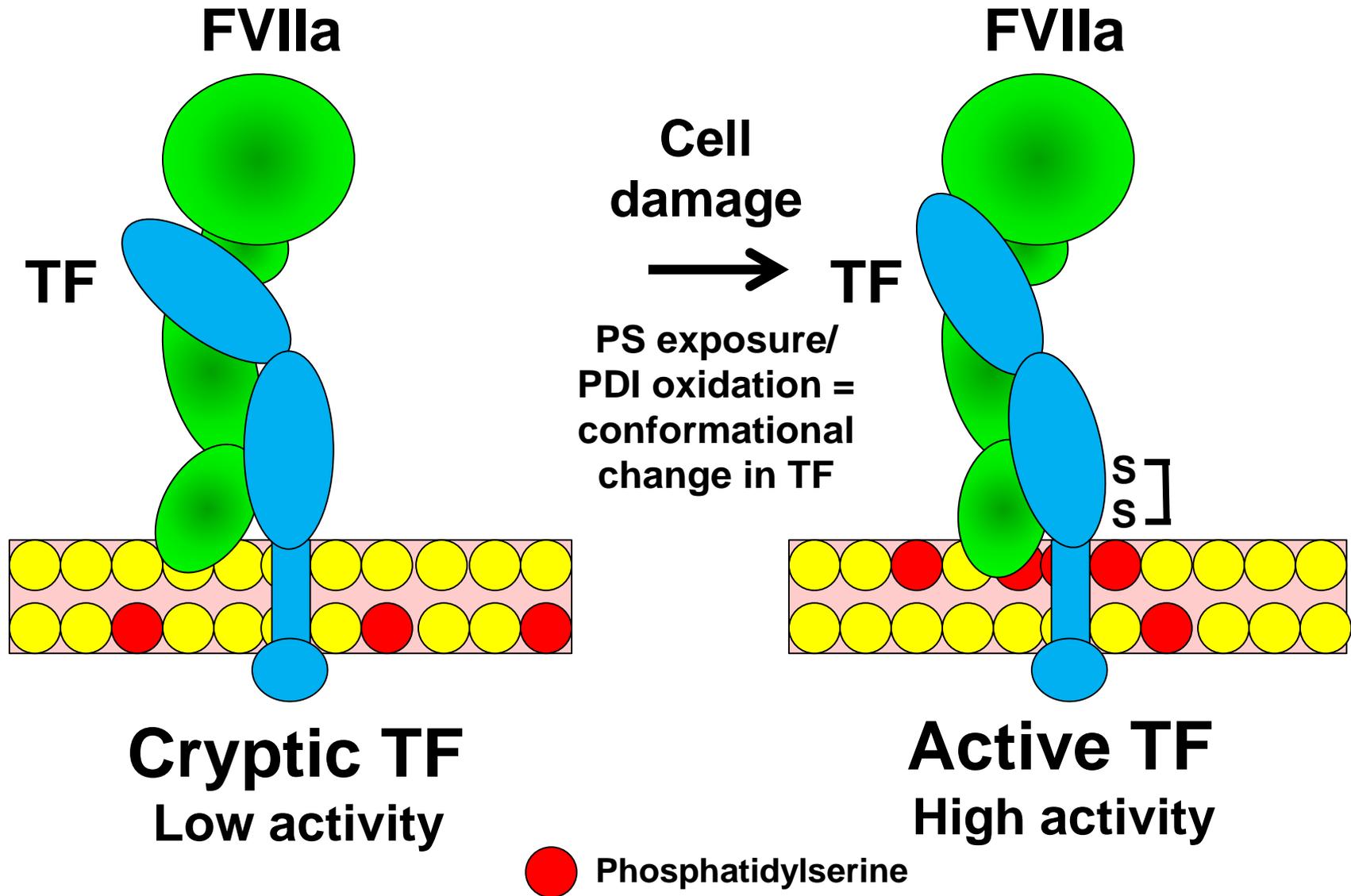
## TAT



An anti-human TF Ab reduces the increases in TAT

Wang et al unpublished data

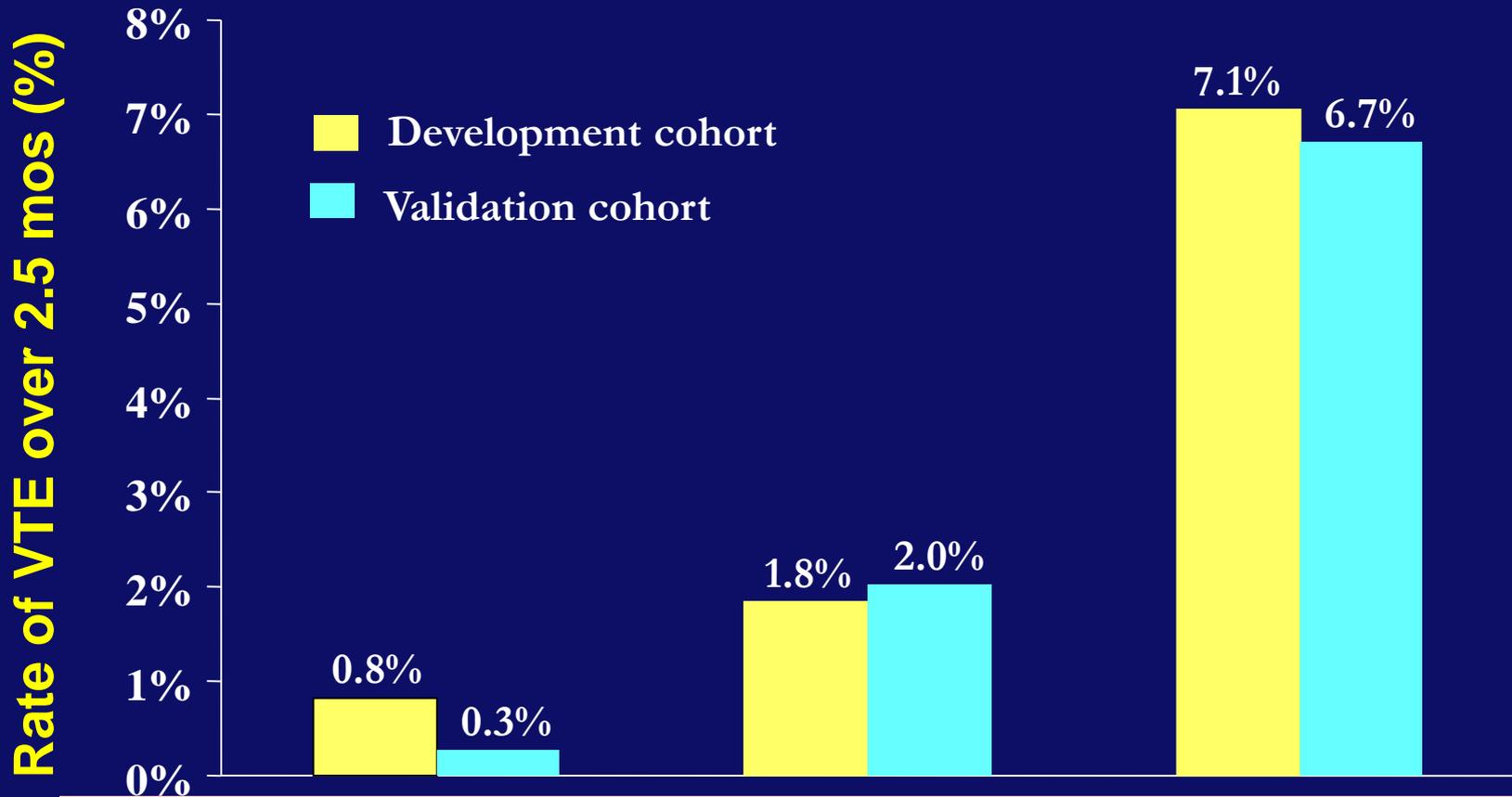
# Regulation of TF Activity



# Predictive Model-Khorana Score

Patient Characteristic	Score
Site of Cancer	
Very high risk (stomach, pancreas)	2
High risk (lung, lymphoma, gynecologic, GU excluding prostate)	1
Platelet count $\geq 350,000/\text{mm}^3$	1
Hgb $< 10\text{g/dL}$ or use of ESA	1
Leukocyte count $> 11,000/\text{mm}^3$	1
BMI $\geq 35$	1

# Predictive Model Validation



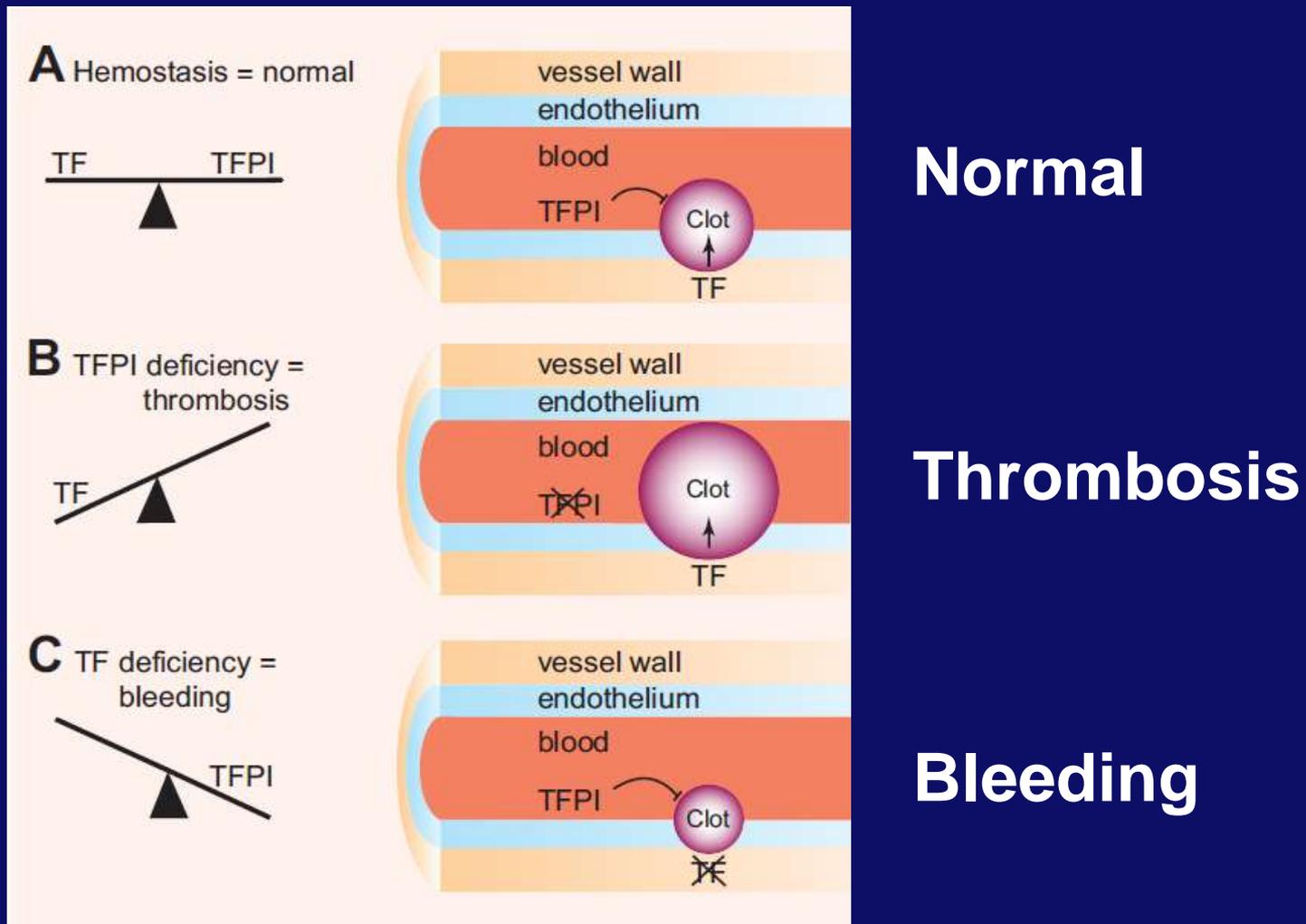
**D-dimer and soluble P-selectin have been added- Ay et al Blood 2010**

# Biomarkers

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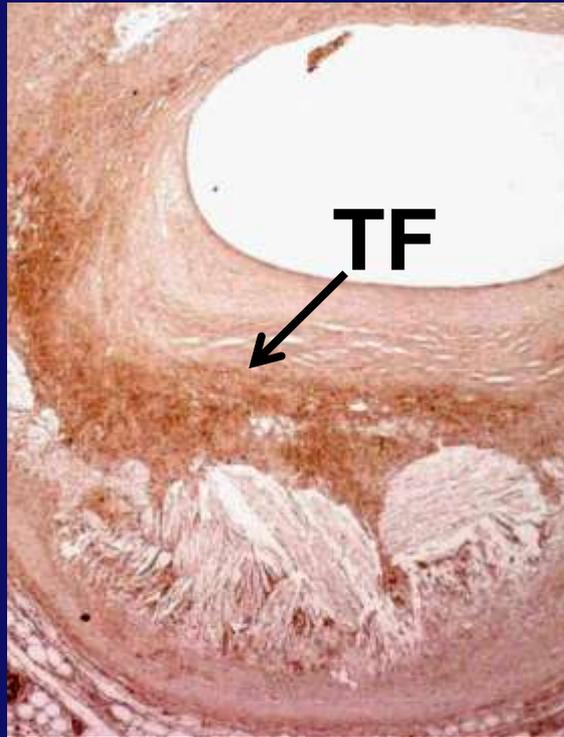
A measurable substance in an organism whose presence is indicative of some phenomenon such as disease, infection, or environmental exposure.

# Balancing Clotting



# High Levels of TF are Present in Atherosclerotic Plaques

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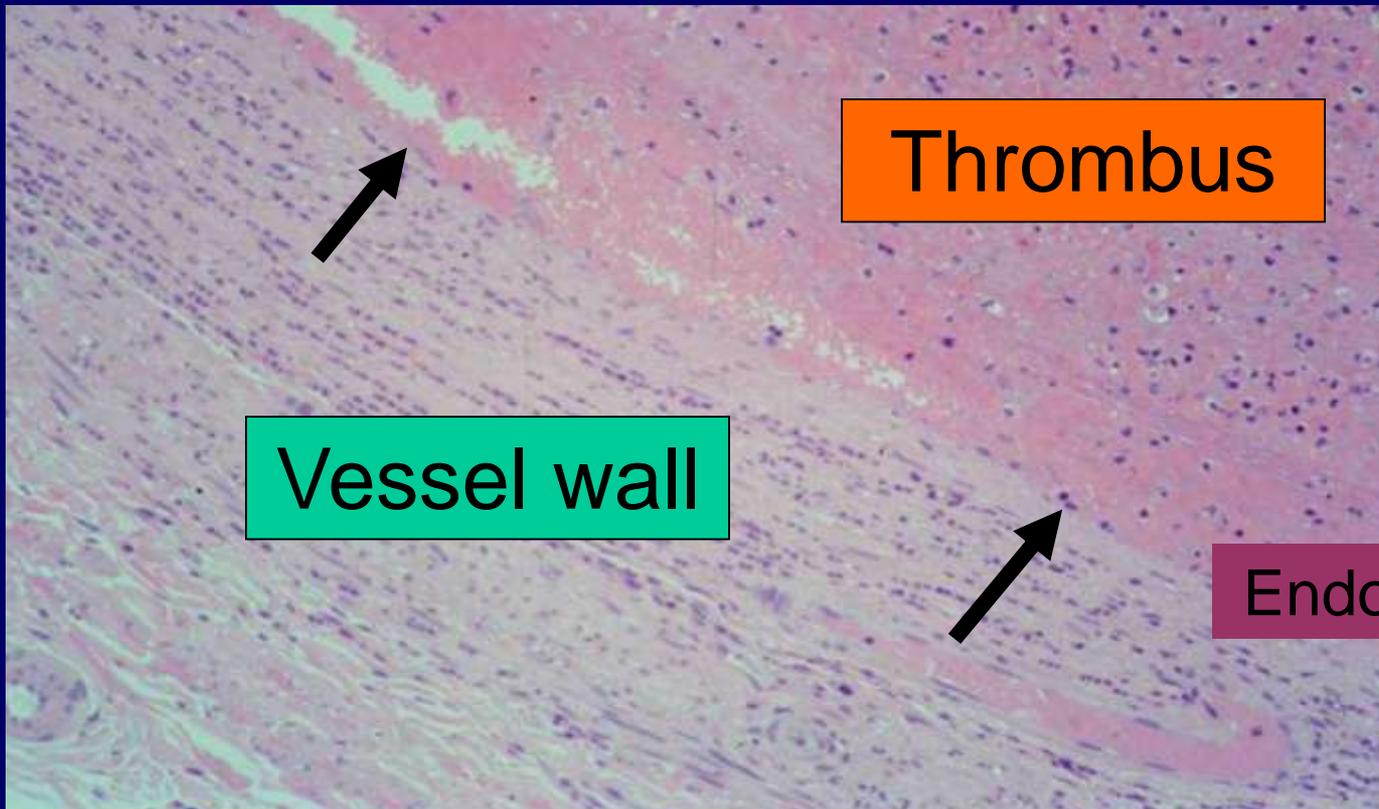


**“Thrombotic” TF**

Slide provided by Dr. Y. Asada

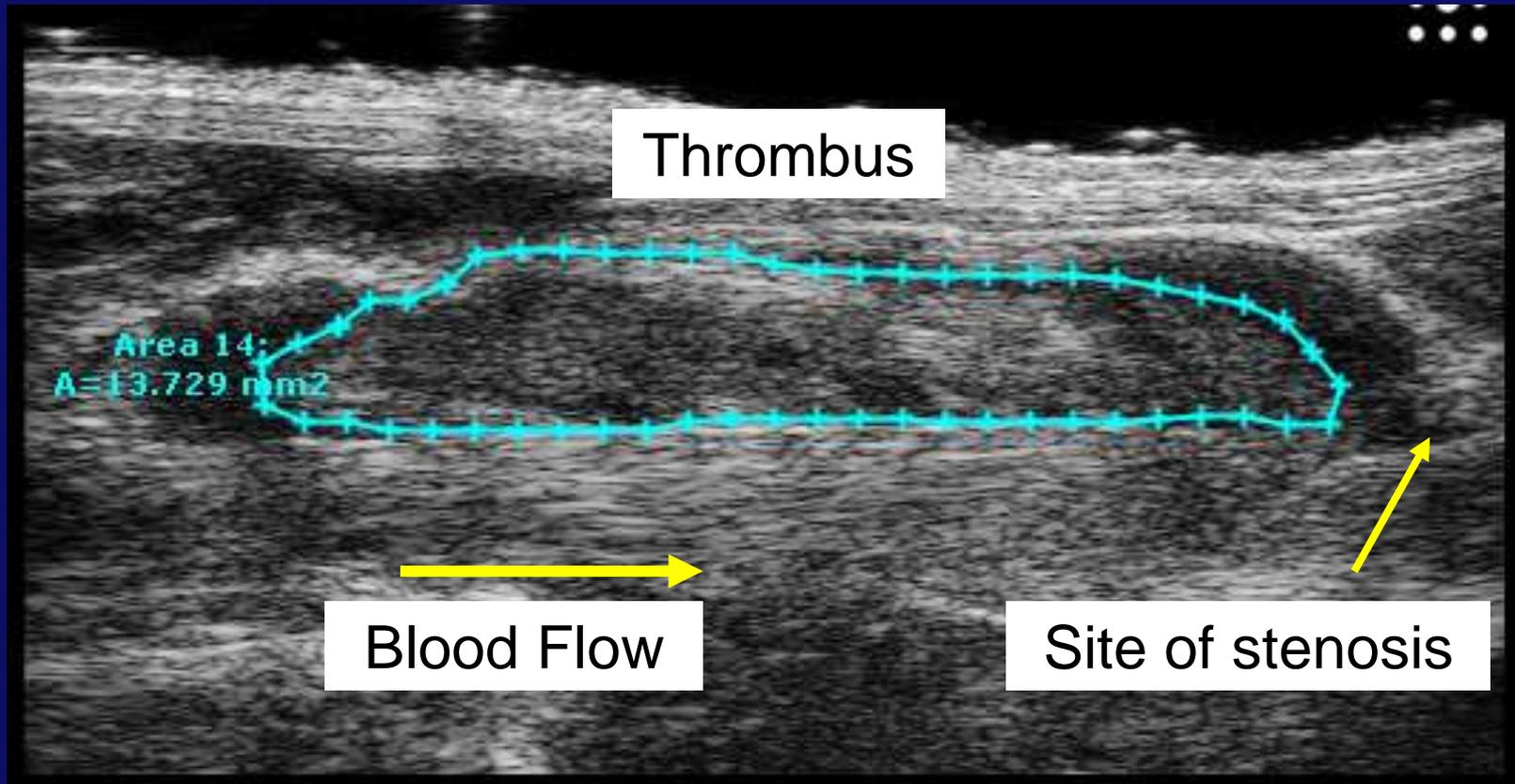
# Venous Thrombosis

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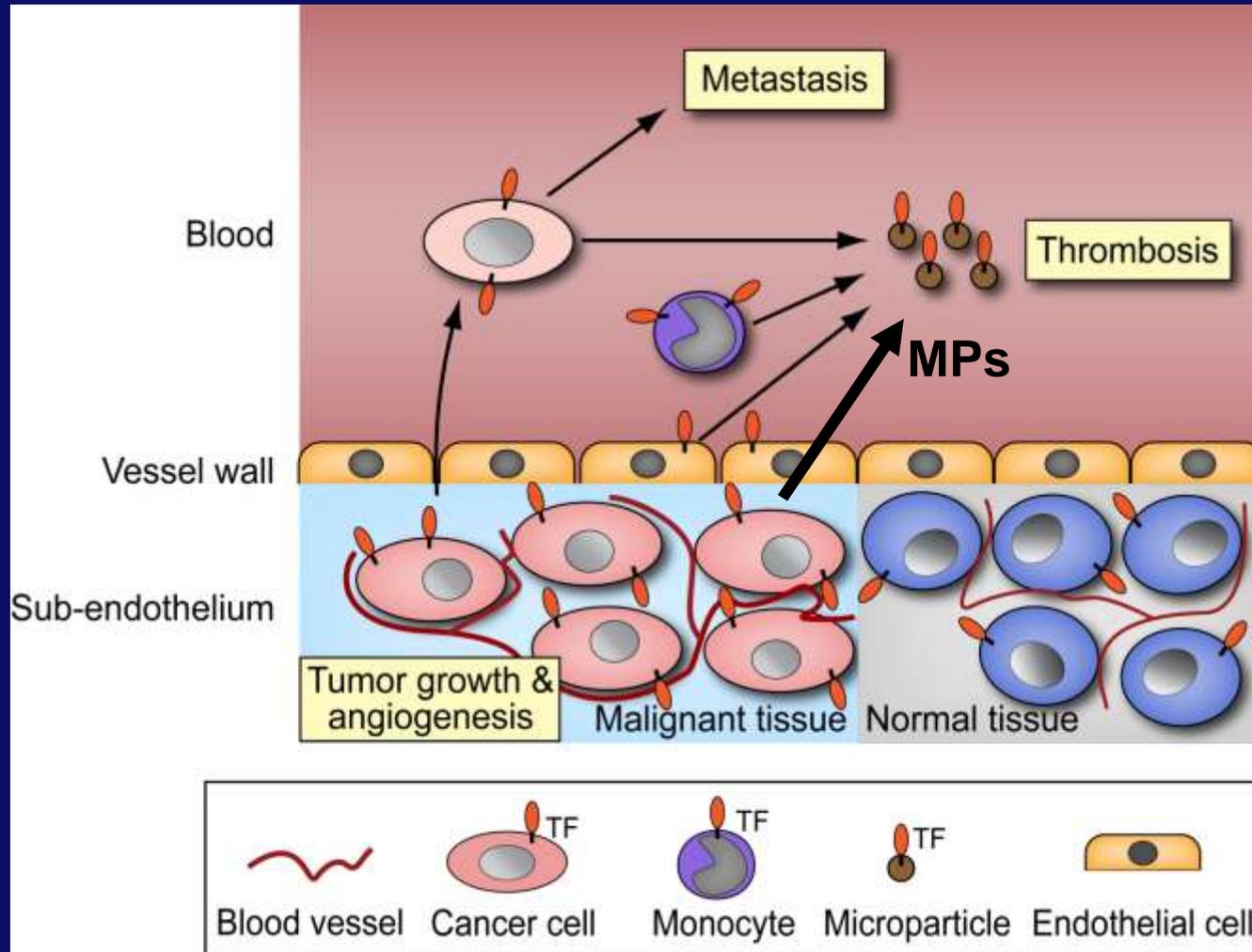
Slide provided by Dr. Wakefield

# Measurement of Thrombus Size by Ultrasound

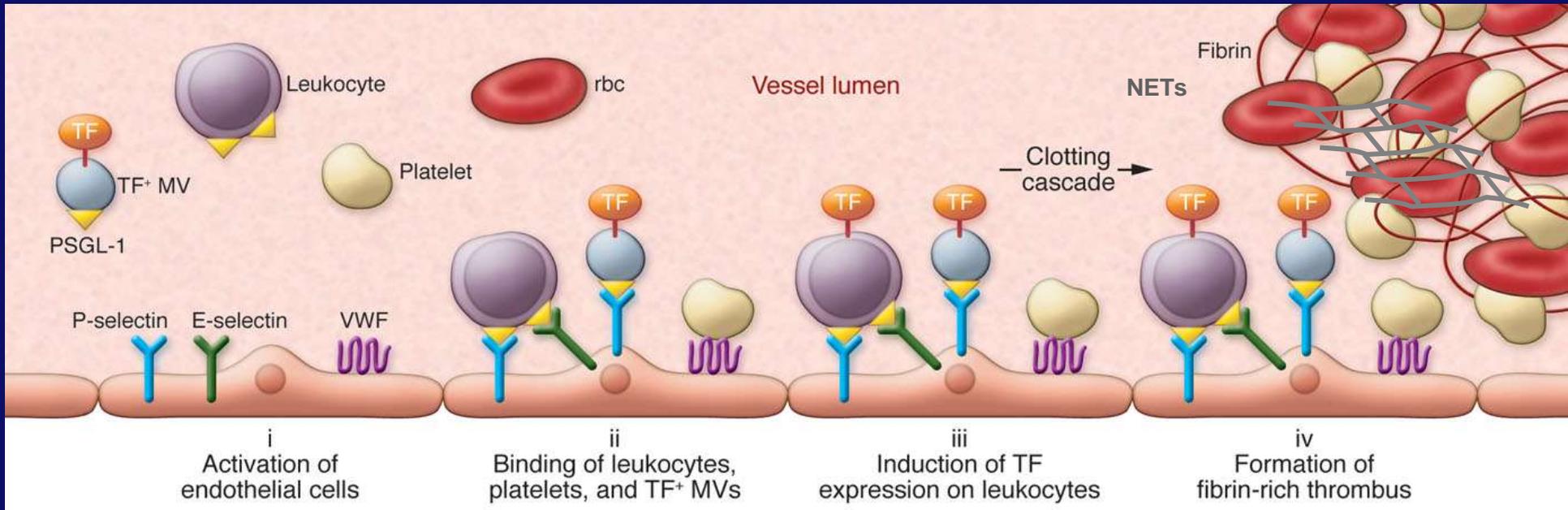


Gedding J unpublished data

# Role of Tissue Factor in Cancer



# Mechanisms of Venous Thrombosis



Mackman JCI 2012



*"That's all Folks!"*